

2019

## Trends in Collection of Microbiological Cultures Across Veterans Affairs Community Living Centers in the United States Over 8 Years

Haley J. Appaneal  
*University of Rhode Island*

Aisling R. Caffrey  
*University of Rhode Island, aisling\_caffrey@uri.edu*

Maria-Stephanie A. Hughes  
*University of Rhode Island*

Vrishali V. Lopes  
Follow this and additional works at: [https://digitalcommons.uri.edu/php\\_facpubs](https://digitalcommons.uri.edu/php_facpubs)  
*University of Rhode Island*

The University of Rhode Island Faculty have made this article openly available.  
Please let us know how Open Access to this research benefits you.

This is a pre-publication author manuscript of the final, published article.

See next page for additional authors  
Terms of Use

This article is made available under the terms and conditions applicable towards Open Access Policy Articles, as set forth in our [Terms of Use](#).

### Citation/Publisher Attribution

Appaneal, H. J., Caffrey, A. R., Hughes, M.-S. A., Lopes, V. L., Jump, R. L.P., LaPlante, K. L. & Dosa, M. M. (2019). Trends in Collection of Microbiological Cultures Across Veterans Affairs Community Living Centers in the United States Over 8 Years. *J. Am. Med. Assoc.*, 21(1), 115-120. <https://doi.org/10.1016/j.jamda.2019.07.003>

Available at: <https://doi.org/10.1016/j.jamda.2019.07.003>

This Article is brought to you for free and open access by the Pharmacy Practice at DigitalCommons@URI. It has been accepted for inclusion in Pharmacy Practice Faculty Publications by an authorized administrator of DigitalCommons@URI. For more information, please contact [digitalcommons@etal.uri.edu](mailto:digitalcommons@etal.uri.edu).

---

---

**Authors**

Haley J. Appaneal, Aisling R. Caffrey, Maria-Stephanie A. Hughes, Vrishali V. Lopes, Robin L.P. Jump, Kerry L. LaPlante, and David M. Dosa

1 **ABSTRACT WORDS 297**

2 **OBJECTIVES:** To describe and evaluate changes in the collection of microbiological cultures across Veterans  
3 Affairs (VA) Community Living Centers (CLCs) nationally.

4 **DESIGN:** Descriptive study.

5 **SETTING:** 146 VA CLCs.

6 **PARTICIPANTS:** We identified both positive and negative microbiological cultures collected during VA CLC  
7 admissions from January 2010 through December 2017.

8 **MEASURES:** We measured the average annual percent change (AAPC) in the rate of cultures collected per  
9 1,000 bed days and per admission, overall and stratified by culture type (i.e. urine, blood, skin and soft tissue,  
10 respiratory). AAPCs were also calculated for the proportion and rate of positive cultures collected, overall and  
11 stratified by culture type and organism (i.e. *Escherichia coli*, *Proteus mirabilis*, *Staphylococcus aureus*,  
12 *Enterococcus species [spp.]*, *Pseudomonas aeruginosa*, *Klebsiella spp.*, *Enterobacter spp.*, *Morganella*  
13 *morganii*, *Citrobacter spp.*, *Serratia marcescens*, *Streptococcus pneumoniae*). Joinpoint regression software  
14 was used to assess trends and estimate AAPCs and 95% confidence intervals (CI).

15 **RESULTS:** Over 8-years, 355,329 cultures were collected. The rate of cultures collected per 1,000 bed days of  
16 care decreased significantly by 6.0% per year (95% CI, -8.7 – -3.2%). The proportion of positive cultures  
17 decreased by 0.9% (95% CI, -1.4 – -0.4%). The most common culture types were urine (48.4%), followed by  
18 blood (27.7%). The rate of cultures collected per 1,000 bed days of care decreased per year by 6.3% for urine,  
19 5.0% for blood, 4.4% for skin and soft tissue, and 4.9% for respiratory. In 2010, *Staphylococcus aureus* was the  
20 most common organism identified and in all subsequent years *Escherichia coli* was the most common.

21 **CONCLUSION AND IMPLICATIONS:** We identified a significant reduction in the number of cultures collected  
22 over time among VA CLCs. Our findings may be explained by decreases in the collection of unnecessary cultures  
23 in VA CLCs nationally due to increased antibiotic stewardship efforts targeting unnecessary culturing and  
24 antibiotic treatment.

