5-2015

Propagation Methods for Growing Spartina alterniflora for Salt Marsh Restoration

Samantha Walker
University of Rhode Island, samantha.walker@uconn.edu

Follow this and additional works at: https://digitalcommons.uri.edu/srhonorsprog

Part of the Plant Biology Commons

Recommended Citation

This Article is brought to you for free and open access by the Honors Program at the University of Rhode Island at DigitalCommons@URI. It has been accepted for inclusion in Senior Honors Projects by an authorized administrator of DigitalCommons@URI. For more information, please contact digitalcommons-group@uri.edu.
Propagation methods for growing *Spartina alterniflora* for salt marsh restoration

**Samantha Walker¹, Hope Leeson², Peter August³**

1. University of Rhode Island Coastal Fellow  
2. Rhody Native/ Rhode Island Natural History Survey  
3. University of Rhode Island Department of Natural Resources

---

**Introduction and Background**

**Salt Marsh Restoration and *Spartina alterniflora***

**Problem:** Coastal salt marshes are one of the most productive ecosystems on earth and provide countless ecosystem services including shoreline protection from storms and flooding, nutrient removal, habitat for fish, birds and other wildlife and provide some of the most beautiful areas for hunting, fishing and recreational activities. However, salt marshes are disappearing along the east coast of the United States due to human development and sea level rise.

**Solution:** In order to protect salt marshes and restore the large portion of them that have been damaged, it is important to focus on the vegetation that help salt marshes function. *S. alterniflora*, smooth cordgrass, is a critical component of the salt marsh vegetation community. *S. alterniflora* is a dominant species that helps to stabilize the ecosystem, retain a seedbank of other species, uptake nutrients, and provide important habitat for wildlife.

**Purpose of Project:** In order to grow *S. alterniflora* for restoration purposes, the species must be grown from seed to provide genetic diversity and high survival rates when planted. However, low germination rates have limited the use of this species for restoration using seeds. While working with, Rhody Native, I tested different methods to achieve high germination rates for *S. alterniflora* including comparing soil mixes, seed colors, and root development stages. This process is essential to find a propagation method that will work effectively to obtain high germination rates for this essential species to then use for restoration in local salt marshes.

---

**Results**

**Seed Color Variation**

- **Light seeds**
  - Empty seeds, lack an embryo
  - Do not germinate
  - Most seeds sorted were light seeds
- **Medium seeds**
  - Lower germination rate than dark seeds
  - 25% germination rate
  - Smallest group of seeds
- **Dark seeds**
  - Contain fully developed embryo, therefore high chance of germination
  - 85-90% germination rate

**Seed Germination Media**

- **Sand**
  - Both seed types germinated at the same rate in each soil mix
  - Leaf color turned yellow over time, due to lack of nutrients in growing media
- **1:1 sand to metromix 510**
  - Seedlings retained dark green leaf color
  - Seedlings had better root development

**Seedling Propagation**

- **Group of seedlings transplanted at 15 days**
  - Less root development
- **Group of seedlings transplanted at 30 days**
  - More root development
  - Higher survival rate and faster growth

**Irrigation System**

Half of the pots were out into an ebb and flow system while half were over head watered. The data from the different irrigation systems are in the process of being recorded.

---

**Methods/ Protocol**

First, the *S. alterniflora* seeds were collected at Succotash marsh, RI, 10/20/14 and put into cold stratification for 3 months.

Second, the seeds were sorted into dark, medium and light colors and sowed into sand or 1:1 metromix 510 to sand.

Third, the seedlings were transplanted to larger pots with 1:1 sand and peat, seedlings with varying root development.

Fourth, the pots were transferred into the ebb and flow system and some were overhead watered. Eventually seedlings will be planted for restoration.

---

**Citations/ Acknowledgements**


I want to thank Hope Leeson for agreeing to let me complete this project with her and taking the time to explain every detail about the plants to me and helping me with every step. I have learned more from you than I could have ever hoped and thank you for helping to develop my passions for native plants. I want to thank Professor Peter August for helping to introduce me to Hope Leeson and encouraging me every step of the way. I want to thank Dr. Laura Meyerson, Melissa Burger and Sara Wigginton for helping to edit my poster and encouraging me as well every step of the way.