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Francois A. Menard
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A PHYSICO-CHEMICAL STUDY
OF THE COMPLEXATION OF CYCLODEXTRINS
WITH PHARMACEUTICAL SUBSTANCES

BY
FRANCOIS A. MENARD

A DISSERTATION SUBMITTED IN PARTIAL FULFILLMENT OF THE
REQUIREMENTS FOR THE DEGREE OF
DOCTOR OF PHILOSOPHY
IN
PHARMACEUTICAL SCIENCES

UNIVERSITY OF RHODE ISLAND

1988

DOCTOR OF PHILOSOPHY DISSERTATION

OF

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UNIVERSITY OF RHODE ISLAND

1988

ABSTRACT

The complexation of drugs with alpha, beta, gamma and hydroxyethyl beta cyclodextrins was investigated using the solubility method described by Higuchi and Connors. Beta cyclodextrin was shown to exhibit the most extensive complexation. The solubility isotherms of complexation were studied at different temperatures and at various pH conditions. The effect of ionization of drugs on complexation was thus evaluated for four drugs viz phenytoin, diazepam, ibuprofen and hydrochlorothiazide. In the case of phenytoin and diazepam, both the non-ionized and the ionized species were shown to form a complex with cyclodextrins. The slope of complexation for ibuprofen and hydrochlorothiazide was found to be pH-independent. The thermodynamic parameters of complexation were determined using the temperature dependence method. In order to characterize more specifically the parameters of complexation, a novel computerized method was then developed. A non-linear regression software was used to evaluate the values of the complexation parameters as well as their standard deviation and 95% confidence limit. The complexation of drugs with hydroxyethyl beta cyclodextrin showed that the substitution of the natural cyclodextrin affected the solubility of the complexing agent without altering its solubilizing power. A rule of prediction was

then proposed, allowing the calculation of potential drug concentration upon complexation with the cyclodextrin derivative. The ratio of solubility of the two kinds of cyclodextrins was shown to correlate very well with the ratio of the concentration of drug complexed. The rule might be applied to all drugs at various pH values. The potential use of beta cyclodextrin as an additive in tableting was also evaluated. The mechanism of dissolution rate enhancement was found to be related not only to the inclusion complex formation but also to mechanical changes during the mixing process.

ACKNOWLEDGEMENTS

I would like to express my thanks to my Major Professor, Dr. Christopher T. Rhodes, for his constant support, his scientific guidance and his great professional approach. I believe that it has been a fruitful and enjoyable teamwork for both of us.

I am very grateful to research committee members, Dr. P. Brown and Dr. M. Dedhiya. Dr. Brown has been very helpful and a great source of enthusiasm. Dr. Dedhiya has been a great advisor and a friend since our first association in 1984.

During my four years in the Department of Pharmaceutics, I have seen people coming and going. I would especially like to thank Dr. B. Birmingham and Dr. M. Gardner, who had been assisting me in the computer aspects of the research project, and Dr. A. Paruta, who has been of great assistance. Also I thank Dr. G. Osborne, who was always there to give me the right advise, and Dr. J. Lausier, who has been of great help and support.

I would like to express my thanks to all the Faculty members of the University, with whom I took courses; they will be forever a part of my life and my future.

I also want to thank my fellow graduate students, those who have already left, those who are leaving soon, and those who are going to carry the torch for the next several years.

I would like to express my gratitude to Miles Pharmaceuticals, the University of Rhode Island, Dr. Rhodes and the Department of Pharmaceutics for the financial support throughout my graduate studies.

I also want to thank Dr. D.C. Monkhouse, who gave me the advise to pursue graduate studies and who served on the committee until 1986.

I would like to thank my parents for their love and support. They have always been there when I needed them.

Finally, I want to dedicate this work to my wife Elena, whom I met when I first came to Rhode Island, and who became my light in the darkness and my hope during the difficult steps.

To my wife Elena

PREFACE

This dissertation follows the format of the MANUSCRIPT PLAN, as described in section 11-3 of the Graduate Manual of the University of Rhode Island. The work is presented in three sections. Section I contains the introduction to the project and the objectives of the study. Section II consists of five manuscripts, which represent the core of the study. At the time of the defense, three manuscripts will have been either accepted or at least submitted to international scientific journals. The two other manuscripts are written according to the format suitable for publication. Section three consists of the specific details related to the various aspects of the study. The appendices thus describe either certain experimental considerations or computer manipulations. Appendix A is a manuscript published in Drug Development and Industrial Pharmacy, that is related to a study performed at Miles Pharmaceuticals, West Haven, CT, where the author of this thesis was awarded a Graduate Student Summer fellowship. Appendix B reports a study performed at the University of Rhode Island, in association with Dr. J. L. Kanig, who is an adjunct Professor in the Department of Pharmaceutics. Appendices C to I allows the reader to repeat certain procedures or to obtain more details regarding the computer work.

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INTRODUCTION

Many of the new chemical entities recently synthesized or discovered in the pharmaceutical industry possess poor physico-chemical properties, which impact adversely on formulation. Thus low solubility, limited stability and erratic bioavailability sometimes make the development of acceptable dosage forms impossible. In addition, some drugs formerly discarded for their formulation problems have recently attracted new interest. One major new tool of solubilization and stabilization of drugs in the pharmaceutical industry has been the use of cyclodextrins.

Cyclodextrins are cyclic oligosaccharides. The three major forms of cyclodextrins viz α , β and γ consist of six, seven or eight units of glucose, respectively (1). The size of the cavity of these molecules gives the unique opportunity to adjust the affinity between the drug and the complexing agent, and therefore to optimize the complexation (2). There has been keen interest in the phenomenon of cyclodextrin complexation for the past decade or so, in the field of chemicals, agriculture and pharmaceuticals (3,4,5) as well as in the food industry. The affinity of certain molecules for cyclodextrins has been the basis of many applications of cyclodextrin complexation. Cyclodextrins have been used as enzymatic model in biochemistry and as a separation material in analytical

chemistry (6). The terms clathrate, complex, inclusion compound and monomolecular encapsulate have all been used to refer to cyclodextrin complexation.

Within the pharmaceutical industry, there are three advantages to cyclodextrin complexation:

- a) the improvement of solubility, dissolution rate and bioavailability (7,8,9);
- b) the stabilization of drugs (10,11);
- c) the amelioration of organo-leptic properties such as smell or taste.

Numerous excellent reviews of the subject of complexation of drugs with cyclodextrins have been published in the past few years. Special references will be given to two books, edited by Sjeztli (1) and Duchene (3). The review articles by Uekama (4) and Hegde (12) give a complete overview of history, structure, toxicity, uses and limitations of cyclodextrins in the pharmaceutical industry.

Numerous publications have appeared in the literature, especially from Japan, during the past ten years. The structure of the work done is usually presented as a sequence of four steps (13,14,15):

- a) results showing the complexation of the drug considered with cyclodextrin using the solubility method, spectroscopy, conductivity or diffusion;
- b) preparation of the complex by freeze-drying or solvent evaporation;

- c) results of the physical properties of the complex prepared using Proton Magnetic Resonance (PMR), X-Ray crystallography, Infra-Red spectroscopy or Differential Scanning Calorimetry (DSC), and elucidation of the stoichiometry of the complex;
- d) evidence of the advantages of complex formation by in-vitro dissolution studies or in-vivo animal bioavailability results.

The two areas lacking in-depth work have been the design of dosage forms of the drug/cyclodextrin complex, and the study of the physico-chemical aspects of the complexation phenomenon.

In this introduction the different aspects of cyclodextrin complexation, which have been suggested and discussed in the literature, will be briefly reviewed. Special attention will be then given to the physical chemistry and the thermodynamics of complexation. The two main thermodynamic parameters considered will be the enthalpy, which corresponds to the heat exchange upon complexation, and the entropy, which may be largely regarded as reflecting the degree of order of the system. In addition, the current regulatory status of cyclodextrins in the Food and Drug industry world-wide will be discussed.

The mechanism of complexation of drugs with cyclodextrins is based on some basic characteristics of the cyclodextrin molecules. Each glucose unit included in the ring contains three hydroxyl groups: C(2) and C(3) are

secondary hydroxyl whereas C(6) is a primary hydroxyl. These hydroxyl groups are also of critical importance in the preparation of cyclodextrin derivatives, which have been synthesized and have great potential applications in the pharmaceutical industry (16). The hydroxyl groups laying on the outer surface of the ring give to the cyclodextrin molecule its hydrophilicity. The inside of the cavity is hydrophobic due to the glucosic oxygen bridges. Upon complexation, the apolar guest molecule will approach the host molecule of cyclodextrin and will fit into the cavity.

The mechanism of cyclodextrin complexation in water or aqueous buffered solutions is closely related to the nature of the solvent. Water can be considered as a mixture of free water molecules and structured material. The water molecules are subject to hydrogen bonding; when a large number of hydrogen bonds are established, a structure called clusters or ice-like hydrogen bonded material is formed (17). From a thermodynamic point of view, water is thus a very versatile solvent, with continual formation and rupture of hydrogen bonds. Water will influence the thermodynamics of complexation at two different levels: the water molecules trapped inside the cyclodextrin cavity are iceberg-like and enthalpy-rich, and a non-polar molecule will be also surrounded by structured water, likely to be perturbed upon complexation.

The complex formation can be represented by the schemes shown in Figures 1 and 2. Figure 1 represents the non polar

aromatic ring of a potential guest molecule, that will replace the energetically rich water molecules from the inside of the cyclodextrin cavity. The result of this displacement will be a favorable non polar/non polar interaction. A more detailed representation is shown in Figure 2.

The different steps occurring during the complexation formation process have been proposed as follows (18,19):

- a) the guest molecule or drug approaches the cyclodextrin molecule;
- b) the ice-like water trapped in the cyclodextrin cavity is displaced, with loss of structure and corresponding release of energy;
- c) the water molecules surrounding the apolar portion of the drug to be included are broken down and water molecules go into solution;
- d) substituents of the drug interact with either the rim or the inside of the cyclodextrin ring;
- e) hydrogen bonds may be formed between the drug and cyclodextrin;
- f) the water structure is reestablished around the drug and cyclodextrin.

The question of the exact driving force in complex formation has been subject to controversy. The different theories proposed are as following:

1. Hydrophobic interaction

The water molecules around the apolar drug are broken down. The increased degree of freedom of the liberated water molecules corresponds to a negative entropy and a positive enthalpy upon complexation.

2. Van der Waals interaction

This type of interaction can originate either from dipole/dipole (Keesom forces), dipole/induced dipole (Debye forces) or induced dipole/induced dipole (London forces) interaction. The Van der Waals forces are weak and are unlikely to establish stable complexes by themselves. These forces would correspond to a negative enthalpy and a negative entropy.

3. Hydrogen bonding

The inclusion complex formation involves three components: the drug or guest molecule, the cyclodextrin or host molecule and the solvent. Different possibilities of interaction exist between these three components. Not only drug and cyclodextrin can interact, but also the solvent interacts with both the drug and the cyclodextrin. Some substances have been shown to function as Hydrogen bond donors. However this mechanism of complexation is generally regarded of minor importance, since the average bond energy is only 2 to 8 Kcal.mol⁻¹.

4. Release of high energy water

Water can be considered as existing with two different structures: water molecules can be tightly bound to water-solvating charged sites to form hydrogen bonding; water molecules enclosed within the cyclodextrin cavity cannot have a full complement of hydrogen bonds and are enthalpy-rich. The expulsion of these enthalpy-rich water molecules into bulk water upon complexation seems to be a major factor in the mechanism. The thermodynamic parameters observed in this case are usually a negative enthalpy and a negative entropy.

5. Release of conformational strain

This mode of action should be especially of critical importance in the case of alpha cyclodextrin complexation, that has the lowest number of glucose units. The torsion of the cyclodextrin molecules would be affected upon penetration of the guest molecule into the cavity, thus releasing energy.

Even though no general rule has been entirely accepted, it appears that a combination of the different factors described earlier is likely to play a role in the complex formation. The relative combination of each factor will depend upon the drug and the type of cyclodextrin. However, the definite point of agreement is that the thermodynamics of complexation deserve attention and clarification. It is generally accepted that both the extent and the sign of the

thermodynamic parameters viz enthalpy and entropy will give a clear indication of the process involved. Furthermore, a variation of pH. leading to a change in the ionization of the drug will undoubtedly affect the polarity of the drug, thus influencing the complexation phenomenon.

One major concern in the pharmaceutical industry has been the regulatory status of cyclodextrins. The nature of the interaction forces involved in the drug/cyclodextrin complex formation should give to the final product the status of materials such as solid dispersions, which means that a simplified approval process with no recognized new chemical entity may apply. In order to be approved for commercial use in food and drug products, a compound has to be Generally Regarded As Safe (GRAS) and usually a Drug Master File (DMF) will then be established with the Food and Drug Administration (FDA). The acceptance of a DMF is based of the results of clinical, toxicological and metabolic studies. Usually not only acute toxicity results, but also results from chronic use of the substance are required by the FDA. In the case of cyclodextrins, it appears that acute and chronic toxicity have been carried out in rats and dogs, but there is limited long term toxicity data. A report by the Food and Agriculture Organization (FAO) classified cyclodextrins as enzymatically modified starches. This position seems to be confirmed by various European Health Agencies, which also recently considered β

cyclodextrin as an enzymatically modified starch. Since 1983, the use of β cyclodextrin is thus approved for stabilization of flavors. In 1986, France granted approval for the use of β cyclodextrin as a flavor carrier. A major breakthrough in the approval process has been the efforts of a Dutch starch company, Aebe, to gather data and to gain approval in Netherlands, Belgium and Luxemburg. German and Spanish authorities seem to follow the same position and are expected to approve cyclodextrins for food products (20). However, as far as the drug industry is concerned, the only products on the market world-wide are Prostaglandin/cyclodextrin complexes in Japan. Thus, Ono Pharmaceuticals has launched a tablet formulation in 1976 and an injectable dosage form in 1979 (21).

The objectives of this study were:

1. To contribute to the characterization of the physico-chemical parameters that control the complexation of drugs with cyclodextrins;
2. To evaluate the effect of pH and temperature on the complexation of some drugs with cyclodextrins and to estimate the thermodynamic characteristics of the system at various conditions;
3. To develop a mathematical model using non-linear regression to calculate enthalpy and entropy of complexation;
4. To evaluate the potential use of a new β cyclodextrin

derivative as a complexing agent;

5. To investigate the properties of β cyclodextrin as an additive in tablet formulation.

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Figure 1

Schematic representation of the formation
of cyclodextrin inclusion complexes

Figure 1
Schematic representation of the formation
of cyclodextrin inclusion complexes

Figure 2

Schematic representation of the thermodynamic process
of drug:cyclodextrin complexation

Figure 2
Schematic representation of the thermodynamic
process of drug:cyclodextrin complexation

SECTION II

MANUSCRIPT I

STUDIES OF THE EFFECT OF pH, TEMPERATURE
AND RING SIZE ON THE COMPLEXATION OF
PHENYTOIN WITH CYCLODEXTRINS

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INTRODUCTION

The molecular encapsulation of pharmaceutical substances by cyclodextrins has been shown to enhance the solubility and the dissolution of a drug (1,2), to decrease drug degradation (3) and to improve the absorption from the gastro-intestinal tract (4) and through the skin (5). Phenytoin, a widely used anticonvulsant, has a low aqueous solubility and follows non linear pharmacokinetics (6). A coprecipitate of phenytoin with polyvinylpyrrolidone has been shown to produce a significant increase in the drug dissolution rate (7). Phenytoin has been reported to interact with beta cyclodextrin and to form a 1:1 complex (8). The complexation of phenytoin with cyclodextrins could improve solubility and dissolution rate of the drug as well as decrease the intra- and inter-variability of the absorption in patients.

Numerous publications in the field of drug/cyclodextrin complexation describe briefly studies of the complexation by the solubility method, but do not always indicate the exact

conditions of the experiments (pH, ionic strength of the solutions). Few published studies are actually focused on the physico-chemical aspects of the complexation of drugs with cyclodextrins. The objective of the study was to investigate the effect of temperature and pH on the thermodynamics of the complexation, and to propose a model of the complexation mechanism, describing how both the unionized and the ionized forms of phenytoin interact with cyclodextrins.

EXPERIMENTAL

Materials

Phenytoin^a and alpha,beta and gamma cyclodextrins^b were used as received. All other reagents or solvents were of analytical grade. Deionized water was used to prepare all buffer solutions, which were adjusted to an ionic strength of 0.3 using potassium chloride.

Methods

Ultra-Violet analysis of phenytoin. Phenytoin was assayed using a Hewlett-Packard HP 8451A diode array Ultra-Violet spectrophotometer at a wavelength of 225nm. The Beer-Lambert law was obeyed and the molar absorptivity at 225nm was found to be 6.72×10^3 .

^aUSP grade. Sigma Chemical Co., St. Louis, MO.

^bAmerican Maize, Hammond, IN.

pKa determination. The pKa of phenytoin was determined from the solubility data using the Benet approach (9).

Solubility studies. The solubility method described by Higuchi and Lach (10) was used under different pH and temperature conditions. An excess of phenytoin was placed into screw-capped vials containing solutions of cyclodextrins at various concentrations. The vials were shaken using a rotating unit in a controlled temperature ($\pm 0.1^{\circ}\text{C}$) waterbath. The time needed to reach equilibrium was determined to be about four hours. The samples were then pipetted, filtered through glass wool and diluted before being analyzed to determine the total concentration of phenytoin. All sampling, filtration and dilution steps were carried out in a constant temperature walk-in oven.

RESULTS AND DISCUSSION

pKa determination

The solubility of phenytoin as a function of pH is shown in Figure 1. From the plot shown in Figure 2, the value of pKa was determined to be 8.2, which is in good agreement with the literature values (11,12).

Heat of solution

The solubility of phenytoin was determined at different temperatures. The Van't Hoff equation gives a relationship between the solubility expressed in mole fraction and the absolute temperature for an ideal solution:

$$-\log X_i = (\Delta H_f / RT) + \text{constant} \quad (1)$$

where X_i is the molar solubility in mole fraction

ΔH_f is the heat of solution

R is the constant of perfect gases

and T is the absolute temperature

Table 1 shows the values of the heat of solution at different pH values. As pH increases, the heat of solution decreases, showing that the ionized form of phenytoin needs less energy to go into solution.