26 **BACKGROUND WORDS 2,345 REFERENCES 31 TABLES 5**

27 Diagnostic uncertainty contributes to antibiotic overuse in long-term care facilities (LTCFs).[1] The presentation  
28 of common infections, such as urinary tract infections, lower respiratory tract infections, and skin and soft tissue  
29 infections may be atypical among older patients.[1, 2] The signs and symptoms of infection in older adults may  
30 include acute confusion or subtle changes in physical state or functioning without classic signs of infection, such  
31 as fever and chills, which may jeopardize accurate diagnosis.[1, 2] Cognitive impairment often limits the ability  
32 of residents to articulate their symptoms, which may further complicate clinical assessments.[3-6] These issues  
33 may prompt LTCF clinicians to respond to any change in status with collection of microbiological cultures,  
34 particularly urine cultures. If cultures result in positive growth due to colonization and/or contamination versus  
35 true infection this can lead to inappropriate and unnecessary antibiotic therapy.[1]

36  
37 Despite the impact clinical microbiological cultures may ultimately have on antibiotic use, there have been no  
38 long-term studies describing the trends in the collection of cultures among LTCFs in the United States at the  
39 national level. Quantifying these trends may inform future targets for diagnostic stewardship intervention among  
40 LTCFs.[7, 8] Diagnostic stewardship has the same goal as antibiotic stewardship to improve antibiotic use, but  
41 intervenes at the level of the diagnostic test, to improve the way in which tests are ordered, performed, and  
42 reported.[8] Additionally, over the past decade, the importance of antibiotic stewardship and infection control  
43 have been increasingly recognized, and may have led to reductions in the collection of cultures.[9, 10] Thus, the  
44 objective of this study was to describe national trends in the rates of microbiological cultures collected at VA  
45 LTCFs (referred to as Community Living Centers or CLCs in the VA Healthcare System) nationally from 2010 to  
46 2017.

47  
48 **METHODS**

49 We conducted a longitudinal study to describe annual trends in the collection of microbiological cultures among  
50 VA CLCs nationally. The study was approved by the Institutional Review Board (IRB) and the Research and  
51 Development (R&D) Committee of the Providence Veterans Affairs Medical Center prior to initiation. National  
52 VA clinical and administrative data, including data hospital and long-term care admissions, outpatient visits,  
53 medication administrations and dispensings, and laboratory results, were used in this study. All microbiology

54 results among LTCF admissions entered in the electronic medical record were included in the study. From this  
55 data, we included all cultures collected during a stay at a VA CLC facility between January 1, 2010 and December  
56 31, 2017. Cultures from 146 VA CLCs and from all culture types were included. We included all cultures, even  
57 in the case of multiple cultures from the same patient, on the same day, of the same culture type. VA bed days  
58 were captured from Veterans Health Administration Support Services Center.

59  
60 For each calendar year, we calculated the number of cultures collected, including positive and negative cultures,  
61 as well as the rate of cultures collected per admission and per 1,000 bed days of care. We described the number  
62 of cultures collected per admission per year, which accounts for turnover but does not account for occupancy,  
63 and the number of cultures collected per bed days of care per year, which accounts for occupancy but does not  
64 account for turnover.[11, 12] Positive cultures were those in which any organism was recovered from the culture  
65 by the microbiological laboratory that tested the specimen and negative cultures were those in which no  
66 organisms were identified.

67  
68 Cultures were categorized by culture type and organism. Culture types were grouped into broad categories  
69 (urine, blood, skin and soft tissue, or respiratory) based on the body site where the specimen was collected (e.g.  
70 an expectorated sputum or endotracheal aspirate were grouped into respiratory and a skin swab or skin lesion  
71 aspirate were grouped into skin and soft tissue). “Other” was used to group cultures that did not fit into these  
72 broad categories. Positive cultures were grouped into the following organism categories: *Escherichia coli*,  
73 *Proteus mirabilis*, *Staphylococcus aureus*, *Enterococcus species (spp.)*, *Pseudomonas aeruginosa*, *Klebsiella*  
74 *spp.*, *Enterobacter spp.*, *Morganella morganii*, *Citrobacter spp.*, *Serratia marcescens*, *Streptococcus*  
75 *pneumoniae*, and other (positive for any another organism). Culture types and bacterial organisms were selected  
76 *a priori* due to their clinical importance and prevalence in the CLC setting.[13, 14] Polymicrobial cultures were  
77 counted as a single positive culture, however each organism identified was counted and categorized separately  
78 into organism types.