Effect of ring size

The solubility measurements were carried out using three cyclodextrins viz alpha, beta and gamma. Figure 3 shows the solubility isotherms of phenytoin with the three cyclodextrins at 25°C. Phenytoin interacts with alpha and gamma cyclodextrins, but to a much lesser extent than with beta cyclodextrin. According to the Higuchi and Lach approach (10), and assuming the formation of a 1:1 complex, the apparent thermodynamic constant of complexation could be calculated from the slope and the intercept of the straight lines obtained in Figure 3.

$$K = S_1 / I (1-S_1) \quad (2)$$

where K is the apparent thermodynamic constant

Sl is the slope of the solubility isotherm
and I is the intercept of the solubility isotherm

The values of the complexation constants were determined to be 110 M^{-1} , 850 M^{-1} and 149 M^{-1} at 25°C and pH 7 for alpha, beta and gamma cyclodextrins respectively.

Effect of temperature

Figure 4 shows the plot of the solubility of phenytoin as a function of cyclodextrin concentration at temperatures ranging from 20°C to 50°C and pH 5. At each temperature, the apparent complexation constant was calculated using equation (2). The apparent change in free energy ΔG was then computed from the following equation:

$$\Delta G = - R T \ln K \quad (3)$$

The thermodynamic parameters of complexation were determined using the Gibbs equation:

$$\Delta G = \Delta H - T \Delta S \quad (4)$$

Figure 5 shows the plot of ΔG as a function of T at pH 5. The slope and the intercept correspond to the entropy (ΔS) and the enthalpy (ΔH) of complexation respectively.

Effect of pH

Figure 6 shows the solubility isotherms of phenytoin/cyclodextrin complexation at various pH values. From the straight lines obtained, the apparent values of the complexation constant and the free energy were determined and are shown in Table 2 at a given temperature of 20°C. As pH increased the slope value increased showing that the ionized form of the drug does interact with beta cyclodextrin. However the interaction of the ionized form is weaker than for the unionized species.

The same experiment was performed at different temperatures. Table 3 shows the values of the overall entropy and enthalpy of complexation, calculated from the Van't Hoff plots, similar to that shown in Figure 5. The values in Table 3 are actually the geometric mean of the individual thermodynamic parameters describing the complexation for both the unionized and the ionized forms of phenytoin.

As pH increased, the intercept of the straight lines shown in Figure 6 increased, due to the ionization of phenytoin, which has a pKa of 8.3. However, as pH increased, the slope of the solubility isotherms also increased, clearly demonstrating a further increase of the total phenytoin concentration as shown in Figure 7.

The following model was developed to describe the complexation process:

$$K_i = (A^-/CD) / (A^-)(CD)_i \quad (6)$$

where K_u and K_i are the apparent thermodynamic complexation constants for the unionized and ionized forms of phenytoin respectively. $(CD)_u$ and $(CD)_i$ correspond to the free cyclodextrin concentration at pH 5 and pH>7 respectively.

At any pH value, the total concentration of phenytoin and cyclodextrin, respectively noted $(Phenytoin)_t$ and $(CD)_t$, can be expressed as:

$$(Phenytoin)_t = (AH) + (AH/CD) + (A^-) + (A^-/CD) \quad (7)$$

$$(CD)_t = (CD)_i + (AH/CD) + (A^-/CD) \quad (8)$$

By substituting at any given temperature one set of data (pH 5) from another (pH>7), it is possible to obtain a plot of $(A^-) + (A^-/CD)$ as a function of $(CD)_t$, as shown in Figure 8. The intercept corresponds to the concentration of the ionized species of phenytoin, and the slope can be written as:

$$\text{slope} = (A^-/CD) / (CD)_t \quad (9)$$

By combining equations (7), (8) and (9):

$$\begin{aligned} \text{slope} = K_i (A^-) (CD)_i / (CD)_i + K_u (AH) (CD)_u \\ + K_i (A^-) (CD)_i \end{aligned} \quad (10)$$

The expression $K_u (AH) (CD)$ corresponds to (AH/CD) at pH 5. The concentration of the free cyclodextrin varies with pH, but can be expressed as a function of $(CD)_u$ at pH 5, according to the following equation:

$$(CD)_i = r (CD)_u \quad (11)$$

where r is the proportionality factor between $(CD)_i$ and $(CD)_u$.

Therefore, at pH>7:

$$\text{slope} = K_i (A^-) r / (r + K_u (AH) + K_i (A^-) r) \quad (12)$$

AH and A^- represent the intercepts shown in Figure 6. AH is the intercept (I) at pH 5, and A^- is the intercept (I^-) at pH>7.

If Sl^- represents the slope of the solubility isotherm at pH 7:

$$Sl^- = K_i I^- r / (r + K_u I + K_i I^- r) \quad (13)$$

$$r Sl^- + K_u I Sl^- + K_i I^- r Sl^- = K_i I^- r \quad (14)$$

$$K_i = (Sl^- (r + K_u I)) / (I^- r (1 - Sl^-)) \quad (15)$$

Equation 15 allows for the determination of the apparent complexation constant of the ionized form of phenytoin with beta cyclodextrin.

The apparent values of the complexation constant (K_i) and the free energy (ΔG_i) were computed at different temperatures, and the Gibbs equation was used to determine the enthalpy (ΔH_i) and entropy (ΔS_i) of the ionized species of phenytoin with beta cyclodextrin.

The values are reported in Table 3. The coefficient of correlation of the linear plot obtained at pH 5 was high ($r=0.966$ in Figure 5). However, in the case of the complexation of the ionized species with beta cyclodextrin, the coefficient of correlation was lower (from $r=0.8$ to $r=0.6$). The relatively poor correlation is probably due to the noise in the experimental values and to the fact that the data used for the plots from pH 7 to pH 10 were obtained by subtraction of one set of data from another. Thus, the thermodynamic parameters obtained for the ionized species, since they are derived from two sets of experimental data, have the potential of twice as much an experimental error than in the case of the unionized species. At pH 7 and pH 8, the plot was linear at a 90% confidence level. The use of a computerized approach, providing a more precise determination of the thermodynamic parameters together with the exact standard deviation of the values, would allow a more extensive discussion of the results.

SUMMARY

Phenytoin interacts much more strongly with beta cyclodextrin than with alpha or gamma cyclodextrins. This

result is in accordance with previously reported work, indicating that geometric parameters play a major role in controlling the mechanism of complexation (the hand in glove theory).

The study at various pH values shows that both the non-ionized and the ionized forms of phenytoin interact with cyclodextrins. If the purely geometric consideration were to be confirmed, the ionized species should either not interact at all, or interact to the same extent. The ionized form of phenytoin is shown here to interact to a much lesser extent, suggesting that some chemical parameters are involved in the complexation.

There are a few examples in the literature show the interaction of both species (15-18). The possible delocalization of electrons or the influence of the energy of solvation have been proposed but no definite explanation has been established.

The biopharmaceutical implications of the interaction of cyclodextrins with both species are quite significant. For an orally administered dosage form of the complex, the change of pH through the gastro-intestinal tract will eventually lead to a different overall amount of bound drug, with a clear effect on the bioavailability of the drug. For an intravenous injection, the dissociation of the complex would occur at pH 7.2 to 7.6. The assumption of a solely unionized form of the bound drug to cyclodextrin with both the unionized and the ionized forms free would then create

an error in the results. For any type of interaction, Riegelman described the free drug as the activity factor and the bound drug as the capacity factor. The possible complexation of the ionized species with cyclodextrins in an intravenous solution would actually decrease the amount of free drug, but would also increase the amount of bound drug, therefore increasing the effective overall biological half-life.

ACKNOWLEDGEMENTS

The authors are grateful to Miles Pharmaceuticals for financial support in the study, and American Maize for providing the cyclodextrin materials.

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Table 1
Apparent heat of solution of phenytoin
at various pH values

pH	ΔH_f (Kcal/mole)
5	4.208
7	4.224
8	3.329
9	2.325

Table 2

Apparent values of the complexation constant
and the free energy of complexation for phenytoin
and beta cyclodextrin at 20° C

pH	K (M^{-1})	ΔG (Kcal/mole)	ΔG (KJ/mole)
5	847.9	- 3.93	- 16.43
7	923.9	- 3.98	- 16.64
8	695.9	- 3.82	- 15.97
9	277.0	- 3.28	- 13.71

Table 3

Enthalpy and entropy values for the
complexation of phenytoin (ionized and unionized)
with beta cyclodextrin at various pHs

	pH 7	pH 8	pH 9	pH 10
ΔH_i (Kcal/mole)	- 5.0	- 6.3	- 7.3	- 5.5
ΔS_i (cal/mole/K)	- 5.0	- 9.7	-14.0	- 9.8
pH 5				
ΔH_u (kcal/mole)	- 9.1			
ΔS_u (cal/mole/K)	-16.0			

Figure 1
Solubility profile of phenytoin at 25°C

SOLUBILITY PROFILE OF PHENYTOIN AT 25°C

Figure 2

Plot of $\log (S/S_o - 1)$ vs pH at 25°C

Figure 3

Complexation of phenytoin with α , β and γ cyclodextrins

Complexation of phenytoin with α , β , and γ cyclodextrins
at 20°C and pH 7

Figure 4

Complexation of phenytoin with β cyclodextrin at pH 5

Figure 5

Determination of the overall enthalpy and entropy
for the complexation of phenytoin with β cyclodextrin

Figure 6

Effect of pH on the complexation of phenytoin
with β cyclodextrin at 20°C

Figure 7

Effect of pH on the complexation of phenytoin
with β cyclodextrin at 20°C
un = non-ionized

The lines noted un are parallel to the pH 5 solubility
isotherm

Effect of pH on the complexation of phenytoin with β cyclodextrin. un = unionized. The lines noted un are parallel to the pH 5 solubility isotherm

Figure 8

Solubility isotherms for the complexation of
ionized phenytoin with β cyclodextrin at pH 8

MANUSCRIPT II

ABSTRACT

A new computer method was proposed to calculate the thermodynamic parameters of drug/cyclodextrin complexation. The method allowed the simultaneous determination of two or more parameters using non-linear regression. The mathematical model was developed and used to obtain the estimates of enthalpy, entropy and solubility parameters. The output of the NONLIN program also provided the value of the standard deviation of the parameters, thus allowing meaningful comparisons between different systems.

Key words: Non-linear regression, Thermodynamics, Phenytoin, Cyclodextrin, Complexation.

Determination of Thermodynamic Parameters of Complexation
using a Non-linear Regression Method

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INTRODUCTION

The complexation of pharmaceutical substances with cyclodextrins has been shown to improve stability (1,2), solubility (2), dissolution rate (3) and bioavailability properties (4). The physico-chemical aspect of the complexation phenomenon and especially the determination of the thermodynamic parameters of complexation are essential to characterize the drug/cyclodextrin interaction. Different methods have been suggested for the determination of the values of both the enthalpy and the entropy of complexation (5). The microcalorimetric method has the advantage of using direct calorimetric measurements and has been proposed as the most reliable method (6). However, this method is not always readily available in the laboratory and requires specific expertise (7). Another widely used method is based on the temperature dependence of the complexation constant (8). The complexation constant and the free energy can be determined from solubility data

using the Higuchi and Connors equation. After constructing the solubility isotherms at different temperatures, the enthalpy and entropy of complexation can be obtained from the plot of the free energy as a function of the absolute temperature. The major limitation of the temperature dependence method is the lack of precision of the thermodynamic parameters (9). The two successive steps requiring the use of linear regression really introduce an error, which can be critical in the comparison process of the parameters. The purpose of this paper is to present a novel computerized method allowing the direct treatment of the raw experimental data to determine the thermodynamic parameters of the drug/cyclodextrin complexation using non-linear regression. A similar approach has already been used by Dr. Fung for the determination of a single parameter viz the energy of activation from raw experimental data of drug stability (10). In this paper we describe a method of general applicability for the simultaneous determination of two or more parameters.

THEORY

The first step is to develop a mathematical model, which describes the relationship between the concentration of the drug, the concentration of cyclodextrin (CD) and temperature (T). The drug noted AH will be used as the model molecule interacting with cyclodextrins. The drug is

subject to ionization, but will be assumed to be in its non-ionized form in the first part of the paper.

Based on a 1:1 stoichiometric ratio, the complexation reaction can be written as follows:

The overall complexation constant K is expressed as:

$$K = [AH/CD] / [AH]f[CD]f \quad (2)$$

where the notation f describes the free concentration.

It is possible to express the apparent free energy of complexation ΔG using the relationship:

$$\Delta G = -R T \ln K \quad (3)$$

where: T is the absolute temperature (K)

and R is the gas constant

The Gibbs equation shows the relationship between free energy (ΔG), enthalpy (ΔH) and entropy (ΔS) of complexation:

$$\Delta G = \Delta H - T \Delta S \quad (4)$$

The overall complexation constant K can be expressed, after rearrangement of equations (3) and (4) as:

$$K = e^{\Delta H/RT - \Delta S/R} = e^A \quad (5)$$

with $A = \Delta H/RT - \Delta S/R$

A typical plot of solubility data for the drug/CD interaction is shown in Fig. 1. The total concentration of drug is plotted versus the total cyclodextrin concentration.

The intercept of the straight line with the y-axis corresponds to the concentration of the free drug molecule, whereas the increase of the drug concentration represents the concentration of the complexed form.

The following equations can be written to ascribe the total concentration of both the drug and the complexing molecule:

$$[AH]_t = [AH]_f + [AH/CD] \quad (6)$$

$$[CD]_t = [CD]_f + [AH/CD] \quad (7)$$

where the notation t stands for total concentration

$$\text{From Eq. (6): } [AH/CD] = [AH]_t - [AH]_f \quad (8)$$

$$\text{From Eq. (7)(8): } [CD]_f = [CD]_t - [AH]_t + [AH]_f \quad (9)$$

By substituting $[AH/CD]$, $[AH]_f$ and $[CD]_f$ in Equation (2):

$$K = [AH]_t - [AH]_f / [AH]_f ([CD]_t - [AH]_t + [AH]_f) \quad (10)$$

The term $[AH]_f$, which is the free guest concentration, represents the actual solubility of the guest molecule, which varies with temperature according to the Hildebrand equation:

$$-\ln [AH]_f = \Delta H_f (T_o - T) / R T T_o + \ln \gamma \quad (11)$$

Eq. (11) can be rearranged, and $(AH)_f$ is then expressed as:

$$[AH]_f = e^{\Delta H_f (T - T_o) / R T T_o - \ln \gamma} = e^B \quad (12)$$

where: ΔH_f = heat of solution

T = absolute temperature

T_o = melting point

γ = activity coefficient

R = constant of perfect gases

The term B can be written as:

$$B = \Delta H_f(T - T_0) / R T_0 - \ln \gamma$$

Eq. (10) can then be arranged according to Higuchi and Connors, as:

$$[AH]_t = K [AH]_f / (1 + K [AH]_f) [CD]_t + [AH]_f \quad (13)$$

The terms K and [AH]_f can be substituted by the exponential expression, and eq. (13) now becomes:

$$[AH]_t = e^A e^B / (1 + e^A e^B) [CD]_t + e^B \quad (14)$$

or:

$$[AH]_t = e^{A+B} / (1 + e^{A+B}) [CD]_t + e^B \quad (15)$$

Eq. (15) fully describes the relationship between the total concentration of the drug (the dependent variable), the total concentration of cyclodextrin (the first independent variable) and the absolute temperature (the second independent variable), which is included in the expression of both terms A and B.

A and B also include the thermodynamic parameters of complexation (ΔH and ΔS) and the free guest solubility parameters (ΔH_f and $\ln \gamma$), respectively.

Eq. (15) will be used as the mathematical model, allowing for the direct treatment of the raw experimental data. Eq. (15) can therefore be applied to any type of interaction, in which the stoichiometric ratio complexation is known to be 1:1.

In the case of the complexation of phenytoin with β cyclodextrin, some previously published work has shown that

the stoichiometry of the complex is indeed 1:1 (11). This result was obtained after preparation of the freeze-dried form of the complex and analysis using X-Ray, PMN and DSC (12).

Figure 2 shows a typical solubility isotherm at pH>7 together with that at pH 5. Assuming that the complexing ability of cyclodextrins is pH-independent, the further increase of the slope corresponds to the complexation of the ionized form of the drug with β cyclodextrin.

Phenytoin is a weak acid, with a pKa of 8.3 (13,14), and has already been reported to interact with β cyclodextrin both as the non-ionized and as the ionized forms (9). At low pH values (pH 5), phenytoin is non-ionized, and Eq. (15) can be applied to study the complexation phenomenon from the solubility data. However, at higher pH values, the complexation of the ionized species with β cyclodextrin has to be taken into consideration. An extension to the model is thus needed.

The symbols used in the development of the equations will take into consideration both complexation phenomena. If AH represents the non-ionized form of the drug and A^- the ionized species, the different equations can be written as follows:

$$K^u = [AH/CD] / [AH]^u f [CD] f \qquad K^i = [A^-/CD] / [A^-]^i f [CD]^i f \quad (18)$$

$$\Delta G^u = - R T \ln K^u \qquad \Delta G^i = - R T \ln K^i \quad (19)$$

$$\Delta G^u = \Delta H^u - T S^u \qquad \Delta G^i = \Delta H^i - T \Delta S^i \quad (20)$$

$$\ln[AH] = Hf^u(T-To)/RTTo - \ln \gamma^u \qquad \ln[A^-] = Hf^i(T-To)/RTTo - \ln \gamma^i \quad (21)$$

Eq. (14) has already been developed:

$$[AH]^u_t = (e^{Au+Bu}) / (1 + e^{Au+Bu}) [CD]_t + e^{Bu} \quad (15)$$

The intercept of the solubility isotherm at pH 7 can be expressed as:

$$I^i = I^u + [A^-] f \quad (22)$$

or

$$[A^-]^i f = I^i - I^u \quad (23)$$

The total drug concentration $[AH]_t$ can be written:

$$[AH]^i_t = [AH]^u + [AH/CD]^u + [A^-]^i f + [A^-/CD]^i \quad (24)$$

or

$$[AH]^i_t = [AH]^u_t + (I^i - I^u) + [A^-/CD]^i \quad (25)$$

$$\text{and: } [A^-/CD]^i = [AH]^i_t - [AH]^u_t - [I^i - I^u] \quad (26)$$

$$[AH/CD]^u = [AH]^u_t - I^u \quad (8)$$

The total cyclodextrin concentration $[CD]_t$ can be written as:

$$[CD]^i_t = [CD]^i f + [AH/CD]^u + [A^-/CD]^i \quad (27)$$

or

$$[CD]^i f = [CD]^i_t - [AH/CD]^u - [A^-/CD]^i \quad (28)$$

The apparent complexation constant K^i under the current conditions can then be expressed as:

$$K^i = [A^-/CD]^i / [A^-]^i_f [CD]^i_f \quad (18)$$

$$K^i = ([AH]^i_t - [AH]^u_t - (I^i - I^u)) / (I^i - I^u) ([CD]^i_t - [AH/CD]^u - [A^-/CD]^i) \quad (29)$$

$$K^i = ([AH]^i_t - [AH]^u_t - (I^i - I^u)) / (I^i - I^u) ([CD]^i_t - ([AH]^u_t - I^u) - ([AH]^i_t - [AH]^u_t - (I^i - I^u))) \quad (30)$$

$$[AH]^i_t - [AH]^u_t - (I^i - I^u) = K^i (I^i - I^u) ([CD]^i_t - ([AH]^i_t - [AH]^u_t - (I^i - I^u)) - ([AH]^u_t - I^u)) \quad (31)$$

$$[AH]^i_t - [AH]^u_t - I^i + I^u = (K^i I^i - K^i I^u) ([CD]^i_t - [AH]^i_t + I^i) \quad (32)$$

After rearrangement:

$$[AH]^i_t = (K^i I^i - K^i I^u) / (1 + K^i I^i - K^i I^u) [CD]^i_t + [AH]^i_t + (I^i - I^u) (1 + K^i I^i) \quad (33)$$

Equation (33) shows the relationship between the total drug concentration at pH 7 (the dependent variable), the total cyclodextrin concentration (the first independent variable) and the temperature, at which the complexation occurs (the second independent variable).

The terms I^u and $[AH]^u_t$ correspond to the free drug concentration and the total drug concentration at pH 5, respectively. Since the complexation ability of cyclodextrin is assumed to remain constant through the pH range studied, I^u and $[AH]^u_t$ remain constant.

COMPUTER TREATMENT OF DATA

The experimental data treated using the novel computer method have been published previously (9). The lack of precision of the thermodynamic parameters was the main limitation of the results, thus an extensive discussion of the thermodynamics of complexation was rather difficult.

The NONLIN computer program was used to determine the final estimates of all thermodynamic parameters. The NONLIN regression approach has been successfully used to determine parameters in many fields such as pharmacokinetics (15) and protein binding (16). The different programs needed to treat the data were:

- 1) the datafile, which included all the experimental data, the initial estimates and the limits of the parameters;
- 2) the subroutine program consisting of the declaration of the different variables and the full mathematical model describing the complexation;
- 3) a link program, which allows for the iterations of the NONLIN program to take place and the data to be called in.

The method of determining the parameters using the NONLIN computer was validated by running a set of simulated data. The simulated data were obtained by giving a specific value to each of the parameters appearing in eq. 15. The enthalpy and the entropy of complexation were assigned a value of $-10.0 \text{ Kcal.mol}^{-1}$ and -10.0 eu respectively. The solubility parameters used were $15.0 \text{ Kcal mol}^{-1}$ for the heat of solution and 5.0 for the activity coefficient. All the concentrations were rounded to two significant digits.

The schematic representation of the programs used is shown in Figure 3. Eq. (15) corresponds to the complexation at pH 5 of the unionized species, whereas Eq. (33) fits the data obtained when both species form a complex. For a set of data at $\text{pH} > 7$, the subroutine program was designed so that the complexation occurring between the unionized species and the β cyclodextrin was taken into account. Thus, for each temperature and at each cyclodextrin concentration, the program regenerated the drug concentrations calculated at pH 5, based on the previous determination of ΔH^u and ΔS^u .

RESULTS AND DISCUSSION

Validation of the program

The datafile created from the set of simulated data was run with a subroutine program allowing the determination of the four unknown values, namely enthalpy, entropy, heat of solution and activity coefficient. The NONLIN program converged to the values shown in Table 1. The values determined by the computer were in excellent agreement with the theoretical values. The results clearly show that this method is perfectly applicable for the determination of four unknown values from a set of 24 concentrations; in this case four temperatures and six cyclodextrin concentrations.

Advantages and general application of the method

All the equations, such as the expression of the equilibrium complexation constant and the Gibbs equation, are already included in the general equation; there is thus no need of the successive preliminary plots to determine the

thermodynamic parameters of complexation. This method not only allows a direct treatment of the raw experimental data, but also gives a clear indication of the statistical significance and the level of confidence associated with the determined parameters.

Although the method has been developed for the case of the complexation of drugs with cyclodextrins, as far as the major assumptions of the model remain acceptable, this method is applicable to many other cases of drug interaction or molecular recognition.

Characterization of the phenytoin/BCD interaction

Since our objective was to evaluate the effect of pH on the interaction of phenytoin with β cyclodextrin, a typical set of data consisted of a set of phenytoin concentrations (independent variable) as a function of β cyclodextrin concentration and temperature (dependent variables). Six levels of β cyclodextrin concentrations and four levels of temperatures were used to build a database of 24 points at different pH values from pH 5 to pH 10.

Table 2 shows the values of the overall thermodynamic parameters determined using the NONLIN program, at different pH values. The enthalpies and entropies actually represent the geometric means for the complexation of both the non-ionized and the ionized species of phenytoin with β cyclodextrin. As pH increases, the relative standard of deviation increases, probably due to the much higher concentrations of phenytoin at pH 9 or 10 compared to those

at pH 5, when the drug is non ionized. The large negative values of enthalpy suggest a strong dipole interaction, either due to the interaction between the drug and the cyclodextrins or due to the change of behavior of the water molecules associated with both the drug and the cyclodextrin. The negative entropies suggest that the solvent has a more ordered structure after complex formation. At this point, it is difficult to be more specific, since the values in Table 2 are not exclusively related to either form of the drug, but rather represent an overall estimate of the thermodynamic changes occurring during the formation of the phenytoin/ β cyclodextrin complex.

In order to further analyze the results, the more sophisticated mathematical model described in eq. (33) had to be used. In order to run the subroutine program for datafiles at pH>7, the respective values $-9,714.9 \text{ cal.mol}^{-1}$ and -10.49 e.u. were used for the enthalpy and the entropy of complexation of the unionized form of phenytoin with β cyclodextrin. It is assumed that the complexation of the non-ionized drug with cyclodextrin is not affected by change in pH used in the study, which also means graphically that a parallel line to the pH solubility isotherm could actually be drawn at any pH value. The results of the treatment of the raw experimental data using the computer method are shown in Table 3. The correlation between the data and the theoretical model of eq. (33) was excellent with a

coefficient of correlation from 0.99 to 0.97, with 24 points ie. 20 degrees of freedom.

The negative values of enthalpy show again a strong involvement of dipole interaction. It can be mentioned that the enthalpy of complexation is lower at pH 7 and pH 8, where the drug is only slightly ionized. The value increases at pH 9 and pH 10, suggesting a possible contribution of the charge of the ionized phenytoin molecule. A widely proposed theory is that the apolar and non-ionized drug molecule leaves behind a structured cavity, which allows the solvent molecules to redistribute upon complexation. This aspect of the mechanism is likely to vary as a function of pH, since the polarity of phenytoin is pH-dependent. The relative standard deviation is very high at pH 7, showing that the low percentage of ionization does not allow an accurate determination of ΔH . The values of entropy become large at pH 9 and pH 10, suggesting a much more ordered solvent after complexation. At pH 7 and pH 8, the standard deviations are very high, and the positive value of entropy probably only indicates little or even no entropy change.

SUMMARY

The new model and method introduced here allow the determination of more than one unknown from one set of raw experimental data. Even though the method was used to treat data from the complexation of phenytoin with cyclodextrins,

the same approach could be exploited for other drug interaction system. The method allows for a meaningful comparison of thermodynamic parameters for the interaction of either one drug with different cyclodextrins or one cyclodextrin with different drugs. The use of the approach could enhance the ability of the pharmaceutical scientist to analyze data from the solubility method to propose a rationale of interaction between drugs and complexing agents.

ACKNOWLEDGEMENTS

The authors are grateful to Dr. Ho-Leung Fung (SUNY at Buffalo) and Dr. Douglas S. Greene (Lederle Labs.) for their useful assistance in this study. Also we thank American Maize for the generous supply of cyclodextrins used in this study.

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Table 1

Thermodynamic parameters of complexation determined
from a set of simulated data

Parameter	Theoretical Value	Final Estimate	Standard Deviation	95% Conf. Limit
Enthalpy				
ΔH (cal.mol ⁻¹)	-10,000	-10,164	60	9,959-10370
Entropy				
ΔS (e.u.)	-10.0	-10.55	0.19	9.91-11.19
Heat				
ΔH_f (cal.mol ⁻¹)	15,000	15,154	57	14,960-15,349
Activity				
$\ln \gamma$	5.0	4.43	1.0	3.87-5.05

Table 2
Overall enthalpies and entropies of complexation
for the phenytoin/ β -cyclodextrin interaction

pH	ΔH (Kcal.mol ⁻¹)	RSD (%)	ΔS (e.u.)	RSD (%)
5.0	- 9,721	1.1	-10.49	3.3
7.0	-11,528	1.2	-16.32	2.8
8.0	- 9,292	1.3	-10.02	3.8
9.0	- 8,802	4.2	-10.18	11.8
10.0	- 8,982	12.9	-12.44	30.5

Table 3
Enthalpies and entropies for the complexation
of phenytoin ion with β -cyclodextrin

pH	r	ΔH^i (cal.mol ⁻¹)	RSD (%)	ΔS^i (e.u.)	RSD (%)
7.0	0.998	- 5,326	40.4	1.68	405.0
8.0	0.999	- 5,352	5.8	1.35	74.0
9.0	0.995	- 8,899	4.0	- 10.75	10.7
10.0	0.973	-12,934	5.2	- 25.41	8.6

Figure 1

Typical solubility isotherm for the complexation
of non-ionized phenytoin with β CD at pH 5

Figure 1: Typical solubility isotherm of the complexation of non-ionized phenytoin with β -cyclodextrin at pH 5

Figure 2
Typical solubility isotherm of phenytoin
with β CD at pH>7

Figure 2: Typical solubility isotherm of the complexation of phenytoin with β -cyclodextrin at pH 7

Figure 3

Schematic representation of the different programs

Figure 3: Schematic representation of the different programs

MANUSCRIPT III

Potential Pharmaceutical Applications
of a New Beta Cyclodextrin Derivative

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INTRODUCTION

The three major natural cyclodextrins viz alpha, beta and gamma, consist of six, seven or eight glucose units, respectively (1). In addition to the natural cyclodextrins, numerous new products have been shown to have potential for the pharmaceutical industry (2,3). A clear distinction has to be made between cyclodextrin polymers, which are mainly used as separation materials in analytical chemistry (4) and cyclodextrin derivatives, which are monomers of substituted cyclodextrins (5). In the synthesis process of the latter, both the C(2) and C(3) secondary and the C(6) primary hydroxyl groups have been the target of many chemical substitutions (6), leading to the development and the characterization of cyclodextrin derivatives of pharmaceutical interest (7). Such derivatives have been successfully used in the design of new drug carrier systems (3,5). Hence, in addition to their increased solubility, these chemically modified cyclodextrins exhibit excellent complexing abilities (8). The different possibilities of modifying cyclodextrins have been reviewed by Seville (2),

Szejtli (1) and Uekama (3). Szejtli also published an extensive list of the major cyclodextrin derivatives under investigation until 1982 (1).

The chemical considerations of the substitution of cyclodextrins have been investigated by Harata et al., who examined the macrolytic conformation of permethylated cyclodextrins (6). The main factor is the loss of the ability to form hydrogen bonds upon substitution. Further studies dealing with the properties of cyclodextrin derivatives have been recently published (9).

The stability of prostaglandins was improved by complexation with methylated cyclodextrins (10). On the other hand, an ammonium beta cyclodextrin derivative was shown to catalyze the hydrolysis of nitrophenyl ester (11). Nakai et al. studied the effect of a tri-methyl beta cyclodextrin on the partition of nitrophenol, and interpreted the data as the formation of a 1:1 complex (12). The use of cyclodextrin derivatives as agents of solubilization has been reported in the literature (13). Muller and Brauns have investigated the complexation of diazepam, hydrocortisone and digitoxin with methyl beta cyclodextrin (14), as well as the effect of the derivatization of gamma cyclodextrin on the phase-solubility of progesterone (15). Otagiri et al. have also studied the complexation of several drugs with tri-methyl cyclodextrin. In most of the studies, the critical factor to be carefully considered in the interpretation of the data is the degree

of substitution (DS) of the cyclodextrin derivatives (16). Muller and Brauns have established a relationship between the average degree of substitution and the complexing ability (14). From the data obtained, the lower the degree of substitution, the more extensive the complexation phenomenon. This result was attributed to the steric blockage of the cyclodextrin cavity by the substituents.

Several researchers have published data regarding specific dosage forms of complexes of drug/cyclodextrin derivatives. Dimethyl beta cyclodextrin has been proposed as a parenteral drug carrier for such drugs as progesterone and hydrocortisone (17). The dissolution of flurbiprofen suppositories was shown to be greatly improved by complexation with dimethyl and trimethyl beta cyclodextrin (18). Also, a comparative study of the complexation of flurbiprofen with both beta cyclodextrin and its derivatives showed that the mode of inclusion was quite different, due to the different number of free hydroxyl groups on the cyclodextrin ring (18). In-vivo studies in rats suggested that the complexation of ketoprofen with methyl beta cyclodextrin could lead to high plasma levels (19). It was also proposed that the surface active properties of methyl beta cyclodextrins could facilitate drug absorption through membranes. However, as discussed further in this paper, the same surface active properties may well be responsible for negative toxic effects on the cells, causing hemolysis (3). Pitha et al. investigated the oral administration of

steroidal hormones. The study showed that the complexation of progesterone and testosterone with beta cyclodextrin derivatives allowed for the correction of serum levels and could efficiently be used for therapeutic applications (17).

Even though the hydrophilic cyclodextrins have been the focus of the attention of many researchers, Uekama recently published data showing that ethylated beta cyclodextrins could be used as hydrophobic drug carriers. An in-vivo study in rats showed that these hydrophobic derivatives produced a sustained release of diltiazem (21). It is very likely that more work will be done in the future to establish the exact potential of cyclodextrin derivatives in sustained release.

The purpose of this paper is to evaluate the complexing ability of hydroxyethyl beta cyclodextrin, a new beta cyclodextrin derivative. Furthermore, the complexation of several drugs with both the natural beta cyclodextrin and its new derivative was investigated in order to establish a possible comparison of the data and to propose a valid prediction of the overall increase in drug solubility upon complexation.

EXPERIMENTAL

Materials

Hydroxyethyl beta cyclodextrin (batch #CDM108) was obtained from American Maize (Hammond, IN). Table 1 shows

some of its basic physico-chemical properties, compared to those of the natural cyclodextrins.

Three of the drugs viz phenytoin, diazepam and hydrochlorothiazide were obtained from Sigma Chemical Co. (St. Louis, MO). Ibuprofen was generously given by Whitehall Laboratories. All drugs were used as received. All solvents and reagents used in the study were of analytical grade and were used as received.

Methodology

The complexation of the drugs with the new modified beta cyclodextrin was studied using the solubility method described by Higuchi and Connors (21). An excess of drug was placed into screw-capped vials containing various concentrations of cyclodextrins. The vials were then rotated at constant temperature ($\pm 0.05^{\circ}\text{C}$) in a waterbath, until the maximum solubility was reached. The samples of drug solutions were then filtered using glasswool, diluted and assayed for drug concentration using a diode-array ultra-violet spectrophotometer (Hewlett-Packard HP 8451A). The drug standards obeyed the Beer's law in all cases, and cyclodextrin was shown not to interfere with the assay.

RESULTS AND DISCUSSION

The solubility isotherms were obtained by plotting the total drug concentration as a function of total cyclodextrin concentration. In all cases the plots were

linear, suggesting a type A phase solubility (21). The intercept represents the concentration of the free drug, while the slope represents the solubilization power of the complexing agent. Figures 1, 2, 3 and 4 show the solubility isotherms of phenytoin, diazepam, ibuprofen and hydrochlorothiazide, respectively. Each plot shows the complexation data with both beta cyclodextrin and the hydroxyethyl derivative. The maximum concentration of beta cyclodextrin used was 15×10^{-3} M, except in the case of ibuprofen, where a maximum concentration of 5.0×10^{-3} M was used, due to the low solubility of the ibuprofen/beta cyclodextrin complex. The concentration of hydroxyethyl derivative varied from 0 to about 150×10^{-3} M (20% w/w) (although, according to the supplier, solutions of up to 60% solids could actually be prepared). The molar concentration was calculated using a molecular weight of 1300, value obtained from the average value of the degree of substitution. The concentrations of both complexing agents are plotted on the X-axis using the same scale; the use of the derivative thus corresponds to an extension of the X-axis.

From Figures 1 to 4, it appears that the slope of the solubility isotherms remained identical for both complexing agents. This result was confirmed statistically using a SAS program. The results thus suggest that even though the overall increase of drug solubility was much higher in the case of the derivative, the solubilizing power was

unchanged. This observation confirms the "hand and glove" theory used to describe how the steric factors control the extent of complexation. Some researchers have reported that the complexation of drugs with cyclodextrin derivatives was associated with a lower solubilizing power (14). These results were obtained in the case of hydroxypropyl beta cyclodextrin, with high degrees of substitution. The decrease of complexing ability was therefore suggested to be due to steric hindrance. In the case of hydroxyethyl beta cyclodextrin, it seems that the low degree of substitution presents the unique advantage of preserving the complexing ability of the cyclodextrin.

The reactions of complexation can be written as following:

The slope of the solubility isotherm corresponds to the concentration of the complexed drug over the total concentration of cyclodextrin, as shown in Figure 5.