79  
80 Average annual percent changes (AAPC) were calculated for the rate of cultures collected per admission and  
81 per 1,000 bed days over the study period, overall and stratified by culture type and organism.[15, 16] AAPCs

were also calculated for proportion of positive cultures collected and the rate of positive cultures collected. The AAPC is calculated as a weighted average of the average percent change (APC) segments from a joinpoint model with the weights equal to the duration of each APC segment. The Joinpoint Regression Program version 4.6.0.0 (National Cancer Institute, Bethesda, MD, USA) was used to calculate AAPCs and 95% confidence intervals (CI). Significance was set at  $p < 0.05$ .

## RESULTS

Between 2010 and 2017, 355,329 cultures were collected from residents admitted to 146 VA CLCs. Positive microbial growth was reported in 42.0% of cultures ( $n=149,069$ ). The number of cultures collected per admission was 1.6 in 2010 and 0.9 in 2017 and decreased significantly by 8.3% per year (95% CI, -9.6 – -6.9%, **Table 1**). Similarly, the number of cultures collected per 1,000 bed days of care decreased significantly by 6.0% per year (95% CI, -8.7 – -3.2%, **Table 2**).

### *Trends in Cultures Collected by Culture Type*

The most common culture type collected (both positive and negative) during the study period was urine (48.4%,  $n=172,081$ ), followed by blood (27.7%,  $n= 98,422$ ), other (11.8%,  $n= 42,093$ ), skin and soft tissue (8.5%,  $n= 30,292$ ), and then respiratory (3.5%,  $n= 12,441$ ). The annual rates and trends in cultures collected by culture type are presented in **Table 1** (rates per admission) and **Table 2** (rates per 1,000 bed days). The rate of cultures collected per 1,000 bed days of care decreased by 6.3% per year for urine (95% CI, -8.2 – -4.4), 5.0% per year for blood (95% CI, -5.8 – -4.3), 4.4% per year for skin and soft tissue (95% CI, -6.6 – -2.3), and 4.9% per year for respiratory (95% CI, -6.9 – -2.7%, **Table 2**).

### *Trends in Positive Cultures Collected*

The rates of positive cultures collected (per admission and per 1,000 bed days) decreased over the study period, as did the proportion of positive cultures (AAPC = -0.9%, 95% CI, -1.4 – -0.4%). The annual distribution and trends in rates (per 1,000 bed days of care) of positive cultures are presented in **Table 3**. Similar results were observed in rates per admission but are not presented in the Tables.

109 The proportion of positive cultures collected remained stable for urine cultures at about 53% (AAPC = 0.2%,  
110 95% CI, -0.1 – 0.5%) and skin and soft tissue cultures at about 62% (AAPC = 0.1%, 95% CI, -1.0 – 1.2%). The  
111 proportion of positive cultures decreased over the study period for blood cultures from about 14% to 12% (AAPC  
112 = -1.7%, 95% CI, -2.9 – -0.6%) and respiratory cultures from about 54% to 43% (AAPC = -2.8%, 95% CI, -4.0 –  
113 -1.7%).

#### 114 115 *Trends in Cultures Collected by Species of Organism*

116 In 2010, *S. aureus* was the most common organism identified. In subsequent years, *E. coli* was the most common  
117 organism identified. The annual distribution and trends in rates (per 1,000 bed days of care) of positive cultures  
118 by species of organism are presented in **Table 4**. Similar results were observed in rates per admission but are  
119 not presented in the Tables. The rate of positive cultures decreased for most organisms (i.e. *S. aureus*,  
120 *Enterococcus spp.*, *E. coli*, *P. mirabilis*, *P. aeruginosa*, *Enterobacter spp.*, *Citrobacter spp.*, *M. morgani*, and  
121 *Serratia marcescens*) except *S. pneumoniae* and *Klebsiella spp.* which remained stable. The trends in rates (per  
122 1,000 bed days of care) of positive cultures by species of organism and culture type are presented in **Table 5**.  
123 The rate of positive cultures growing *S. aureus* decreased for all culture types (urine, blood, skin and soft tissue,  
124 or respiratory).

## 125 126 **DISCUSSION**

127 We identified a significant reduction in the rate of microbiological cultures collected over an 8 year period, which  
128 may be related to increased infection control and antibiotic stewardship throughout the VA healthcare  
129 system.[17-19] Increased antibiotic stewardship efforts may lead to reduced culture collection rates through  
130 reduction of inappropriate or unnecessary microbiological cultures.