Therefore:

$$S = [\text{Drug}/\beta \text{ CD}] / [\beta \text{ CD}] \quad (3)$$

$$S' = [\text{Drug}/\beta' \text{ CD}] / [\beta' \text{ CD}] \quad (4)$$

where S and S' represent the slope for the complexation of drugs with β and β' cyclodextrins respectively.

Assuming that the slope remains unchanged, the following equality can be written:

$$S = S' \quad (5)$$

$$[\text{Drug}/\beta \text{ CD}] / [\beta \text{ CD}] = [\text{Drug}/\beta' \text{ CD}] / [\beta \text{ CD}] \quad (6)$$

$$\text{and} \quad [\beta' \text{ CD}] = 10 \times [\beta \text{ CD}] \quad (7)$$

$$\text{Therefore:} \quad [\text{Drug}/\beta' \text{ CD}] = 10 \times [\text{Drug}/\beta \text{ CD}] \quad (8)$$

According to this model, the concentration of the complexed drug obtained upon complexation with the beta cyclodextrin derivative should be about ten times that observed in the case of the natural product. Both concentrations were calculated at different temperatures and at various pH values. The ratio was then computed and the results are reported in Table 2. The ratios are in good agreement with the theory; values were in the range of 8.6 to 11.0. Two values were calculated for phenytoin and diazepam, since for these drugs both the ionized and the non-ionized species have been shown to interact with cyclodextrins. In the case of ibuprofen, the value was found to be 26.6. Once the adjustment of the concentration ratio is made, this value corresponds to 8.9. It should be mentioned that the values of concentration of complexed drug reported in Table 2 represent almost the total concentration for ibuprofen, for which the interaction is very extensive. However, in the case of hydrochlorothiazide, the value

represents merely half of the concentration of the free drug, suggesting a very weak interaction.

A SASGRAPH software was used to plot the drug solubility as a function of cyclodextrin concentration and temperature. The three dimensional plots obtained are shown in Figures 6, 7, 8 and 9, for phenytoin, diazepam, ibuprofen and hydrochlorothiazide respectively. The overall volume of these plots allows a good visualization of the extent of complexation.

The practical implications of the results of this study can be discussed at two levels. Firstly, it appears that it may be possible to predict the solubility of drug upon complexation with hydroxyethyl beta cyclodextrin, based on the data obtained with the natural form. Beta cyclodextrin, which is a much more readily available material, could be used as a model to evaluate whether a drug concentration in the range desired could be achieved through beta cyclodextrin derivative complexation. The empirical rule suggested here corresponds to the multiplication of the solubility observed with beta cyclodextrin by the ratio factor between the solubility of beta cyclodextrin and its derivative. This rule has been shown here to stand very well in the case of four drugs, at different pH values and different temperatures.

The second point of interest resides in examining the range of solubility obtained after complexation. The complexation of drugs with cyclodextrins can be particularly

appropriate either to improve the physico-chemical properties or the organoleptic properties of drugs. Clearly, the concentrations of ibuprofen attained in this study are very high and would even make possible a liquid dosage form of the drug. However, ibuprofen has a bitter taste, that even a 60 to 70% complexation level may not totally mask. In the case of the other drugs, phenytoin is complexed at about 10% and diazepam around 20%. This percentage would be a critical factor in the development of any dosage form of a drug/cyclodextrin complex. Obviously, on a development point of view, the closer to a 100% complexation, the better. The results of the study indicate that the use of hydroxyethyl beta cyclodextrin may offer clear advantage to the pharmaceutical formulator.

CONCLUSIONS

The complexation ability of β' -cyclodextrin has been shown to be similar to that of the natural β cyclodextrin. The slope of the solubility isotherms was indeed identical for the four drugs studied. It thus appears that the substituents do not affect the basic mechanism of complexation. The particular advantages of β' cyclodextrin are its high solubility and its low degree of substitution. The range of drug concentrations achieved in the study suggests real new possibilities of formulation. A liquid dosage form of ibuprofen with high bioavailability could very well be developed. One of the major concerns in the

pharmaceutical industry has been the toxicity of the cyclodextrin derivatives, in particular unacceptable hemolytic properties. However, it has also been shown that the lower the degree of substitution, the lower the hemolytic properties. From the preformulation data presented here in the study, hydroxyethyl β cyclodextrin seems to be an excellent candidate for future drug/cyclodextrin complex formulation.

ACKNOWLEDGEMENTS

The authors thank American Maize for the generous supply of cyclodextrin samples. The offices of Mr. D. Downing and Mr. G. Reed worked with a particular diligence. The cooperation of Dr. A. Smith from Whitehall Laboratories is greatly appreciated in the supply of ibuprofen. The authors are also grateful to Dr. B. Birmingham for his expertise in plotting data on three-dimensional SAS graphs.

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TABLE I
BASIC PROPERTIES OF CYCLODEXTRINS

CD	# GLUCOSE UNITS	MW	# WATER MOLECULES	SOLUBILITY gm/100ml
α	6	972	6	14.5
β	7	1135	11	1.8
γ	8	1297	17	23.2
β'	7	1300	11	60.0

TABLE II
Concentrations of drug complexed with
 β and β' cyclodextrins at 50°C

DRUG	pH	[DRUG]f (mM)	[Drug/ β CD] (mM)	[Drug/ β' CD] (mM)	Ratio [*]
Phenytoin	5.0	0.26	1.28	12.69	9.9
	8.0	0.68	1.62	13.98	8.6
Diazepam	3.0	0.80	1.12	12.32	11.0
	6.0	0.29	0.71	7.82	11.0
Hydrochlorothiazide	5.0	7.65	3.05	32.30	10.6
Ibuprofen	5.0	0.16	0.34	9.08	8.9 ^{**}

* Ratio = [Drug/ β' CD] / [Drug/ β CD]

** The value of 8.9 was obtained by dividing 26.6 by 3
The maximum concentration of β CD was 5.0×10^{-3} M in
the case of ibuprofen and 15.0×10^{-3} M for the other
drugs

Figure I
Complexation of phenytoin with β CD at 40°C and pH 5

Figure II
Complexation of diazepam with β CD at 30°C and pH 6

Figure III
Complexation of ibuprofen with β CD at 50°C and pH 5

Figure IV
Complexation of HCTZ with β CD at 50°C and pH 5

COMPLEXATION OF HCTZ WITH β CD AT 50°C AND pH 5

Figure V
Complexation of drugs with β and β' cyclodextrins

Figure V
Complexation of drugs with both β CD and β' CD

Figure VI

Solubility of phenytoin as a function of
temperature and β' CD concentration at pH 5

PHENYTOIN, β , CD, FH 5

Figure VII

Solubility of diazepam as a function of
temperature and β' CD concentration at pH 6

DIAZEPAM/ ρ CD PH 6

Figure VIII

Solubility of ibuprofen as a function of
temperature and β' CD concentration at pH 5

IBUPROFEN/PCD P115

Figure IX

Solubility of hydrochlorothiazide as a function of
temperature and β' CD concentration at pH 5

HYDROCHLOROTHIAZIDE / β CD PH 5

MANUSCRIPT IV

ABSTRACT

The complexation of several drugs with beta cyclodextrins was studied using the Higuchi and Connors solubility method. Solubility isotherms were plotted for each drug at various pH and temperature conditions. Both the non-ionized and the ionized species of diazepam were shown to form an inclusion complex. In the case of ibuprofen and hydrochlorothiazide, only the non-ionized species interacted with beta cyclodextrin. The thermodynamic parameters of complexation were estimated using a computer method based on non-linear regression. The enthalpy of complexation was large and negative in all cases, while the entropy of complexation was found to depend more upon the nature of the drug and the experimental conditions.

Key words: Cyclodextrins, complexation, thermodynamics, ibuprofen, diazepam, hydrochlorothiazide.

PHYSICO-CHEMICAL ASPECTS OF THE COMPLEXATION
OF SOME DRUGS WITH CYCLODEXTRINS

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INTRODUCTION

The three major cyclodextrins consist of six, seven and eight units of glucose, respectively (1). Cyclodextrins readily form inclusion complexes with a wide number of organic compounds. The complexation of drugs with cyclodextrins has been shown to have great potential for the pharmaceutical industry (2). The main advantages of the drug-cyclodextrin complex formation include: a) enhancement of organo-leptic properties (i.e. taste and smell); b) stabilization of the drug; and c) improvement of the solubility, dissolution and bioavailability of drugs. A number of papers have been published by pharmaceutical scientists during the last decade that focused on the preparation, the physical analysis and the formulation of drug/cyclodextrin complexes (3-7). However, the physico-chemical aspects of complexation have received relatively little attention.

The objectives of this paper were: to investigate the effect of pH and temperature on the phase-solubility of three drugs upon complexation with cyclodextrins and to determine the thermodynamic parameters of complexation using a novel computer approach based on a non-linear program.

EXPERIMENTAL SECTION

Reagents and Chemicals- All solvents and reagents were of analytical grade and were used as received.

Hydrochlorothiazide and diazepam were obtained from Sigma Chemical Co. (St. Louis, MO). Ibuprofen was received from Whitehall Laboratories. The cyclodextrin materials used were generously offered by American Maize (Hammond, IN).

Methods- The solubility method used was based on that described by Higuchi and Connors (8). An excess of solid drug was added into screw-capped tubes containing solutions of cyclodextrins at various known concentrations. The tubes were rotated in a waterbath at constant temperature ($\pm 0.05^{\circ}\text{C}$) until saturation was reached. The filtration and dilution steps were conducted in a walk-in oven. The complexation of ibuprofen, diazepam and hydrochlorothiazide were studied at various temperatures from 20°C to 50°C , and at different pH values. The solubility isotherms were obtained by plotting the solubility of the drug as a function of cyclodextrin concentration.

Computer treatment of Data- The thermodynamic parameters of complexation were determined using a computer

method based on non-linear regression. Three programs were used: 1) a datafile; 2) a subroutine; and 3) a JCL program which links the datafile and the subrouitne to the NONLIN software. The datafile included a full set of data consisting of drug concentration, cyclodextrin concentration and temperature at a given pH value and the initial estimates of the thermodynamic parameters. The three major parameters considered were the enthalpy (ΔH) and entropy (ΔS) of complexation, and the heat of solution (ΔH_s). The subroutine program contained the following mathematical equation:

$$[AH]_t = e^{A+B} / (1+e^{A+B}) [CD]_t + e^B \quad (1)$$

where: $[AH]_t$ = total concentration of drug in solution

$[CD]_t$ = total concentration of cyclodextrin

$$A = \Delta H/RT - \Delta S/R$$

$$B = \Delta H_s (T-T_o)/RTT_o - \ln \gamma$$

with: ΔH = enthalpy of complexation

ΔS = entropy of complexation

ΔH_s = heat of solution

R = gas constant

T = absolute temperature ($^{\circ}K$)

T_o = melting point

γ = activity coefficient

This equation was based on the following assumptions: 1) the stoichiometry of the drug/cyclodextrin complex was 1:1; 2)

the change of the free energy of complexation with temperature obeyed the Gibbs equation; 3) the complexing ability of the cyclodextrins was pH independent between pH 2 and pH 9; and 4) the free drug solubility could be described by the general Van't Hoff equation. The output of the NONLIN program used gave the final estimates of the thermodynamic parameters, their standard deviation and their 95% confidence limits.

Statistical Analysis- An analysis of variance (ANOVA) was performed on the slope of the solubility isotherms using a SAS statistical package. Temperature and pH were the two possible sources of variation. A SASGRAPH program was used to plot the solubility of the drug as a function of cyclodextrin concentration and pH. A G3D grid technique was used in all cases.

RESULTS AND DISCUSSION

Determination of pKa- The Benet and Peck approach was used to calculate the apparent pKa of each drug from the intercept data of the solubility isotherms built at a given temperature (9). The solubility C varies as a function of pH according to the following equation:

$$\log (C/C_0 - 1) = \text{pH} - \text{pKa} \quad (2)$$

where C_0 = solubility of the unionized species

The pKa value corresponds to the pH at which the solubility is twice that of the unionized species. The apparent pKa determined from the data available were 4.8, 3.6 and 8.0 for

ibuprofen, diazepam and hydrochlorothiazide respectively. These values are in good agreement with the values available in the literature. In the case of diazepam, which is a weak base, 3.6 is the pKa of the conjugated acid.

Determination of Heat of solution- The heat of solution (ΔH_s) of the free drug represents the intercept of the solubility isotherm.

The Van't Hoff and Hildebrand equations can be used to show the solubility-temperature relationship (10). Although both equations have the same theoretical basis, the plot of the logarithm of the solubility expressed in mole fraction (X) as a function of the reciprocal of the absolute temperature is referred to as the Van't Hoff plot (11,12).

$$\log X = -(\Delta H/2.303 R) (1/T) + C_t \quad (3)$$

The Van't Hoff plots for ibuprofen, diazepam and hydrochlorothiazide are shown in Figures 1, 2 and 3 respectively. Each straight line represents a set of data at a given pH value. The slope of the linear plots varied with pH indicating that the heat absorbed by the system during dissolution varied with the extent of ionization.

Table I shows the values for the heat of solution of the three drugs determined by both the manual method using equation 2 and linear regression and the computer method using non-linear regression. The advantage of the computer approach is that each parameter is estimated with a 95%

confidence interval. An excellent correlation was observed between the results obtained from the two different methods for all three drugs tested. In the case of ibuprofen and hydrochlorothiazide, which are both weak acids, an increase of pH resulted in lower heat of solution because of the increased ionization of the drug. The lack of variation in the H_s with pH for hydrochlorothiazide can be correlated to the almost parallel lines observed in Figure 3. For diazepam, the unionized species at pH 6 required more energy to go into solution. In all cases, the solubility process was endothermic as indicated by the positive values of heat of solution.

Effect of pH on complexation- The solubility isotherms of the complexation of the three drugs with β cyclodextrin at various pH values and constant temperature are presented in Figures 4, 5 and 6. As the pH increased, the intercept increased for ibuprofen and hydrochlorothiazide and decreased for diazepam due to the ionization of the drug. The solubility plots obtained for diazepam (Figure 5) indicate that there is an increase of the slope as the pH decreased. Since diazepam is completely ionized at pH 2, the observed increase of the slope would correspond to the interaction of β cyclodextrin with the ionized species. Without such interaction, the pH 2 line should have been parallel to those obtained at the other pH values. However, in the case of ibuprofen and hydrochlorothiazide, the SAS treatment showed that there was no significant difference in

the slope as a function of pH i.e. for these two drugs the ionized species does not interact significantly with β cyclodextrin.

In order to rationalize the differences in complexation behavior, one has to consider the basic mechanism of inclusion complex formation. The cavity of cyclodextrin is an hydrophobic environment that can host an hydrophobic molecule. It seems that β cyclodextrin can fit a group similar to the benzene ring, which is one of the groups present in diazepam and phenytoin that has previously been shown to form a complex at the ionized state. For these two molecules the complexing group may remain mostly unaffected by the ionization and therefore the ionized species is still able to form a complex with cyclodextrin. Ibuprofen and hydrochlorothiazide do not form such a complex after ionization and the slope of the solubility isotherms is pH independent. It is possible that the ionization of both drugs greatly affects the polarity or the hydrophobicity of the complexing group, thus preventing the formation of a complex at higher pH values.

Effect of temperature on complexation- The solubility study was also conducted at different temperatures for each of the three drugs studied. Figure 7 shows an example of the plots obtained at each pH value. The analysis of variance using SAS did not show any significant difference of the slopes tested, which means that the solubilization power was essentially temperature-independent.

Three-dimensional plots obtained at given pH values using SASGRAPH are shown in Figures 8, 9 and 10 for ibuprofen, hydrochlorothiazide and diazepam, respectively.

Determination of thermodynamic parameters- The classical method to determine thermodynamic parameters of complexation is based on the temperature dependence of the complexation constant. However, this method based on two sequential steps of linearization of data, does not allow a precise determination of the enthalpy and entropy of complexation, which can make difficult further comparisons. The computer method developed in this study utilized datafiles that contained a set of solubility data at one pH value similar to that plotted in Figures 8 to 10.

Tables II, III and IV summarize the results of the treatment of the solubility data for the three drugs. The thermodynamic parameters of complexation can be used as a basis for discussion of the mechanism of complexation. The driving forces of complexation have long been a subject of controversy (13,14). The different possibilities proposed include hydrophobic interaction, hydrogen bonding, Van der Waals interaction, release of enthalpy rich water molecules and release of conformational strain (15). The contribution of each of these factors depends upon the nature of the drug. The overall thermodynamic parameters, namely enthalpy (ΔH) and entropy (ΔS) are related to the apparent free energy (ΔG) by the Gibbs equation:

$$\Delta G = \Delta H - T \Delta S \quad (4)$$

The free energy can be considered as the sum of the energy changes due to each of the factors listed earlier. Therefore, the values given in Tables III to IV can be seen as overall thermodynamic parameters, that indicate which factors may be predominant in the mechanism of complexation.

In the case of ibuprofen, the apparent enthalpy of complexation is negative at all pH values indicating that the system is releasing energy upon complexation. The extent of the energy released decreases as the drug becomes more ionized. There is no definite trend in the change in enthalpy from pH 2 to pH 5. However, the large negative enthalpies probably indicate a strong involvement of dipoles and Wan der Waals interaction during complexation. Also this loss of heat upon complexation could correspond to the energy released by the enthalpy rich water molecules trapped within the cyclodextrin cavity. The entropy factor is positive at all pH values, showing that the order of the systems decreases upon complexation. At pH 6, the entropy increase is particularly significant despite the high standard deviation. Ibuprofen is quite ionized at pH 6 and it is likely that some water molecules of solvation are still in contact with the drug molecule. If one considers that the change in entropy corresponds to the difference in hydrogen bonds before and after complexation, one could suggest that the larger overall balance is due to the smaller number of hydrogen bonds broken.