131  
132 The overall rate of cultures collected over time was largely driven by a reduction in urine cultures, which may be  
133 related to an increased awareness of overdiagnosis and testing for UTIs over the study period.[4, 20] The  
134 appropriate diagnosis and treatment of UTIs has become one of the most important antibiotic stewardship targets  
135 among LTCFs, due to the high prevalence of asymptomatic bacteriuria among residents.[9] Previous smaller  
136 studies have demonstrated that UTI-focused antibiotic stewardship interventions are associated with reductions

137 in the collection of urine cultures.[4, 20, 21] For example, a single center study in a VA CLC demonstrated a  
138 reduction in urine culture collection rate (from 3.7 to 1.5 per 1,000 resident days) after implementation of a  
139 pathway to limit urine testing without urinary symptoms.[20] Our research shows this trend on a national scale  
140 and suggests that the trend towards reduced urine testing has been occurring across the entire VA CLC system.

141  
142 In addition to a reduction in urine cultures, we also observed significant reductions in blood, skin and soft tissue,  
143 and respiratory cultures collected from VA CLC residents, which suggests diagnostic stewardship may extend  
144 beyond urine testing. Similar to urine cultures, limiting over collection of skin and soft tissue cultures is an  
145 important stewardship target in LTCFs.[22] Wounds and pressure ulcers, which are common in older adults, are  
146 colonized by bacteria.[22] As such, collection of microbiological samples from skin or wounds in the absence of  
147 signs or symptoms of infection could lead to inappropriate antibiotic use.[3, 13] For respiratory and blood cultures,  
148 the role of diagnostic stewardship on culture reduction rates we observed in VA CLCs may be more related to  
149 efforts to reduce contamination rather than efforts to reduce unnecessary cultures. The role of respiratory  
150 cultures in LTCFs are generally limited, as residents are often not be able to produce expectorated sputum and  
151 even when respiratory cultures are available they are often contaminated. [23] Similarly, the role of blood cultures  
152 in LTCFs is limited, as they generally have a low yield in LTCF residents and most residents with suspected  
153 bacteremia are transferred to acute-care facilities.[23] Nonetheless, reduction of blood culture contamination is  
154 an important tenant of diagnostic stewardship and overall quality improvement in many healthcare settings.[24-  
155 26] Interestingly, when stratified by culture type, we did find the proportion of positive cultures collected remained  
156 stable for urine cultures and skin and soft tissue cultures, but the proportion of positive cultures decreased for  
157 blood cultures and respiratory cultures.

158  
159 An important finding from this research is the overall reduction in the rate of positive cultures collected for  
160 particularly virulent organisms, such as *S. aureus*, *Enterococcus spp.*, *E. coli*, and *P. aeruginosa*, among the VA  
161 CLC population. While we did not assess antibiotic resistance, previous work has demonstrated that the burden  
162 of infections due to methicillin-resistant *S aureus* has significantly declined in the VA since 2007 when a  
163 prevention initiative was implemented among all VA medical centers.[17, 18, 27, 28] This initiative may have  
164 also led to reductions in other non-*S. aureus* infections as well, through expanded infection control programs



165 and resources.[29] However, similar trends have been observed nationally outside the VA, which may suggest  
166 shifts in strain epidemiology and may also contribute to our findings.[30] Further work in this area will need to  
167 occur to identify why the trends we observed occurred.

168  
169 There are limitations inherent in our work. Our study included all cultures that were collected and did not assess  
170 the clinical significance of the cultures. Any culture in which any organism or any bacteria were recovered were  
171 defined as positive and as such, we cannot distinguish what proportion of positive cultures represent true  
172 infection versus colonization and/or contamination. Additionally, in our assessment of culture by organism there  
173 may be an overestimation of bacteria in which multiple positive cultures are obtained from the same patient with  
174 the same results. Moreover, the indications for the cultures are unknown. For example, the collection of a urine  
175 culture may be indicated for a resident with sepsis, whether or not a positive result represents a UTI or  
176 asymptomatic bacteriuria. The generalizability of results among VA CLCs to community non-VA LTCFs may be  
177 limited. The residents of VA CLCs have more complex medical needs and the staffing levels in are higher than  
178 in non-VA LTCFs.[31] Finally, while many facilities ensure that cultures results obtained from outside the VA  
179 system are captured within the electronic medical record, it is impossible to ascertain if all cultures collected at  
180 outside laboratories are included in this analysis. Finally, our categorization of cultures by source and organism  
181 are based on the ordering, interpretation, and reporting of the providers and microbiology laboratory that ordered  
182 and handled the clinical culture. Additionally, the specificity of the culture collection site varies, and we cannot  
183 always identify the specific body sites where cultures were collected (for example sputum versus endotracheal  
184 among respiratory cultures, or skin swab versus lesion aspirate among skin cultures). As such, we used  
185 used broad culture type categories (e.g. urine, blood, skin and soft tissue, or respiratory).