The thermodynamic parameters of complexation for the diazepam/ β cyclodextrin interaction are presented in Table III. The enthalpy of complexation is again negative at all pH values. As pH decreases and drug ionization increases, the absolute value of ΔH decreases. This lower value of ΔH indicates a weaker binding and a lower loss of heat upon complexation. It has also been proposed that the complexation of an ionized species would occur on a more random fashion than for the non-ionized species (16). The entropy value is positive and increases as the pH is increased. As for ibuprofen, the entropy can be correlated to the forming and breaking of hydrogen bonds upon complexation. The ionized form of diazepam probably keeps a water solvation in the surroundings and gains hydrogen bonding without losing such a great amount as in pH 6. The overall result is a larger entropy upon complexation. Also, as the pH decreases the water molecules are more tightly bound to the drug molecule and therefore are only slowly removed, partially explaining the high entropy values.

The results shown in Figure IV for the complexation of hydrochlorothiazide with β cyclodextrin are less dramatic but possibly easier to interpret. Hydrochlorothiazide is a weak acid with a pK_a around 8.0. It seems that the ionization of the drug has very little effect on the thermodynamic parameters of complexation. This result also confirms the fact that hydrochlorothiazide does not form a complex with β cyclodextrin in its ionized form. The

negative values of enthalpy testifies again of the strong involvement of the enthalpy rich water during complexation. The large negative values of entropy suggest that the system becomes more ordered upon complexation.

SUMMARY

From the results for the complexation of three drugs with cyclodextrins, it appears that the main driving forces of complexation reside in the release of energy of the water molecules entrapped in the cyclodextrin cavity. Although the negative enthalpy value was a common point between the drugs tested, the extent of the pH effect varied from one molecule to another. The entropy is probably more related to the solvent participation in the complexation phenomenon. The consideration of the interaction between water and non-polar drug, water and cyclodextrin before complexation as well as the breakage and establishment of hydrogen bonds, allowed for a description of the complexation mechanism for each drug. However, this drug/cyclodextrin interaction depended on the nature of the drug and the pH of the solution. The practical biopharmaceutical implications of the results can be quite significant. A drug /cyclodextrin complex dosage form either for parenteral or oral administration, would encounter various pH conditions in-vivo. The therapeutic effect might well be affected by the mechanism of complexation. Further work is still needed to fully explain

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SUMMARY

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the exact mechanism of drug/cyclodextrin complex formation and to eventually optimize complex formulation.

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TABLE I: Heat of solution of the drugs at various pH values

DRUG	pH	ΔH_s (linear) (cal.mol ⁻¹)	ΔH_s (non-linear) (cal.mol ⁻¹)
IBUPROFEN	2	11,158.8	8,718.9 $\pm$ 1,266.2
	4	10,534.1	9,447.9 $\pm$ 1,223.0
	5	6,287.5	6,207.5 $\pm$ 375.7
	6	4,685.9	4,157.7 $\pm$ 385.8
DIAZEPAM	2	311.2	1,246.8 $\pm$ 543.2
	3	1,565.2	1,459.9 $\pm$ 289.6
	4	3,724.9	5,368.5 $\pm$ 716.0
	6	5,381.4	6,509.3 $\pm$ 565.4
HYDROCHLORO- THIAZIDE	5	10,392.2	9,420.3 $\pm$ 327.4
	8	9,756.2	10,313.4 $\pm$ 332.2
	9	8,808.9	9,157.6 $\pm$ 330.3

TABLE II: Ibuprofen/ β cyclodextrin complexation
thermodynamic parameters

pH	r	ΔH (cal.mol ⁻¹)	std. dev.	ΔS (e.u.)	std. dev.
2	0.999	-6,925.5	324.9	+3.49	1.06
4	0.999	-7,797.9	321.7	+0.88	1.05
5	0.999	-7,045.5	300.0	+0.72	0.97
6	0.991	-4,036.4	1,246.6	+8.15	4.06

TABLE III: Diazepam/ β cyclodextrin complexation
thermodynamic parameters

pH	r	ΔH (cal.mol ⁻¹)	std. dev.	ΔS	std. dev. (e.u.)
2	0.960	-51.4	1502.4	+16.63	4.89
3	0.998	-797.9	145.7	+16.50	0.47
4	0.998	-4,070.7	184.2	+ 5.22	0.60
6	0.999	-4,416.4	104.3	+ 4.44	0.34

TABLE IV: Hydrochlorothiazide/ β cyclodextrin complexation
thermodynamic parameters

pH	r	ΔH (cal.mol ⁻¹)	std. dev.	ΔS	std. dev. (e.u.)
5	0.997	-9,642.3	432.3	+ 14.7	1.4
8	0.997	-9,322.1	574.1	+ 14.1	1.9
9	0.996	-10,012.2	715.9	+ 16.7	2.3

Figure 1
Van't Hoff plots for ibuprofen

Figure 2
Van't Hoff plots for diazepam

Figure 3
Van't Hoff plots for hydrochlorothiazide

Figure 4

Effect of pH on the solubility isotherms of
ibuprofen at 50°C

Figure 5
Effect of pH on the solubility isotherms of
diazepam at 30°C

Figure 6

Effect of pH on the solubility isotherms of
hydrochlorothiazide at 40°C

COMPLEXATION OF HCTZ WITH β CD AT 40°C

Figure 7

Effect of temperature on the solubility isotherms of
diazepam at pH 4

Figure 8

3D plot of the solubility of ibuprofen at pH 5

IBUPROFEN/ ρ CD PH 5

Figure 9
3D plot of the solubility of HCTZ at pH 8

HYDROCHLOROTHIAZIDE / λ_{CD} PH 8

Figure 10

3D plot of the solubility of diazepam at pH 6

DIA. TEM/3 CD. FIG.

MANUSCRIPT V

POTENTIAL USE OF BETA CYCLODEXTRIN
AS AN ADDITIVE IN TABLET FORMULATION

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INTRODUCTION

The dissolution of poorly soluble drugs can be the major limitation in the design of suitable solid dosage forms (1). The absorption rate is usually much higher than the dissolution rate in the gastro-intestinal tract. Therefore an increase in the dissolution properties for many drugs is likely to lead to higher bioavailability (2). Much research has already been done in the field of pharmaceutics to improve the dissolution of poorly absorbed drugs (3,4). Different techniques have been proposed such as micronization (5,6), grinding (7) and preparation of solid dispersions (8,9,10). The term solid dispersion refers to the dispersion of an active drug in an inert carrier or matrix (11), such as polyethylene glycol (PEG), urea, mannitol or polyvinylpyrrolidone (PVP). The complexation of drugs with beta cyclodextrin is a particular case of solid dispersion, in which the drug is actually encapsulated on a

molecular basis in an hydrophilic oligosaccharide composed of seven units of glucose (12).

Cyclodextrins have been shown to have the ability to form inclusion complexes with drugs, which exist both in solution and in the solid state. This is the basis of the preparation of a solid dosage form containing a drug/cyclodextrin complex as the active ingredient. The dry method consists of a mechanical mixing of the drug with cyclodextrins. Different ratios of the two components and various methods of mixing can be used (13). Such formulations have been successfully prepared using grinding and roll-mixing . The advantages of the dry method include simplicity, one-step process and exact knowledge of the blend content. The second or wet method consists of a two-step process. The first step is the formation of the drug/cyclodextrin complex in solution. Drug and cyclodextrin are solubilized in a solvent according to a definite ratio and the solution is shaken until equilibrium is reached (14). The second step is the removal of the solvent. Different techniques have been proposed to prepare the dry complex, such as solvent evaporation (15), kneading, spray-drying (12) and freeze-drying (16). Upon drying, the complex obtained is usually amorphous (17), with two specific properties: a smaller particle size and a reduced crystal growth, due to the fast removal of water molecules (18). It has been well reported that amorphous forms have the largest energy and exhibit faster dissolution. Rarely

in the literature are the dissolution profiles of drug/cyclodextrin complexes compared with those of the pure drug subjected to the same process (19). It is therefore often difficult to interpret the data, that compare dissolution rate of an amorphous form with a crystalline form.

Two main criteria should be taken into consideration in the development of a solid dosage form of drug/cyclodextrin complex: the stoichiometry of the complex and the degree of complexation, that can be evaluated from the slope of the phase-solubility diagram. The stoichiometry of the complex can be determined using different techniques such as Job's plots, titrations or physical analysis by X-Ray crystallography, Infra-Red and Differential Scanning Calorimetry. T. Higuchi also determined the stoichiometry of complexes from the plateau region observed in the phase-diagram solubility of certain insoluble complexes (20). The slope of the solubility isotherms is also an useful indicator of the extent of complexation. It represents a ratio of the respective amounts of drug and cyclodextrin actually entering the complex. A slope higher than one suggests the formation of a $1:x$ complex ($x>1$). A slope lower than the unity can correspond either to a $1:1$ complex with low affinity between drug and cyclodextrin or to the formation of a $y:1$ ($y<1$). For such a complex, the solution ready for drying does not contain exclusively the drug:cyclodextrin inclusion compound. There is also in

solution a substantial amount of free drug and free cyclodextrin. An intermediate step of filtration or purification should then be used to ensure the complete isolation of the complex. Finally, the advantage of the precipitation method is the higher purity of the product obtained.

The drug considered in this study is phenytoin, a widely used anticonvulsant. Phenytoin has a low aqueous solubility and follows non-linear pharmacokinetics (21,22). The complexation of phenytoin with cyclodextrin is expected to result in higher bioavailability and decrease in the intra- and inter-variability of the absorption in patients.

The objectives of the study are to investigate the potential use of beta cyclodextrin as dissolution enhancer in mechanical mixtures of phenytoin and cyclodextrin. Also, the study focusses on the dissolution profiles of direct compression tablets, prepared using various drug/cyclodextrin ratios and different methods of mixing.

EXPERIMENTAL

Materials

Phenytoin was obtained from Sigma Chemicals and used as received. The cyclodextrin materials were a generous gift of American Maize. Avicel PH101, Primojel, magnesium stearate and sodium bicarbonate were all of USP grade suitable for use in pharmaceutical research.

Methods

Formulations

Several formulations were prepared. Tables 1 and 2 show the amount of each ingredient (% w/w) in the different blends. The percentage of cyclodextrin in Table 1 corresponds to 1:0, 1:1 and 1:2 as drug:cyclodextrin ratio (% w/w). In Table 2, a 1:1 ratio was used for all the formulations, which contain various percentages of sodium bicarbonate. This 1:1 stoichiometric ratio has been shown to be that of the complex formed in solution. Blends of 25 grams were prepared using a Turbula Rapid Blender, in three steps:

1. Mix phenytoin and cyclodextrin for ten minutes.
2. Add Avicel pH 101 and mix for ten minutes.
3. Add magnesium stearate and mix for five minutes.

The formulation variables considered were the order of mixing and the method of mixing, as described in Table 3. The blends were then compacted on a model F single-punch Stokes tablet press. The target specifications of the tablets manufactured are listed in Table 4.

Evaluation of tablets

All the tablets were tested according to USP standards. The tablets from all batches did meet the desired specifications. Dissolution studies were conducted using the USP paddle method, in pH 7.4 phosphate buffer and distilled water. Samples were taken manually at 15, 30 and

60 minutes in triplicate. The medium was replaced after each sample withdrawal and final concentrations were computed by taking into account medium replacement. Phenytoin concentrations were determined using a Hewlett-Packard diode-array ultra-violet spectrophotometer. The standard calibration curve obtained at 222 nm obeyed the Beer's law.

RESULTS AND DISCUSSION

Different models have been developed to describe the dissolution of drugs. The following equation shows the relationship between dissolution rate and some critical parameters using the simplest theory:

$$dM/dt = K S (C_s - C) \quad (1)$$

where dM/dt is the dissolution rate

K is the ratio of diffusion coefficient
and thickness of the stationary layer

S is the effective surface area

C_s is the saturation solubility of the drug
and C is the drug concentration at time t

This equation shows that both particle size and saturation solubility directly affect the dissolution rate. For instance, the effect of micronization of the drug on dissolution rate can be readily explained from this empirical equation.

Figure 1 shows the dissolution profiles of phenytoin from tablets containing different amounts of cyclodextrin.

As the ratio of drug:cyclodextrin increases, the dissolution rates increases, suggesting that cyclodextrin promotes the transport of the drug from the dosage form into the dissolution medium. Cyclodextrin may behave either as an extra-granular disintegrating agent or as an in-situ solubilization agent. The larger the amount of cyclodextrin, the higher the probability for the drug molecules to interact with cyclodextrin. It can be mentioned that the increase in dissolution rate, although it was statistically significant, was not dramatic.

Figure 2 shows the effect of grinding on the dissolution rates of phenytoin. Grinding resulted in a significant increase in dissolution, which can be explained by two different factors. Grinding is a process mainly used to decrease particle size of certain drugs. It has been shown that ground phenytoin formulations exhibited higher dissolution characteristics and better bioavailability (7,23). It is thus likely that the increase observed in Figure 2 corresponds to an increase of the specific surface area. Due to the intimate character of grinding, it is also possible that the tortuosity of the system is changed, with a change in the diffusion properties of the drug molecules from the tablet into the bulk (18). This effect would also explain the results shown in Figure 3, where the dissolution rate of the tablets made using method 2 is significantly lower than that observed using the first method. The lack

of interaction between the drug and cyclodextrin during mixing may be related to the negative effect on dissolution.

Sodium bicarbonate has the property to change the microenvironment around the dissolution particles and to promote dissolution of acidic drugs. The dissolution profiles of the tablets made using various amounts of sodium bicarbonate are shown in Figure 4 and 5. The two dissolution media used were pH 7.4 phosphate buffer and distilled water. A significant increase was observed in Figure 4, that shows that sodium bicarbonate affects the pH of the dissolution medium, thus resulting in higher concentrations. However, the increase does not appear to be very extensive. Figure 5 confirms that sodium bicarbonate does not have a dramatic effect on the dissolution rates of phenytoin. The data overlapped after 45 minutes, showing no trend or difference from one formulation to another.

SUMMARY

From the results of this study, it appears that beta cyclodextrin does enhance the dissolution of phenytoin. However, the relatively small extent of the increase in dissolution rate is unlikely to justify the use of this approach simply to increase dissolution rate, although, of course, cyclodextrin has a clearly identified role in increasing equilibrium solubility. Grinding actually seemed to have been the main factor, and can be compared to freeze-drying as an additional formulation process, that can

interfere with the basic effect of cyclodextrin complexation in drug dissolution. A physical analysis would allow a better establishment of the crystal nature on a qualitative basis.

In conclusion, the use of beta cyclodextrin as a non-complexed additive in direct compression formulation seems for phenytoin to be of limited value. A more positive effect might well be observed using wet granulation, in which water or other solvents would allow the formation of the inclusion complex. Cyclodextrin could then play the role of intra-granular disintegrating agent and is likely to promote a larger increase in dissolution rate. Also preformulation data such as solubility studies, degree of interaction and stoichiometry determination should be the basis of the development of drug:cyclodextrin complexation.

ACKNOWLEDGEMENTS

The authors thank American Maize for the generous supply of the cyclodextrin materials.

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Table 1: Phenytoin: β cyclodextrin formulations (% w/w)

	drug: β CD ratio		
	1:0	1:1	1:2
Phenytoin	8	8	8
β cyclodextrin	0	36	72
Avicel pH 101	89	53	17
Primojel	2	2	2
Magnesium stearate	1	1	1

Table 2: Phenytoin:β cyclodextrin formulations
containing sodium bicarbonate (% w/w)

	Batch #		
	Na12	Na24	Na36
Phenytoin	8	8	8
β cyclodextrin	36	36	36
Avicel pH 101	41	29	17
Primojel	2	2	2
Magnesium stearate	1	1	1
Sodium bicarbonate	12	24	36

Table 3: Process variables

Mixing process	Order of mixing	Method
<hr/>		
regular mixing	(phenytoin + cyclodextrin) + Avicel	1
	(phenytoin + Avicel) + Cyclodextrin	2
grinding	(phenytoin + cyclodextrin) + Avicel	1

Table 4: Tablet specifications

Parameter	Unit	Specification
-----------	------	---------------

Weight	(mg)	325 ± 15
Hardness	(Kg)	4 - 7
Friability	(%)	LT 0.8
Disintegration time(s)		LT 180
Diameter	(inch)	0.378 ± 0.001
Thickness	(inch)	0.205 0.005

Figure 1

Dissolution profiles of phenytoin at 37°C in water
Effect of drug:CD ratio

Figure 2
Dissolution profiles of phenytoin at 37° in water
Effect of mixing process

Figure 3
Dissolution profiles of phenytoin at 37°C in water
Effect of order of mixing

Figure 4
Dissolution profiles of phenytoin at 37°C
Effect of dissolution medium

Figure 5

Dissolution profiles of phenytoin at 37°C and pH 7.4
Effect of sodium bicarbonate

DISSOLUTION PROFILES AT 37°C AND pH 7.4

the drugs studied, thus preventing any interaction of the ionized species with cyclodextrin.

A novel computerized method of determination of thermodynamic parameters of the complexing group was developed and validated. The method allowed for a precise determination of enthalpy and entropy of complexation with known standard deviation. A large negative enthalpy was observed for all four drugs, suggesting a rather enthalpy controlled mechanism. This result shows the involvement of the enthalpy-rich water molecules trapped in the cyclodextrin cavity during complexation. Strong dipole interactions as well as Van der Waals forces are likely to accompany the complexation process. Classical hydrophobic interaction does not seem to play a major role. The results for the entropy of complexation indicate both the importance of the solvent and the differences in complexation behavior of the different drugs. Water seems to play a major role in the complexation phenomenon due to its ability to form hydrogen bonding. According to the nature and the polarity of the guest molecule, the extent of hydrogen bonding might vary and thus affect the stability of the complex.

The new chemically modified beta cyclodextrin was shown to have a great potential for the pharmaceutical industry. The solubilization power observed from the solubility data was basically identical to that obtained with the natural beta cyclodextrin. The total concentration reached in the case of phenytoin and ibuprofen could very well lead to new

CONCLUSIONS AND SUGGESTIONS FOR FUTURE WORK

CONCLUSIONS

The solubility method has been shown to be an excellent practical mean of studying the complexation of drugs with cyclodextrins. From the solubility data obtained, a linear relationship was observed between the drug and the cyclodextrin concentrations. The general theory of "hand and glove" or "lock and key" applied for the four drugs studied. Beta cyclodextrin was shown to have the best complexing ability, suggesting that the steric factors are likely to control the complexation phenomenon. This result was also confirmed from the comparative study of the natural and the chemically modified beta cyclodextrin. A change in the external structure of the host molecule did not affect the solubilization power. However, the steric factors are not sufficient to explain the differences from one drug to another or from one degree of ionization to another. For two drugs, ibuprofen and hydrochlorothiazide, the ionized species have been shown not to interact with beta cyclodextrin, whereas the ionized form of phenytoin and diazepam had the ability to form a complex. These differences in behavior might well be due to the structure of the drugs. The ionization seemed to have an effect on the hydrophobic character of the complexing group of two of

dosage forms, with improved bioavailability or better organo-leptic properties.