186  
187 We did not assess whether changes in culture collection corresponded with changes in antibiotic use, resource  
188 utilization and costs, or resident outcomes, such as hospitalizations, which should be explored further.

## 189 190 **CONCLUSIONS AND IMPLICATIONS**

191 Our data revealed a significant reduction in the number of cultures collected among VA CLCs nationally over an  
192 8-year period, with a large reduction in urine cultures. This reduction in cultures likely reflects a reduction in

193 collection of unnecessary cultures in VA CLCs nationally and may be driven by increased awareness for over-  
194 testing and over-treatment of presumed urinary tract infection. Moreover, this reduction may have had important  
195 clinical and economic impact through reduction of additional unnecessary testing and antibiotic use, reduced  
196 resource utilization and costs, and improved overall resident care and outcomes.

197

## References

1. Beckett CL, Harbarth S, Huttner B. Special considerations of antibiotic prescription in the geriatric population. *Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases* **2015**; 21(1): 3-9.
2. Gavazzi G, Krause KH. Ageing and infection. *Lancet Infect Dis* **2002**; 2(11): 659-66.
3. Dyar OJ, Pagani L, Pulcini C. Strategies and challenges of antimicrobial stewardship in long-term care facilities. *Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases* **2015**; 21(1): 10-9.
4. Crnich CJ, Jump R, Trautner B, Sloane PD, Mody L. Optimizing Antibiotic Stewardship in Nursing Homes: A Narrative Review and Recommendations for Improvement. *Drugs Aging* **2015**; 32(9): 699-716.
5. Nicolle LE, Long-Term-Care-Committee S. Urinary tract infections in long-term-care facilities. *Infection control and hospital epidemiology : the official journal of the Society of Hospital Epidemiologists of America* **2001**; 22(3): 167-75.
6. Hooton TM, Bradley SF, Cardenas DD, et al. Diagnosis, prevention, and treatment of catheter-associated urinary tract infection in adults: 2009 International Clinical Practice Guidelines from the Infectious Diseases Society of America. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America* **2010**; 50(5): 625-63.
7. Koch C, Roberts K, Petrucci C, Morgan DJ. The Frequency of Unnecessary Testing in Hospitalized Patients. *Am J Med* **2018**; 131(5): 500-3.
8. Morgan DJ, Malani P, Diekema DJ. Diagnostic Stewardship-Leveraging the Laboratory to Improve Antimicrobial Use. *JAMA* **2017**; 318(7): 607-8.
9. Centers for Disease Control and Prevention (CDC). The Core Elements of Antibiotic Stewardship for Nursing Homes. **Atlanta, GA: US Department of Health and Human Services, CDC; 2015.** Available at: <http://www.cdc.gov/longtermcare/index.html>. Accessed 20 Sept 2015.
10. Dellit TH, Owens RC, McGowan JE, Jr., et al. Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America guidelines for developing an institutional program to enhance antimicrobial stewardship. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America* **2007**; 44(2): 159-77.
11. Schechner V, Temkin E, Harbarth S, Carmeli Y, Schwaber MJ. Epidemiological interpretation of studies examining the effect of antibiotic usage on resistance. *Clin Microbiol Rev* **2013**; 26(2): 289-307.
12. Bertollo LG, Lutkemeyer DS, Levin AS. Are antimicrobial stewardship programs effective strategies for preventing antibiotic resistance? A systematic review. *American journal of infection control* **2018**; 46(7): 824-36.
13. Montoya A, Mody L. Common infections in nursing homes: a review of current issues and challenges. *Ageing health* **2011**; 7(6): 889-99.
14. Nicolle LE. Infection control in long-term care facilities. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America* **2000**; 31(3): 752-6.
15. Clegg LX, Hankey BF, Tiwari R, Feuer EJ, Edwards BK. Estimating average annual per cent change in trend analysis. *Stat Med* **2009**; 28(29): 3670-82.

- 239 16. Kim HJ, Fay MP, Feuer EJ, Midthune DN. Permutation tests for joinpoint regression with applications to  
240 cancer rates. *Stat Med* **2000**; 19(3): 335-51.
- 241 17. Evans ME, Kralovic SM, Simbartl LA, Jain R, Roselle GA. Eight years of decreased methicillin-resistant  
242 *Staphylococcus aureus* health care-associated infections associated with a Veterans Affairs prevention  
243 initiative. *American journal of infection control* **2017**; 45(1): 13-6.
- 244 18. Evans ME, Kralovic SM, Simbartl LA, et al. Nationwide reduction of health care-associated methicillin-  
245 resistant *Staphylococcus aureus* infections in Veterans Affairs long-term care facilities. *American*  
246 *journal of infection control* **2014**; 42(1): 60-2.
- 247 19. Chou AF, Graber CJ, Jones M, et al. Characteristics of Antimicrobial Stewardship Programs at  
248 Veterans Affairs Hospitals: Results of a Nationwide Survey. *Infect Control Hosp Epidemiol* **2016**; 37(6):  
249 647-54.
- 250 20. Zabarsky TF, Sethi AK, Donskey CJ. Sustained reduction in inappropriate treatment of asymptomatic  
251 bacteriuria in a long-term care facility through an educational intervention. *American journal of infection*  
252 *control* **2008**; 36(7): 476-80.
- 253 21. Trautner BW, Grigoryan L, Petersen NJ, et al. Effectiveness of an Antimicrobial Stewardship Approach  
254 for Urinary Catheter-Associated Asymptomatic Bacteriuria. *JAMA Intern Med* **2015**; 175(7): 1120-7.
- 255 22. Nicolle LE, Bentley DW, Garibaldi R, Neuhaus EG, Smith PW. Antimicrobial use in long-term-care  
256 facilities. SHEA Long-Term-Care Committee. *Infection control and hospital epidemiology : the official*  
257 *journal of the Society of Hospital Epidemiologists of America* **2000**; 21(8): 537-45.
- 258 23. High KP, Bradley SF, Gravenstein S, et al. Clinical practice guideline for the evaluation of fever and  
259 infection in older adult residents of long-term care facilities: 2008 update by the Infectious Diseases  
260 Society of America. *Clinical infectious diseases : an official publication of the Infectious Diseases*  
261 *Society of America* **2009**; 48(2): 149-71.
- 262 24. Madden GR, Weinstein RA, Sifri CD. Diagnostic Stewardship for Healthcare-Associated Infections:  
263 Opportunities and Challenges to Safely Reduce Test Use. *Infection control and hospital epidemiology :*  
264 *the official journal of the Society of Hospital Epidemiologists of America* **2018**; 39(2): 214-8.
- 265 25. Youssef D, Shams W, Bailey B, O'Neil TJ, Al-Abbadi MA. Effective strategy for decreasing blood  
266 culture contamination rates: the experience of a Veterans Affairs Medical Centre. *J Hosp Infect* **2012**;  
267 81(4): 288-91.
- 268 26. Robertson P, Russell A, Inverarity DJ. The effect of a quality improvement programme reducing blood  
269 culture contamination on the detection of bloodstream infection in an emergency department. *J Infect*  
270 *Prev* **2015**; 16(2): 82-7.
- 271 27. Jain R, Kralovic SM, Evans ME, et al. Veterans Affairs initiative to prevent methicillin-resistant  
272 *Staphylococcus aureus* infections. *The New England journal of medicine* **2011**; 364(15): 1419-30.
- 273 28. Evans ME, Kralovic SM, Simbartl LA, et al. Prevention of methicillin-resistant *Staphylococcus aureus*  
274 infections in spinal cord injury units. *American journal of infection control* **2013**; 41(5): 422-6.
- 275 29. Goto M, O'Shea AMJ, Livorsi DJ, et al. The Effect of a Nationwide Infection Control Program Expansion  
276 on Hospital-Onset Gram-Negative Rod Bacteremia in 130 Veterans Health Administration Medical  
277 Centers: An Interrupted Time-Series Analysis. *Clin Infect Dis* **2016**; 63(5): 642-50.
- 278 30. Kourtis AP, Hatfield K, Baggs J, et al. Vital Signs: Epidemiology and Recent Trends in Methicillin-  
279 Resistant and in Methicillin-Susceptible *Staphylococcus aureus* Bloodstream Infections - United States.  
280 *MMWR Morbidity and mortality weekly report* **2019**; 68(9): 214-9.