The formulation of phenytoin with cyclodextrin using a dry method without the intervention of solvents, did not show a great improvement in dissolution profiles. This part of the study pointed out the importance of water or other solvents to ensure an efficient interaction between drug and cyclodextrin.

SUGGESTIONS FOR FUTURE WORK

The physico-chemical aspects of complexation still deserve and necessitate a great amount of research before a clear elucidation of the complexation process can call for a full development of this new formulation tool. The main areas that can be mentioned are:

1. To investigate the thermodynamics of a series of drug analogs, and to compare the parameters of complexation. The computer method developed in this thesis could be valuable in the comparison process. A specific of the effect of particular substituents upon complexation could be identified.
2. To carry a full study in hydro-alcoholic mixtures. The change of polarity of the solvent would affect the stability of the complex and the thermodynamics of the mechanism. An

optimization of the solvent polarity could lead to an improved solubilization of drugs.

3. To evaluate the potential of complexation applications of different cyclodextrin derivatives. Two classes of new compounds could be evaluated. Hydrophilic derivatives with various structures, substituents and degree of substitution could be used to increase optimally the solubility of poorly water-soluble drugs. Also a new class of hydrophobic derivatives could be investigated for their potential use as carriers in sustained release of soluble drugs.

4. To investigate the use of cyclodextrin on the dissolution of tablets prepared by wet granulation. A possible correlation between preformulation data formulation performance could be then established.

SECTION III

APPENDIX A

RELATIONSHIP BETWEEN COMPRESSION PROFILE
AND PHYSICAL PROPERTIES OF A
LITHIUM CARBONATE FORMULATION
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ABSTRACT

Relationship between the compression profile and physical properties of a directly compressed lithium carbonate tablet formulation was examined using an instrumented tablet press. The measured physical parameters included hardness, disintegration time, and dissolution. A linear correlation was observed

X To whom all inquiries should be directed.

between the compression force, 4000 to 7020 lbs. and tablet hardness, 5 to 9 Kp. The disintegration time, 80 to 100 seconds, did not significantly change with the increasing compression force. Dissolution of lithium carbonate tablets complied with the USP limits at all compression forces. A dissolution rate maximum related to compression force was observed. The overall results indicate that the processing parameter, compression force, affects the physical properties of the tablet formulation.

INTRODUCTION

The compression force is one of the critical physical factors which affects the dissolution and disintegration time and hardness of tablet dosage form (1-8). Several investigators (9) have examined methods to study relationship between the applied compression force and tablet properties. The purpose of this investigation was to characterize a relationship between the compression force and physical properties of a directly compressed lithium carbonate formulation. Once the correlation is established, the compression force measurement can be used to monitor the physical properties such as hardness and dissolution of the active drug.

MATERIALS

A directly compressed lithium carbonate formulation included the following ingredients: lithium carbonate USP, corn starch USP, calcium phosphate dibasic USP, polyethylene glycol USP, magnesium stearate USP, and sodium lauryl sulfate USP.

METHODS

Two kilograms of direct compressible blend was prepared by mixing 60% of the formula lithium carbonate, corn starch, calcium phosphate, dibasic; and polyethylene glycol in a laboratory V-blender for 20 minutes. A sufficient quantity of lubricant mix containing the remaining materials was added and the mixing was continued for an additional 10 minutes. The particle size of the blend was determined using a sieve analysis, bulk and tapped densities were measured using a JEL apparatus¹. The blend was compressed using a rotary tablet press with a standard round tooling of 3/8 inch diameter and the compression force was measured using strain

1 J. Engelsmann, AB., Ludwigshafen, West Germany.

guages² and a high speed analog recorder³. The physical properties of tablets were determined as follows: tablet hardness was measured with a Schleuniger⁴ hardness tester and disintegration⁵ and dissolution⁶ tests were performed according to the USP procedures. Lithium carbonate analysis was performed using an atomic absorption⁷ spectrophotometric analysis and products complied with all USP requirements.

RESULTS AND DISCUSSION

Table 1 shows the physical properties of the major ingredients of the formulation. The percentage compressibility was computed as follows:

$$\% \text{ compressibility} = (D_t - D_b) / D_t$$

where D_t and D_b are tapped and bulk densities, respectively.

2 Manesty B3B tablet press, Manesty Machine Ltd., Liverpool, England.

3 Gould 2200 Recorder, Gould Inc., Cleveland, OH USA.

4 Schleuniger Hardness Tester, Vector Corp., Marion, IA USA.

5 Tablet Disintegration Tester, Van Kel Industries, Chatham, NJ USA.

6 Dissolution Apparatus, Hanson Research Corporation, Northridge, CA USA

7 Instrumentation Laboratory AA Spectrophotometer, Instrumentation Laboratory, Inc., Lexington, MA USA.

Lithium carbonate, which showed poor flow properties represented 60% w/w of the formulation. The three ingredients listed in Table 1 accounted for more than 95% w/w of the granulation. The particle size distribution of the different ingredients used in this formulation is shown in Figure 1. Lithium carbonate exists as a very fine powder with 90% w/w less than 44 microns. The plot also shows that the final blend exhibited a poor particle size distribution with 90% w/w of the blend containing particles less than 74 microns.

A typical compression profile of the lithium carbonate formulation is shown in Figure 2. The highest peak represents the maximum compression force developed during tableting. In this study, the maximum compression force varied from 4000 to 7020 lbs. Below 4000 lbs compression force, the tablet friability exceeded a 1% limit and above 7020 lbs, tablet capping was observed.

Relationship between compression force and hardness of lithium carbonate tablets is represented in Figure 3. The results show that the compression force exhibits a linear relationship with hardness ($r^2 > 0.99$), leading to an acceptable product in

terms of compressibility. The compression force and hardness ranges are adequate for incorporating these values in scale up manufacturing of the product.

Disintegration time was measured using the USP procedure. All tablets disintegrated within a time range from 80 to 100 seconds. No significant differences were observed in the disintegration time from one compression force to another.

Dissolution of lithium carbonate tablets was examined using the USP XXI basket procedure. The dissolution profiles at different compression forces are shown in Figure 4. This formulation exhibits the dissolution compression force relationship in which the initial dissolution at 15 minutes is faster to a maximum as the compression force is increased and then further increases in the compression force slow the initial dissolution of lithium carbonate. These results correlate extremely well with those obtained by Smith et. al (6) on dissolution of tablets prepared by a wet granulation of lithium carbonate. These investigators studied the dissolution at 20 minutes and tablet hardness relationship of a lithium carbonate formulation containing lithium carbonate, lactose, cornstarch and magnesium stearate. In their

study, the tablet hardness was increased using higher compression force. The dissolution at 20 minutes showed two maxima as the hardness of the tablets were increased.

Levy et. al (5) showed that the initial dissolution rates of salicylic acid tablets increased when the tableting compression force was increased. This increase in dissolution of salicylic acid was attributed to fracturing of the drug particles at higher force yielding smaller particles with increased surface area. In case of the directly compressed lithium carbonate, initial increase in the compress force probably results to fracture of lithium carbonate and subsequent increase in the compression force leads to bonding and or the fusion. The fracture and bonding mechanism is postulated to increase and decrease the apparent surface area of lithium carbonate. This increase or decrease, therefore, enhances or retards the initial dissolution flux of lithium carbonate. The thermodynamic solubility governing mechanism, independent of the surface area, controls the amount of drug dissolved after 45 minutes as indicated by a constant plateau value. It is possible that such fracture and bonding of lithium carbonate can yield a

dissolution rate maximum.

CONCLUSION

Results of this study show that the compression force is directly correlated to the hardness of lithium carbonate tablets. The results of initial dissolution rate show a maximum when the compression force is increased. It is suggested that the dissolution behavior is caused by the fracture and bonding of lithium carbonate. The method described in correlating the compression force with physical properties provides a process monitoring procedure for lithium carbonate tablets.

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TABLE I
Physical Properties of Ingredients

Ingredients	Bulk Density	Tapped Density	% Compressibility
	(g / cc)	(g / cc)	
Lithium Carbonate	0.53	0.80	34
Dicalcium Phosphate	0.92	1.47	37
Corn Starch	0.59	0.78	24
Formulation Blend	0.63	0.90	30

Figure 1
Particle size distribution of the
lithium carbonate blend

FIGURE 1
Particle size distribution

Figure 2
Compression profile

FIGURE 2
Compression profile

Figure 3
Hardness/compression profile

FIGURE 3

Hardness/Compaction profile

Figure 4
Dissolution profiles of lithium carbonate

FIGURE 4

Dissolution profiles of lithium carbonate

APPENDIX B

A Comparative Study of Several Lactose
Products for Direct Compression Tableting

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ABSTRACT

Physical characteristics such as density, flow and compaction properties of four brands of anhydrous lactose were evaluated. The drug capacity of the four products was studied by preparing 10% and 20% hydrochlorothiazide tablets. Plots of the tablet hardness as a function of the compaction force are presented. It was possible to make tablets with acceptable properties with all four products. A lubrication study was conducted by varying the amount of lubricant from 0.2% to 0.4%. The ejection force was plotted as a function of the compaction force. The lubrication profiles showed that an increase from 0.2% to 0.3% of lubricant led to a significant decrease of the ejection force. All tablets were evaluated for hardness, thickness, friability and disintegration. Furthermore, the reworkability was shown to be technically feasible with all products.

INTRODUCTION

It is widely recognized that different brands of a same ingredient used in a tablet formulation can lead to a significant change in the physico-chemical properties and the stability of tablets as well as the in vivo performance of the drug (6,7). The objective of the study was to compare four commercially available anhydrous lactose products. Three of the products considered had the same structure, whereas the fourth product, despite a different structure, could very well be used for similar formulation purposes. The study was limited to one drug and two lots of each product.

MATERIAL AND METHODS

MATERIALS

The anhydrous lactose products were obtained from Sheffield Products¹, Foremost², Dairy Crest³ and De Melkindustrie Veghel⁴. Two lots of each product were used. The drug used in the formulations was hydrochlorothiazide (HCTZ) USP⁵. Magnesium Stearate⁶ was used as a lubricant.

METHODS

BULK/TAP DENSITY.

Bulk density was determined by weighing the amount needed to fill a 50 ml graduated cylinder. Tap density was measured by tapping the material until complete settling. All determinations were done in triplicate.

¹Sheffield Products, Norwich, NY

²Foremost Whey Products, Baraboo, WI

³DeMelkindustrie Veghel, Veghel, The Netherlands

⁴Dairy Crest - Whey Division, Suney, England

⁵Sigma Chemical Company, St. Louis, MO

⁶Fisher Scientific Company, Fair Lawn, NJ

PARTICLE SIZE DISTRIBUTION.

Particle size distribution was determined by shaking a 50 gram sample of material on a Fisher Wheeler Sieve shaker for 20 minutes.

POWDER FLOW.

Powder flow was measured using a Mettler PR 1200 scale connected to a Mettler GA12 chart recorder (1).

PREPARATION OF TABLETS.

Two hundred gram batches of the formulations were blended in a Wab Ambula mixing unit for ten minutes. The blend was further blended for three minutes after the addition of the lubricant.

TABLET COMPACTION.

Tablets were compressed using a Stoke's sixteen station rotary press. The press was instrumented with piezo transducers and interfaced to a Apple II personal computer. The details of methods used in tablet formulation and evaluation have already been published by Rhodes and his research associates (2,3,4,5).

TABLET EVALUATION.

The weight of 20 tablets was determined using a standard analytical balance. The average tablet weight and its standard deviation were computed and expressed in mg.

The thickness of 20 tablets was measured using a standard micrometer. The average tablet thickness and its standard deviation were calculated.

The hardness of 20 tablets was determined using a Erweka hardness tester. The standard deviation was calculated and represented as a vertical bar on the graphs.

The friability of 20 tablets was measured by comparing the initial weight with that after 4 minutes at 100 rpm.

The disintegration time of 6 tablets was evaluated using the apparatus described in the USPINF XX -test #701.

RESULTS AND DISCUSSION.

PHYSICAL CHARACTERISTICS.

From the results of the bulk and tap densities, the coefficient of compressibility was calculated as follows:

$$\% \text{ compressibility} = (D_T - D_B) / D_T$$

where D_T = tap density

D_B = bulk density

Table 1 shows the percent compressibility and the flow rate of the four products. The highest flow rate was observed for the Foremost product, which also showed the lowest % compressibility. A lower flow rate was obtained in the case of the Sheffield product. However, when actually making the tablets, no problem of flow was observed from the hopper to the press table, nor from the table into the die, for any of the products. Tablet weight uniformity was acceptable.

The particle size distribution study showed that the Sheffield and the DMV products had similar properties with a respective percentage underweight of 51% and 41% through the sieve #150. The two other products appeared to be finer materials with 79% for Foremost and 90% for Dairy Crest under the same conditions.

The results of the compaction study are shown in Figure 1, where the hardness of the tablets is plotted as a function of compaction force. For all products, as the compaction force increased, the hardness increased until capping occurred.

DRUG CAPACITY.

Figures 2 and 3 show the hardness of the 10% and 20% HCTZ tablets respectively, as a function of the compression force. The formulation of 10% HCTZ tablets showed acceptable levels for all products. At a 20% level of drug, the hardness of all tablets was lower. The differences observed from product to product in Figures 2 and 3 followed the trend shown in Figure 1.

LUBRICATION.

For each product, blends containing 0.2%, 0.3% and 0.4% of magnesium stearate were prepared and compressed at different compaction forces. Plots of the ejection force as a function of the compaction force has been previously used to evaluate the lubrication profile (8). The lubrication profile of the Sheffield product is thus shown in Figure 4. As the compaction force increased, the ejection force increased, and the lower the percentage of lubricant, the higher the ejection force. An increase from 0.2% to 0.3% showed a significant decrease of the ejection force, whereas an increase from 0.3% to 0.4% did not show any further significant decrease. A comparison of the four products was made at one single compression force of 5 kN. Table 2 shows that the results were similar for all products. It appeared that 0.3% would be a reasonable level of lubricant.

EVALUATION OF TABLETS.

The properties of the tablets made using the four different lactose products are shown in Table 3. All the tablets had a same weight of 350 mg and were compressed at a similar compaction force. Even though some variations were observed, it would have been possible to make tablets with acceptable specifications with all products.

Rework was technically feasible for all four matrices, although the quality of the tablets obtained varied for the different excipients.

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Table 1: Flow Properties

<u>Lactose product</u>	<u>Lot #</u>	<u>% compressibility</u>	<u>flow rate</u> gm./sec.
Sheffield	4NL13	24	10.3 (0.6)
	4NL14	28	11.4 (1.2)
Foremost	1RA527	11	24.2 (1.9)
	2RH431	10	29.0 (1.9)
Dairy Crest	635024	22	23.6 (0.5)
	635025	23	22.6 (1.0)
DMV	A1369	25	23.8 (2.2)
	A0741	24	22.0 (1.1)

^a Average (Standard Deviation) from 3 runs.

Table 2: Lubrication study (Compaction Force = 5 kN)

	<u>Ejection Force</u> (N)		
	<u>Magnesium Stearate</u>		
	<u>0.2%</u>	<u>0.3%</u>	<u>0.4%</u>
Sheffield	390	320	300
Foremost	380	330	310
DMV	430	350	330
Dairy Crest	350	250	240

Table 3: Evaluation of 10% HCTZ tablets.

Lactose product	<u>Thickness</u> inch	<u>Hardness</u> kg	<u>Disintegration time</u> min.	<u>Content uniformity</u> %
Sheffield	.173 (.001) ^a	10.5 (0.5) ^b	10	1.1
Foremost	.177 (.001)	3.9 (0.7)	17	2.4
DMV	.176 (.001)	4.8 (1.3)	6	2.0
Dairy Crest	.177 (.001)	3.5 (0.8)	20	4.4

^aStandard Deviation from 10 tablets.

^bStandard Deviation from 20 tablets.

Table 4: Evaluation of 10% HCTZ tablets after rework.

<u>Lactose product</u>	<u>Thickness</u> inch	<u>Hardness</u> kg	<u>Friability</u> %	<u>Disintegration time</u> min.
Sheffield	.168 (.002) ^a	4.8 (1.3) ^b	0.34	10
Foremost	.165 (.002)	2.1 (0.8)	0.25	20
DMV	.170 (.002)	2.8 (0.7)	0.56	12
Dairy Crest	.165 (.002)	2.0 (0.6)	0.55	20

^aStandard Deviation from 10 tablets

^bStandard Deviation from 20 tablets

FIGURE 1
COMPACTION PROFILES

FIGURE 1
COMPACTION PROFILES

FIGURE 2
DRUG CAPACITY (10% HCTZ)

FIGURE 2
DRUG CAPACITY (10% HCTZ)

FIGURE 3
DRUG CAPACITY (20% HCTZ)

FIGURE 3
DRUG CAPACITY (20% HClZ)

FIGURE 4
LUBRICATION STUDY

FIGURE 4
LUBRICATION STUDY

APPENDIX C

DESCRIPTION OF THE EXPERIMENTAL PROTOCOL

1. Principle of the solubility method
2. Preparation of the buffered solutions of cyclodextrins
3. Preparation of the solubility vials
4. Description of the solubility apparatus
5. Time to reach equilibrium
6. Direction to approach the desired temperature
7. Sampling, filtration and dilution of samples
8. Ultra-Violet analysis of the drugs
9. Plot of solubility isotherm

1. Principle of solubility method

Higuchi and Connors described the solubility method in the 5th chapter of Advanced Analytical Chemistry and Instrumentation. An excess of solid drug is placed in vials containing various known concentrations of the complexing agent. The solubility isotherm is then obtained by plotting the total drug concentration as a function of the total cyclodextrin concentration. Different types of phase-solubility diagrams may be observed. The type observed in this study was in all cases the A-type, in which there is linearity throughout the range of concentrations of the complexing agent.