281 31. Thomas KS, Cote D, Makineni R, et al. Change in VA Community Living Centers 2004-2011: Shifting  
282 Long-Term Care to the Community. *J Aging Soc Policy* **2018**; 30(2): 93-108.  
283

284

**Table 1. Annual rates (per admission) and trends in cultures collected at VA Community Living Centers nationally from 2010-2017**

<b>Year</b>	<b>Overall cultures</b>	<b>Urine cultures</b>	<b>Blood cultures</b>	<b>Skin cultures</b>	<b>Respiratory cultures</b>
2010	1.6	0.692	0.389	0.125	0.052
2011	1.4	0.648	0.373	0.111	0.049
2012	1.2	0.613	0.342	0.105	0.040
2013	1.1	0.577	0.315	0.097	0.040
2014	1.1	0.538	0.304	0.090	0.039
2015	1.0	0.505	0.300	0.094	0.036
2016	0.9	0.474	0.276	0.086	0.034
2017	0.9	0.429	0.262	0.080	0.034
<b>AAPC (95% CI)</b>	-8.3% (-9.6 – -6.9)*	-6.4 (-6.8 – -6.0)*	-5.4 (-6.2 – -4.7)*	-5.6 (-6.9 – -4.2)*	-5.9 (-7.8 – -4.0)*

The Joinpoint Regression Program was used to calculate average annual percent change (AAPC) and 95% confidence intervals (95% CI). AAPC is calculated as a weighted average of the average percent change (APC) segments from a joinpoint model with the weights equal to the duration of each APC segment.

\*Indicates significant trend at p<0.05.

CLC= Community Living Center

**Table 2. Annual rates (per 1,000 bed days) and trends in cultures collected at VA Community Living Centers nationally from 2011-2017**

Year	Overall cultures	Urine cultures	Blood cultures	Skin cultures	Respiratory cultures
2011	14.3	6.8	3.9	1.2	0.5
2012	13.4	6.6	3.7	1.1	0.4
2013	12.8	6.5	3.6	1.1	0.5
2014	12.0	6.1	3.4	1.0	0.4
2015	11.7	5.8	3.5	1.1	0.4
2016	10.5	5.3	3.1	1.0	0.4
2017	9.3	4.6	2.8	0.9	0.4
<b>AAPC</b>	-6.0%	-6.3%	-5.0%	-4.4%	-4.9%
<b>(95% CI)</b>	(-8.7 – -3.2)*	(-8.2 – -4.4)*	(-5.8 – -4.3)*	(-6.6 – -2.3)*	(-6.9 – -2.7)*

The Joinpoint Regression Program was used to calculate average annual percent change (AAPC) and 95% confidence intervals (95% CI). AAPC is calculated as a weighted average of the average percent change (APC) segments from a joinpoint model with the weights equal to the duration of each APC segment.

\*Indicates significant trend at p<0.05.

CLC= Community Living Center

**Table 3. Annual distribution and trends in positive cultures collected at VA Community Living Centers nationally from 2011-2017**

<b>Year</b>	<b>Proportion of positive cultures</b>	<b>Positive cultures per 1,000 CLC bed days</b>
2011	43.3%	6.2
2012	42.3%	5.7
2013	42.8%	5.5
2014	41.8%	5.0
2015	41.2%	4.8
2016	40.6%	4.3
2017	40.1%	3.8
<b>AAPC (95% CI)</b>	<b>-0.9 ( -1.4 -- 0.4)*<sup>a</sup></b>	<b>-7.0 (-9.7 -- -4.2)*<sup>b</sup></b>

The Joinpoint Regression Program was used to calculate average annual percent change (AAPC) and 95% confidence intervals (95% CI). AAPC is calculated as a weighted average of the average percent change (APC) segments from a joinpoint model with the weights equal to the duration of each APC segment.

\* Indicates significant trend at  $p < 0.05$ .

CLC= Community Living Center

<sup>a</sup>AAPC presented for change in proportion of positive cultures collected from 2010 to 2017. The proportion of positive cultures in 2010 was 42.2%.

<sup>b</sup>AAPC presented for change in rate of positive cultures per 1,000 bed days of care. Similar results were observed for rate per admission (data not presented in table).



**Table 4. Annual rates (per 1,000 bed days) and trends in positive cultures by organism collected at VA Community Living Centers nationally from 2011-2017**

Year	<i>Staphylococcus aureus</i>	<i>Enterococcus spp.</i>	<i>Streptococcus pneumoniae</i>	<i>Escherichia coli</i>	<i>Proteus mirabilis</i>	<i>Pseudomonas aeruginosa</i>	<i>Klebsiella spp.</i>	<i>Enterobacter spp.</i>	<i>Morganella morganii</i>	<i>Citrobacter spp.</i>	<i>Serratia marcescens</i>
2011	0.989	0.743	0.011	0.997	0.908	0.564	0.535	0.139	0.114	0.078	0.050
2012	0.841	0.750	0.014	0.930	0.819	0.535	0.493	0.143	0.117	0.078	0.047
2013	0.802	0.752	0.011	0.947	0.826	0.534	0.543	0.140	0.114	0.081	0.043
2014	0.738	0.625	0.007	0.881	0.759	0.451	0.511	0.140	0.099	0.074	0.042
2015	0.651	0.619	0.008	0.856	0.713	0.456	0.511	0.110	0.093	0.075	0.044
2016	0.593	0.533	0.011	0.771	0.684	0.462	0.479	0.114	0.074	0.064	0.040
2017	0.520	0.484	0.010	0.758	0.596	0.379	0.395	0.120	0.076	0.065	0.038
<b>AAPC</b>	-9.6	-7.4	-3.8	-4.5	-6.1	-5.7	-3.6	-4.0	-8.0	-3.6	-3.9
<b>(95% CI)</b>	(-10.8 – -8.5)*	(-10.2 – -4.6)*	(-14.2 – -7.8)	(-5.8 – -3.2)*	(-7.6 – -4.6)*	(-8.3 – -3.0)*	(-7.2 – -0.1)	(-7.5 – -0.3)*	(-11.2 – -4.6)*	(-6.0 – -1.1)*	(-5.7 – -2.2)*

The Joinpoint Regression Program was used to calculate average annual percent change (AAPC) and 95% confidence intervals (95% CI). AAPC is calculated as a weighted average of the average percent change (APC) segments from a joinpoint model with the weights equal to the duration of each APC segment. \*Indicates significant trend at p<0.05.

AAPC presented for change in rates per 1,000 bed days of care. Similar results were observed for rates per admission (data not presented in table).

CLC= Community Living Center; spp.= species

**Table 5. Trends in rates (per 1,000 bed days) of positive cultures by organism and culture type collected at VA Community Living Centers nationally from 2011-2017**

	<i>Staphylococcus aureus</i>	<i>Enterococcus spp.</i>	<i>Escherichia coli</i>	<i>Proteus mirabilis</i>	<i>Pseudomonas aeruginosa</i>	<i>Klebsiella spp.</i>	<i>Enterobacter spp.</i>	<i>Morganella morganii</i>	<i>Citrobacter spp.</i>	<i>Serratia marcescens</i>
<b>Urine</b>	-8.0	-7.5	-4.9	-6.8	-5.7	-3.7	-4.5	-8.3	-4.3	-3.7
<b>AAPC</b>	(-9.8 – -6.1)*	(-11.3 – -3.5)*	(-6.3 – -3.5)*	(-8.9 – -4.8)*	(-8.1 – -3.3)*	(-7.1 – -0.1)*	(-10.0 – -1.3)	(-11.6 – -4.9)*	(-6.8 – -1.7)*	(-10.0 – -3.1)
<b>Blood</b>	-6.0	-8.2	-2.7	-2.3	-10.9	-0.4	-6.6	-1.2	-1.2	-15.7
<b>AAPC</b>	(-11.5 – -0.2)*	(-14.7 – -1.3)*	(-8.2 – -3.1)	(-11.1 – -7.4)	(-23.4 – -3.6)	(-11.7 – -12.3)	(-18.7 – -7.2)	(-33.4 – -46.4)	(-48.6 – -90.0)	(-38.5 – -15.6)
<b>Skin</b>	-6.7	-3.7	-0.6	-2.4	-2.6	-1.6	2.8	-10.2	3.8	-0.9
<b>AAPC</b>	(-8.6 – -4.7)*	(-6.2 – -1.1)*	(-3.1 – -1.9)	(-4.6 – -0.2)*	(-6.9 – -1.9)	(-8.8 – -6.1)	(-3.2 – 9.2)	(-15.8 – -4.2)*	(-4.5 – 12.8)	(-9.0 – 7.9)
<b>Respiratory</b>	-9.8	-11.0	-11.3	-16.4	-10.4	-10.2	-17.6	-10.1	-17.9	-8.0
<b>AAPC</b>	(-12.7 – -6.7)*	(-41.4 – -35.3)	(-18.7 – -3.2)*	(-24.3 – -7.7)*	(-14.8 – -5.7)*	(-12.8 – -7.5)*	(-21.8 – -13.1)*	(-38.4 – -31.2)	(-32.9 – 0.4)	(-19.7 – 5.4)

The Joinpoint Regression Program was used to calculate average annual percent change (AAPC) and 95% confidence intervals (95% CI). AAPC is calculated as a weighted average of the average percent change (APC) segments from a joinpoint model with the weights equal to the duration of each APC segment. \*Indicates significant trend at  $p < 0.05$ .

AAPC presented for change in rates per 1,000 bed days of care. Similar results were observed for rates per admission (data not presented in table).