2. Preparation of the buffered solutions of cyclodextrins

The solutions of cyclodextrins were prepared by dissolving known amounts of solid cyclodextrins in buffered solutions adjusted to the required pH. The buffered solutions were prepared by using monobasic and dibasic phosphate. The four characteristics of the buffer are: nature, pH, concentration and ionic strength.

The buffers made were phosphate buffers. The buffer concentration used was 0.1 mole per liter. The ionic strength was adjusted to 0.3 mole per liter using potassium chloride (KCl). The first step was to make two stock

solutions, one of monobasic phosphate and one of dibasic phosphate. The concentration and the ionic strength of these two solutions were known. The second step was to mix the two stock solutions such that the pH desired would be exactly obtained. All the pH values were also checked at the end of the experiment at room temperature.

3. Preparation of the solubility vials

Test tubes of ten or twenty milliliters were used to carry out the solubility experiments. The excess of drug was approximatively measured so that the solution would be saturated after equilibrium. The amount varied from drug to drug and from one cyclodextrin to another. The solutions were then added to the solid drug in the vial. The tubes were then tightly closed using screw-caps, and Parafilm strips were used to secure a perfect sealing of the vials.

4. Description of the solubility apparatus

The tubes prepared were placed and attached on the rotating unit. The rotating unit was immersed in a waterbath at a constant temperature in the range of 20°C to 50°C ($\pm 0.05^\circ\text{C}$). A cooling unit was also used to ensure a double source of energy thus allowing a better control of temperature. The capacity of the apparatus was about 70 tubes, which corresponded to fourteen clamps each holding up

to five tubes. The tubes were rotated until saturation was reached.

5. Time to reach equilibrium

Two solutions of cyclodextrins, namely 3×10^{-3} M and 12×10^{-3} M were used to evaluate the time needed to reach equilibrium in the case of phenytoin at 20°C and pH 7. Samples were withdrawn and analyzed for drug concentration after intervals of time. The drug concentration was then plotted as a function of time, as shown in Figure 1. Even though the maximum concentration of drug was reached after about one hour, all the results shown in this study were obtained after a rotating time of two to three hours.

6. Direction to approach the desired temperature

In the study of the effect of temperature on the complexation of drugs with cyclodextrins, the same tube containing drug and cyclodextrin solution was brought to temperatures from 20°C to 50°C . There were two ways to change the temperature of the waterbath, one is to go from low to high temperatures and the other from high to low temperatures. In order to evaluate whether the direction to approach the desired temperature would affect the solubility results, a study was performed, in which the same samples were treated according to the two possibilities. As shown

in Table 1, there was no effect on the final drug concentration. In all the experiments, the temperature was then increased from 20°C to 50°C.

7. Sampling, filtration and dilution of samples

Samples of drug/cyclodextrin complex solutions of one to three milliliters were withdrawn from the tubes using a pipette. The samples were then filtered using glasswool, and filtration was shown not to interfere with the final concentration. An aliquot of 500 μ l or one milliliter was then used to dilute the sample before analysis of the drug concentration. All manipulations of the samples were performed at the respective temperature by either working in a cool place (at 20°C) or in a walk-in oven (30°C to 50°C).

8. Ultra-Violet analysis of the drug

The diluted drug samples were analyzed using a diode array ultra-violet spectrophotometer. Standards of the respective drug were prepared by dissolving an exactly amount of pure drug placed in a hundred milliliters volumetric flask in twenty or thirty milliliters of acetonitrile. The volume was adjusted using distilled water. The standards had a concentration from ten to fifty μ g/ml for phenytoin, diazepam and hydrochlorothiazide, and twenty

to hundred ug/ml for ibuprofen. The Beer's law was obeyed in all cases. The calibration curves are shown in Figures 2, 3, 4 and 5 for phenytoin, ibuprofen, diazepam and hydrochlorothiazide, respectively. Cyclodextrins were shown not to interfere with the assay.

9. Plot of solubility isotherm

In the case of phenytoin at pH 7 and 25°C, six tubes were prepared for each cyclodextrin concentration. The protocol was followed as described earlier. The final concentrations were plotted by considering the average value and its standard deviation as a function of cyclodextrin concentration. The results are presented in Figure 6.

Table 1
Direction of approach to equilibrium

T (°C)	[CD] x 10 ³ M					
	0		6		12	
	[Drug] mg/ml					
40	4.5	3.1	18.0	17.3	31.8	31.2
25	2.7	1.6	15.0	14.4	27.4	27.4
10	2.0	1.3	14.4	13.7	26.6	26.5

Figure 1
Time to reach equilibrium

Figure 2
U.V. calibration curve for phenytoin

Figure 3
U.V. calibration curve for ibuprofen

Figure 4
U.V. calibration curve for diazepam

Figure 5

U.V. calibration curve for hydrochlorothiazide

Figure 6
Plot of solubility isotherm for phenytoin

PLOT OF SOLUBILITY ISOTHERM FOR PHENYTOIN

APPENDIX D

SUMMARY OF SOLUBILITY ISOTHERMS

The following section contains the plots of the solubility isotherms obtained for the four drugs considered at different pH and temperature conditions. Some of these plots are already presented in Manuscript I and Manuscript IV. However, it was not possible to include all the data in the manuscripts, and this summary may be useful to review the experimental results.

The plots are presented drug by drug, as follows:

1. Phenytoin

Complexation with β 'CD at pH 5 and pH 8

2. Ibuprofen

Complexation with β CD at pH 5 and 30°C

Complexation with β CD at pH 2, 4, 5 and 6

Complexation with β CD at 20, 30, 40 and 50°C

3. Diazepam

Complexation with β CD at pH 2, 3, 4, and 6

Complexation with β CD at 20, 30, 40, and 50°C

4. Hydrochlorothiazide

Complexation with α , β and CD at pH 5 and 30°C

Complexation with β CD at pH 5, 8 and 9

Complexation with β CD at 20, 30, 40 and 50°C

FIGURE 1
COMPLEXATION OF PHENYTOIN WITH SUBST. β CD AT pH 5

FIGURE 2

FIGURE 3

FIGURE 4
COMPLEXATION OF IBUPROFEN WITH β CD AT pH 2

FIGURE 5

COMPLEXATION OF IBUPROFEN WITH CD AT pH 4

FIGURE 6
COMPLEXATION OF IBUPROFEN WITH CD AT pH 5

FIGURE 7
COMPLEXATION OF IBUPROFEN WITH β CD AT pH 6

FIGURE 8
COMPLEXATION OF IBUPROFEN WITH β CD AT 20°C

FIGURE 9

COMPLEXATION OF IBUPROFEN WITH β CD AT 30°C

FIGURE 10

COMPLEXATION OF IBUPROFEN WITH β CD AT 40°C

FIGURE 11

COMPLEXATION OF IBUPROFEN WITH β CD AT 50°C

FIGURE 12
COMPLEXATION OF DIAZEPAM WITH β -CD AT pH 2

FIGURE 14
COMPLEXATION OF DIAZEPAM WITH β CD AT pH 4

FIGURE 16

FIGURE 17

FIGURE 20
COMPLEXATION OF HCTZ WITH CYCLODEXTRINS

FIGURE 21
COMPLEXATION OF HCTZ WITH β CD AT pH 5

FIGURE 22

COMPLEXATION OF HCTZ WITH β CD AT pH 8

FIGURE 23
COMPLEXATION OF HCTZ WITH β CD AT pH 9

FIGURE 24

FIGURE 25

COMPLEXATION OF HCTZ WITH β CD AT 30°C

FIGURE 26
COMPLEXATION OF HCTZ WITH β CD AT 40°C

FIGURE 27

APPENDIX E

STATISTICAL TREATMENT OF THE DATA

The solubility isotherms obtained in this study correspond to the complexation of several drugs with cyclodextrins, at different pH values and different temperatures. The effect of these two variables on the slope of the plots of drug concentration vs. cyclodextrin concentration was tested for their statistical significance using a SAS program.

The following section describes the datafiles, the Job Control Language (JCL), which was identical in all cases, and the results of SAS treatment of the data. The datafiles are listed for phenytoin, ibuprofen, diazepam and hydrochlorothiazide, respectively. The results listed are those obtained at a 95% confidence level ($\alpha = 5\%$), for the four drugs in the same order as above. The data were also tested at the 1% and 10% levels.

SAS			
DFS	TYPE	PH	GLPGE
1	203	5	0.0777
2	203	7	0.0817
3	203	0	0.1046
4	203	0	0.2670
5	203	10	0.3700
6	203	5	0.0802
7	203	7	0.0868
8	203	8	0.1118
9	203	9	0.2902
10	203	10	0.4118
11	213	5	0.0847
12	213	7	0.0902
13	213	8	0.1249
14	213	0	0.3325
15	213	10	0.2447
16	223	5	0.0851
17	223	7	0.0940
18	223	8	0.1384
19	223	0	0.2657
20	223	10	0.4055

545

GENERAL LINEAR MODELS PROCEDURE

DUNCAN'S MULTIPLE RANGE TEST FOR VARIABLE: SLOPE
 NOTE: THIS TEST CONTROLS THE TYPE I COMPARISONWISE ERROR RATE,
 NOT THE EXPERIMENTWISE ERROR RATE

ALPHA=0.05 DF=12 MSE=0.0017349

NUMBER OF MEANS 2 3 4
 CRITICAL RANGE 0.0572062 0.0600093 0.0618292

MEANS WITH THE SAME LETTER ARE NOT SIGNIFICANTLY DIFFERENT.

MEAN	GROUPING	MEAN	N	TEMP
	A	0.19794	5	303
	A	0.19629	5	303
	A	0.19012	5	293
	A	0.17672	5	313

SAS

GENERAL LINEAR MODELS PROCEDURE

SIMONIS MULTIPLE RANGE TEST FOR VARIABLE: SLOPE
 NOTE: THIS TEST CONTROLS THE TYPE I COMPARISONWISE ERROR RATE
 NOT THE EXPERIMENTWISE ERROR RATE

ALPHA=0.05 DF=12 MSFE=0.017348

NUMBER OF MEANS 2 3 4 5
 CRITICAL RANGE 0.064048 0.0670324 0.0921272 0.0701934

MEANS WITH THE SAME LETTER ARE NOT SIGNIFICANTLY DIFFERENT.

TULCAN	GROUPING	MEAN	N	PH
	A	0.76792	4	10
	B	0.23060	4	9
	C	0.11932	4	8
	D	0.03847	4	7
	E	0.04137	4	5

TABLE 1			
DATE	TIME	PH	SLCPE
1	227	2	0.0330
2	227	3	0.0351
3	227	4	0.0368
4	227	5	0.0325
5	227	6	0.0320
6	227	7	0.0391
7	227	8	0.0325
8	227	9	0.0362
9	227	10	0.0366
10	227	11	0.0397
11	227	12	0.0491
12	227	13	0.0410
13	227	14	0.0417
14	227	15	0.0447
15	227	16	0.0491
16	227	17	0.0476

SAS

GENERAL LINEAR MODEL PROCEDURE

BUNGAN'S MULTIPLE RANGE TEST FOR VARIABLE: SLOPE
 NOTE: THIS TEST CONTROLS THE TYPE I COMPARISONWISE ERROR RATE
 NOT THE EXPERIMENTWISE ERROR RATE

ALPHA=0.05 DFB=9 MSF=0.002424

NUMBER OF MEANS 2 3 4
 CRITICAL RANGE 0.0788072 0.0021244 0.004298

MEANS WITH THE SAME LETTER ARE NOT SIGNIFICANTLY DIFFERENT.

BUNGAN	SE MEANS	MEAN	N	TEST
A		0.15557	4	327
A		0.15305	4	313
A		0.13405	4	303
A		0.09005	4	293

 GENERAL LINEAR MODELS PROCEDURE

DUNCAN'S MULTIPLE RANGE TEST FOR VARIABLE SLOPE
 NOTE: THIS TEST CONTROLS THE TYPE I COMPARISONWISE ERROR RATE,
 NOT THE EXPERIMENTWISE ERROR RATE

ALPHA=0.05 DFE=9 MSSE=0.002424

NUMBER OF MEANS 2 3 4
 CRITICAL RANGE 0.0755272 0.0821244 0.084298

MEANS WITH THE SAME LETTER ARE NOT SIGNIFICANTLY DIFFERENT.

DUNCAN	GROUPING	MEAN	N	SN
	A	0.78803	4	2
	B	0.06815	4	3
	B	0.04437	4	4
	B	0.03933	4	5

TABLE 4

DATE	PRICE	QTY	GLD PR
1	10.1	7	0.653
2	10.1	6	0.740
3	10.1	8	0.713
4	10.1	5	0.754
5	10.1	7	0.693
6	10.1	6	0.700
7	10.1	5	0.713
8	10.1	5	0.783
9	10.1	8	0.777
10	10.1	6	0.731
11	10.1	6	0.733
12	10.1	7	0.737
13	10.1	8	0.773
14	10.1	5	0.703
15	10.1	7	0.633
16	10.1	5	0.731

GENERAL LINEAR MODELS PROCEDURE
 DUNCAN'S MULTIPLE RANGE TEST FOR VARIABLE: SLOPE
 NOTE: THIS TEST CONTROLS THE TYPE I COMPARISONWISE ERROR RATE,
 NOT THE EXPERIMENTWISE ERROR RATE

ALPHA=0.05 DFB=9 MSSE=6.7E-04

NUMBER OF MEANS: 2 3 4
 CRITICAL RANGE: 0.041887 0.0418762 0.0418801

MEANS WITH THE SAME LETTER ARE NOT SIGNIFICANTLY DIFFERENT.

DUNCAN	GROUPING	MEAN	N	TEMP
	A	0.75100	4	727
	A			
	A	0.73775	4	313
	A			
	A	0.73400	4	703
	A			
	B	0.71800	4	297

SAS

GENERAL LINEAR MODELS PROCEDURE

DUNCAN'S MULTIPLE RANGE TEST FOR VARIABLE: SLOPE
 NOTE: THIS TEST CONTROLS THE TYPE I COMPARISONWISE ERROR RATE
 NOT THE EXPERIMENTWISE ERROR RATE

ALPHA=0.05 DF=9 MSE=6.3E-04

NUMBER OF MEANS 2 3 4
 CRITICAL RANGE 0.0401007 0.0419752 0.0430821

MEANS WITH THE SAME LETTER ARE NOT SIGNIFICANTLY DIFFERENT.

DUNCAN	GROUPING	MEAN	N	PH
A		0.76025	4	4
A				
A		0.75875	4	6
B		0.70750	4	2
B				
B		0.70575	4	5

SAS

DBS	TEMP	PH	SLOPE
1	29.3	5	0.240
2	29.3	4	0.216
3	29.3	3	0.242
4	30.3	5	0.207
5	30.3	2	0.216
6	31.3	2	0.235
7	31.3	5	0.247
8	31.3	3	0.247
9	31.3	3	0.235
10	32.3	5	0.262
11	32.3	3	0.257
12	32.3	2	0.270

SAS

GENERAL LINEAR MODEL: PROCEDURE

ROUTINE MULTIPLE RANGE TEST FOR VARIABLE : SLOPE
 NOTE: THIS TEST CONTROLS THE TYPE I COMPARISONWISE ERROR RATE
 AT THE EXPERIMENTWISE ERROR RATE

ALPHA=0.05 DF=6 MS=3.1E-04

NUMBER OF MEANS 2 3 4
 CRITICAL RANGE 0.0418753 0.0430155 0.0439209

VALUES WITH THE SAME LETTER ARE NOT SIGNIFICANTLY DIFFERENT.

MEAN	GROUPING	MEAN	N	TEMP
0.2110	A	0.2110	3	323
0.2110	A	0.2110	3	317
0.2107	A	0.2107	3	293
0.2107	A	0.2107	3	303

GENERAL LINEAR MODEL PROCEDURE

DISJOINT MULTIPLE RANGE TEST FOR VARIABLE: SLOPE
 NOTE: THIS TEST CONTROLS THE TYPE I COMPARISONWISE ERROR RATE
 NOT THE EXPERIMENTWISE ERROR RATE

ALPHA=0.05 DFB 4 MSF=4.4E-04

NUMBER OF MEANS 2

CRITICAL RANGE 1.0361797 0.0374257

MEANS WITH THE SAME LETTER ARE NOT SIGNIFICANTLY DIFFERENT.

COULCAN	TREATING	MEAN	N	DF
	A	0.24575	4	9
	A	0.23400	4	8
	A	0.23400	4	5

APPENDIX F

NONLIN: PROGRAMS AND RESULTS

As described in Manuscript II, the determination of the thermodynamic parameters using NONLIN, requires the use of three different programs: a datafile, a subroutine program and a Job Control Language (JCL).

The following section contains the listing of the programs used for the four drugs considered in this study. An example of datafile is shown for phenytoin, ibuprofen, diazepam and hydrochlorothiazide, respectively. Two different listings of subroutine programs are then presented. The first set of programs was used for the complexation of the non-ionized species of the drug with cyclodextrins. These programs are given in the order: phenytoin, ibuprofen, diazepam and hydrochlorothiazide. Following is the listing of the subroutine program, used in the case of phenytoin and diazepam, for the complexation of the ionized species with cyclodextrins. Finally, an example of JCL program and a listing of a complete output of the NONLIN program are shown.

TABLE 1. $\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^0 p$ and $\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^+ n$ at $\sqrt{s} = 0.5, 1.0, 1.5, 2.0, 3.0, 4.0, 5.0, 6.0, 7.0, 8.0, 9.0, 10.0$ GeV

$\sqrt{s}$ (GeV)	1	2	3	4	5	6	7	8	9	10
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^0 p$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^+ n$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^0 p$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^+ n$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^0 p$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^+ n$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^0 p$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^+ n$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^0 p$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^+ n$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^0 p$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\sigma_{\text{tot}}^{\text{pion}}(p, p) \rightarrow \pi^+ n$ (mb)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00

TABLE 1. (continued)


```

      DEL 'UP1,LEV101,DIB' NOF LINE NUMBERED, USING NOCOM

      SUBROUTINE DFUNC(F,P,UBB,VAL,X,Y,Z,ISPEC,XVEL,Y,B,RES)
      IMPLICIT REAL*16 (A-H,X,F,D-Z)
      DIMENSION ISPEC(1),NUBB(1)
      DOUBLE PRECISION P(1),VAL(1),F,X,UBB(1),Y(1),W(1),XVEL(1)
      H=XX*(1B,0)*0.000001)
      I1=I+NUBB(1)
      I=XVEL(I1)
      A=(F(1)/(1.9B/*1))-(F(2)/1.9B/)
      B=(F(3)*(1-34B,0))/(1.9B/*34B,0*1)-F(4)
      C=EXP(A+B)
      D=L/(L+1)
      E=EXP(B)
10  F=(D*H)+E)*(206.2B/1B,0)*1000000,0
      IF (ISPEC(B),NE,9) RETURN
      END

```

```

E
DATA SET ('DEF,LIJ0101,DDM' NO) LINE NUMBERED= USING NUNUM
E
      SUBROUTINE DFUNC(F,P,CON,VAL,X,I,J,ISPEC,XVEC,Y,W,NUBS)
      IMPLICIT REAL*16 (A-H,K,O-Z)
      DIMENSION ISPEC(1),NUBS(1)
      DOUBLE PRECISION F(1)*VAL(1)+F*X*CON(1),Y(1)*W(1),XVEC(1)
      H=X*(18.0*0.000001)
      I1=1+NUBS(1)
      F=XVEC(I1)
      A=(F(1)/(1.98/*I))-(F(2)/1.98/)
      B=(F(3)*(1-405.0)/(1.98/*405.0*1))-F(4)
      C=EXP(A+B)
      D=C/(C+1)
      E=EXP(B)
10  F=((D*H)+E)*(284./4/18.0)*1000000.0
      IF (ISPEC(8).NE.9) RETURN
      END
END OF DATA

```

```

DATA SET 'URI,UJVI01,DHC' NOT LINE NUMBERED, USING MINUM
E
      SUBROUTINE DFUNC(F,P,CON,VAL,X,1,2,3,4,5,XVEC,Y,W,ALB)
      IMPLICIT REAL*16 (A-H,K,U-Z)
      DIMENSION ISPEC(1),NOBS(1)
      DOUBLE PRECISION P(1),VAL(1),F,X,CON(1),Y(1),W(1),XVEC(1)
      H=X*(18.0*0.000001)
      J=1+H*NOBS(1)
      P=XVEC(1)
      A=(P(1)/(1.98*H))-P(23/1.98)
      B=(P(3)*(1-541.0))/(1.98*541.0*H)-P(4)
      C=EXP(A+B)
      J=J/(C+1)
      E=EXP(B)
10  F=(1*H+E)*(297.73/18.0)*1000000.0
      IF (ISPEC(8),NE,9) RETURN
      END
END OF DATA

```

DATA SET 'URI,LJV101,ION4' NOT LINE NUMBERED, USING NUNUM
E

```

SUBROUTINE DFUNL(F,F,CUN,VAL,X,I,J,ISPEC,XVEC,Y,W,NUBS)
IMPLICIT REAL*16 (A-H,K,Q-Z)
DIMENSION ISPEC(1),NUBS(1)
DOUBLE PRECISION F(1),VAL(1),F,X,CUN(1),Y(1),W(1),XVEC(1)
H=X*(18.0*0.000001)
I1=1+NUBS(1)
I=XVEC(I1)
A=(9714.95/(1.98/*I))-(10.4/1.98/)
B=(10040.4*(I-568.0))/(1.98/*568.0*I)-5.505
C=EXP(A+B)
U=C/(C+1)
E=EXP(B)
G=((I*H)+E)
U=(F(1)/(1.98/*I))-(F(2)/1.98/)
V=(F(3)*(I-568.0))/(1.98/*568.0*I)-F(4)
UU=EXP(U)
VV=EXP(V)
Z=(UU*VV)-(UU*E)
Q=Z/(1+Z)
K=(VV-E)*(1+(UU*VV))
10 F=((Q*H)+G+K)*(252.23/18.0)*1000000.0
IF (ISPEC(8).NE.9) RETURN
END
END OF DATA

```

```

DATA SET 'URI.LJV101.NONLIN' NOT LINE NUMBERED, USING NONUM
E
//LJV101A JOB (LJV101),'MENARD',NOTIFY=LJV101
/*PASSWORD KLBG
// EXEC NONLINPH,REGION=2048K
//DFUNC.SYSPRINT DD DUMMY
//DFUNC.SYSTEM DD DUMMY
//DFUNC.SYSIN DD DSN=URI.LJV101.DM10N,DISP=SHR
//LKED.SYSPRINT DD DUMMY
//DATA.FT05F001 DD DSN=URI.LJV101.DM14,DISP=OLD
//DATA.FT06F001 DD SYSOUT=A,HOLD=YES
END OF DATA

```


1. The estimated net value of the property is \$100,000.00.

2. The estimated net value of the property is \$100,000.00.

3. The estimated net value of the property is \$100,000.00.

4. The estimated net value of the property is \$100,000.00.

5. The estimated net value of the property is \$100,000.00.

6. The estimated net value of the property is \$100,000.00.

7. The estimated net value of the property is \$100,000.00.

8. The estimated net value of the property is \$100,000.00.

9. The estimated net value of the property is \$100,000.00.

10. The estimated net value of the property is \$100,000.00.

11. The estimated net value of the property is \$100,000.00.

12. The estimated net value of the property is \$100,000.00.

13. The estimated net value of the property is \$100,000.00.

14. The estimated net value of the property is \$100,000.00.

15. The estimated net value of the property is \$100,000.00.

16. The estimated net value of the property is \$100,000.00.

17. The estimated net value of the property is \$100,000.00.

18. The estimated net value of the property is \$100,000.00.

19. The estimated net value of the property is \$100,000.00.

20. The estimated net value of the property is \$100,000.00.

Table 1 (Cont.)

σ_1	σ_2	$10\delta_1 \pm \gamma$	$10\delta_2 \pm \gamma$
5.0×10^{-10}	5.0×10^{-10}	19 ± 160	1.00000
5.0×10^{-10}	1.0×10^{-9}	106 ± 660	1.00000
6.0×10^{-10}	1.0×10^{-9}	201 ± 660	1.00000
9.0×10^{-10}	1.0×10^{-9}	276 ± 10	1.00000
1.0×10^{-9}	5.0×10^{-10}	391 ± 660	1.00000
1.0×10^{-9}	1.0×10^{-9}	437 ± 40	1.00000
5.0×10^{-10}	1.0×10^{-9}	259 ± 60	1.00000
5.0×10^{-10}	5.0×10^{-10}	146 ± 70	1.00000
5.0×10^{-10}	5.0×10^{-10}	76 ± 640	1.00000
1.0×10^{-9}	5.0×10^{-10}	483 ± 10	1.00000
1.0×10^{-9}	5.0×10^{-10}	509 ± 80	1.00000
1.0×10^{-9}	5.0×10^{-10}	850 ± 70	1.00000
1.0×10^{-9}	1.0×10^{-9}	56 ± 410	1.00000
5.0×10^{-10}	1.0×10^{-9}	20 ± 110	1.00000
6.0×10^{-10}	1.0×10^{-9}	33 ± 630	1.00000
9.0×10^{-10}	1.0×10^{-9}	508 ± 10	1.00000
1.0×10^{-9}	1.0×10^{-9}	658 ± 90	1.00000
1.0×10^{-9}	1.0×10^{-9}	309 ± 70	1.00000
5.0×10^{-10}	1.0×10^{-9}	119 ± 10	1.00000
5.0×10^{-10}	5.0×10^{-10}	302 ± 250	1.00000
5.0×10^{-10}	5.0×10^{-10}	465 ± 70	1.00000
5.0×10^{-10}	5.0×10^{-10}	668 ± 60	1.00000
1.0×10^{-9}	5.0×10^{-10}	387 ± 560	1.00000
5.0×10^{-10}	5.0×10^{-10}	1034 ± 60	1.00000

TABLE 12

P 1		P 2		P 3		P 4	
Q 1000	P 1000	Q 1000	P 1000	Q 1000	P 1000	Q 1000	P 1000
1	0	5.155E+04	10.000	2.0000E+04	5.000	3.000	
2	1	5.0800E+04	1.7071E+04	2448.	4.196		
3	2	5.155E+04	1.7005E+04	4329.	3.444		
4	3	5.098E+04	1.737E+04	1785.	4.124		
5	4	5.1313E+04	9875.	14.397	4.754		
6	5	5.180054E+04	1.0791E+04	7250.	4.511		
7	6	1.077547E+06	1.0508E+04	8780.	4.585		
8	0	1.077547E+06	1.0508E+04	8780.	4.585		
9	1	1.2793.	9405.	10.56	3.835		
10	2	878729.	8361.	6.928	3.137		
11	3	103519.	9258.	10.07	3.736		
12	0	103519.	9258.	10.07	3.736		
13	1	48777.3	8849.	7.921	3.083		
14	2	356719.	9449.	6.058	2.483		
15	3	176347.	8904.	8.593	3.294		
16	0	44777.3	8849.	7.921	3.083		
17	1	12777.7	8888.	7.400	2.643		
18	2	31671.0	8977.	6.896	2.232		
19	3	13369.0	8895.	7.311	2.570		
20	0	13369.0	8895.	7.311	2.570		
21	1	2667.47	9147.	7.753	2.269		
22	2	7081.46	9403.	8.208	1.984		
23	3	33511.77	9276.	7.983	2.122		
24	0	33511.77	9276.	7.983	2.122		
25	1	3709.13	9264.	8.789	1.912		
26	2	7115.10	9856.	9.632	1.711		
27	3	3597.17	9346.	8.737	1.925		
28	0	3677.17	9346.	8.737	1.925		
29	1	1561.05	9753.	9.273	1.786		
30	2	1981.51	9878.	9.822	1.652		
31	3	1735.67	9876.	9.620	1.701		

11	0	1.17500E-05	9.75E-06	9.4E-06	1.48388E+03	1.4701
11	1	8.97100E-05	9.75E-05	9.4E-05	1.49724E+04	1.4848
11	2	4.12000E-04	1.00000E+00	10.004	1.49999E+04	1.5095
11	3	1.71000E-03	1.00000E+00	10.006	1.50000E+04	1.5090
11	0	1.91800E-05	1.00000E+04	10.006	1.50000E+04	1.5090
11	1	1.01000E-04	1.00000E+04	10.023	1.50099E+04	1.5094
11	2	1.24400E-03	1.00100E+03	10.031	1.50111E+04	1.5118
11	3	1.01400E-02	1.00080E+04	10.029	1.50000E+04	1.5041
11	0	1.01400E-02	1.00080E+04	10.029	1.50000E+04	1.5041
11	1	5.41100E-01	1.01000E+04	10.032	1.50099E+04	1.5025
11	2	5.41100E-01	1.01000E+04	10.034	1.50120E+04	1.5009
11	3	2.99900E-01	1.01000E+04	10.045	1.50120E+04	1.5007
11	0	2.99900E-01	1.01000E+04	10.045	1.50120E+04	1.5007
11	1	2.99900E-01	1.01000E+04	10.039	1.50106E+04	1.5000
11	2	2.99900E-01	1.01000E+04	10.052	1.50140E+04	1.4972
11	3	2.99900E-01	1.01000E+04	10.053	1.50140E+04	1.4971
11	0	2.99900E-01	1.01000E+04	10.053	1.50140E+04	1.4971
11	1	2.99900E-01	1.01000E+04	10.054	1.50150E+04	1.4970
11	2	2.99900E-01	1.01000E+04	10.054	1.50150E+04	1.4988
11	3	2.99900E-01	1.01000E+04	10.054	1.50150E+04	1.4988
11	0	2.99900E-01	1.01000E+04	10.054	1.50150E+04	1.4988
11	1	2.99900E-01	1.01000E+04	10.055	1.50150E+04	1.4988
11	2	2.99900E-01	1.01000E+04	10.055	1.50150E+04	1.4987
11	3	2.99900E-01	1.01000E+04	10.055	1.50150E+04	1.4987
11	0	2.99900E-01	1.01000E+04	10.055	1.50150E+04	1.4987
11	1	2.99900E-01	1.01000E+04	10.055	1.50150E+04	1.4987
11	2	2.99900E-01	1.01000E+04	10.055	1.50150E+04	1.4987
11	3	2.99900E-01	1.01000E+04	10.055	1.50150E+04	1.4987

RESIDUE ERROR TO FOURTH ORDER OF SQUARES LESS THAN 1.000000E-05
 DEPARTMENT OF PHARMACY
 SCHOOL OF PHARMACY
 UNIVERSITY OF RHODE ISLAND
 KINGSTON, RHODE ISLAND

TABLE 14. THE 1970-71 ESTIMATION

OF THE 14 EIGENVALUES, THE ESTIMATES AND THE VARIABILITY ARE:

THE EIGENVALUES	STANDARD DEVIATION	95% CONFIDENCE LIMITS	UNIVAR S PLANE
1 101.64	50.3155	100.81.7 99.99.40	10290.7 10370.4
2 10.5496	0.110079	10.4365 9.90850	10.9427 11.1907
3 15173.4	57.1266	15035.7 14960.0	15273.5 15348.7
4 1.33700	3.59607E-02	1.40567 1.35941	1.50852 1.61960

THE CORRELATION-CORRELATION MATRIX OF ESTIMATES ---

1 101.64	1.0000
2 10.5496	0.0000
3 15173.4	0.0000
4 1.33700	0.0000

THE CORRELATION MATRIX OF ESTIMATES ---

1 101.64	1.0000	0.0000	0.0000	0.0000
2 10.5496	0.0000	1.0000	0.0000	0.0000
3 15173.4	0.0000	0.0000	1.0000	0.0000
4 1.33700	0.0000	0.0000	0.0000	1.0000

THE EIGENVALUES OF CORRELATION MATRIX ---

THE EIGENVALUES

1 101.64	0.0000	0.5144	0.0000	0.0000
2 10.5496	0.0000	0.492	0.0000	0.0000
3 15173.4	0.0000	0.0000	0.0000	0.0000
4 1.33700	0.0000	0.0000	0.0000	0.0000

APPENDIX G

SASGRAPH: PROGRAMS AND RESULTS

The data of drug solubility as a function of cyclodextrin concentration and temperature were plotted using a SASGRAPH program. An example of the programs used is shown in this section for each of the drugs studied: phenytoin, ibuprofen, diazepam and hydrochlorothiazide. The results of the three-dimensional plots are then shown in the following order:

1. Phenytoin: β CD: pH 5, 7, 8 and 9
2. Ibuprofen: β CD: pH 2, 4, 5 and 6
3. Diazepam: β CD: pH 2, 3, 4 and 6
4. Hydrochlorothiazide: β CD: pH 5, 8 and 9


```

E
10000 //LJV101A JOB (LJV101),'MENARD',TIME=2,NOTIFY=LJV101
20000 /*JOBPARM L=20
30000 // EXEC SAS
40000 //GO,FT11F001 DD SYSOUT=A,HOLD=YES
50000 //GO,FT12F001 DD SYSOUT=A,HOLD=YES
60000 //GO,AOUT DD SYSOUT=A,DEST=QMS1
70000 //GO,SYSIN DD *
80000 DATA FILE1;
81000 INPUT CD TEMP CONCEN;
82000 CARDS;
83000 0 293 24.34
84000 3 293 80.424
85000 6 293 142.96
86000 9 293 204.00
87000 12 293 263.58
88000 15 293 311.82
89000 0 303 23.97
89100 3 303 84.36
89200 6 303 142.84
89300 9 303 208.59
89400 12 303 266.25
89500 15 303 330.02
89600 0 313 34.90
89700 3 313 99.35
89800 6 313 160.87
89900 9 313 228.24
90000 12 313 296.53
90100 15 313 351.87
90200 0 323 65.64
90300 3 323 137.39
90400 6 323 189.02
90500 9 323 259.75
90600 12 323 328.75
90700 15 323 387.45
90800 ;

```

```

91000  GOPTIONS  NOTEXT82  DEVICE=QMS1200E  NOPOLYGRAPHIC
100000  COLORS=(BLACK)  NOCIRCLEARC  NOCHARACTERB
110000  NODISPLAY  NOPROMPT  AUTOFEED  NOFILL
120000  NODASH  HANDSHAKE=XUNX00  GSELEN=132
130000  GEND=808A0X  GSENAME=PAULI  GSESCALE=AFRONT
140000  TITLE1  J=C  H=2  F=SIMPLEX  C=BLACK
150000  'PHENYTOIN/  CD  F=5  #
160000  TITLE2  J=C  H=1.5  F=SIMPLEX  C=BLACK
170000  '
231000  PROC SORT  *  BY TEMP CD#
232000  PROC PRINT;
233000  PROC G3GRID DATA=STATS;
234000  GRID TEMP*CD=CONCN /
235000  AXIS2=0 TO 15 BY 1
236000  AXIS1=293 TO 323 BY 1.5;
237000  PROC G3D DATA=STATS;
238000  PLOT TEMP*CD=CONCN / SIDE GRID
239000  ROTATE  =60
239100  TILT  =60
239200  ZMAX  =800
239300  ZMIN  =0
240000  /* -
250000  //
END OF DATA

```

```

E
10000 //LJ0101A JOB (LJ0101), MENARD, TIME=2,NB(1)=LJ0101
20000 /*JOBPARM L=20
30000 // EXEC SSB
40000 //GU.F111F001 DD SYSOUT=A,HOLD=YES
50000 //GU.F112F001 DD SYSOUT=A,HOLD=YES
60000 //GU.AOUT DD SYSOUT=A,DEST=RMS1
70000 //GU.SYSIN DD *
80000 DATA FILE1:
81000 INPUT CD TEMP CONCIN:
82000 CARDS:
83000 0 293 35.5
84000 3 293 62.0
85000 6 293 86.1
86000 9 293 114.1
87000 12 293 149.1
88000 15 293 172.4
89000 0 303 47.5
89100 3 303 77.5
89200 6 303 105.8
89300 9 303 138.4
89400 12 303 169.7
89500 15 303 202.2
89600 0 313 64.6
89700 3 313 97.9
89800 6 313 129.2
89900 9 313 166.4
90000 12 313 204.7
90100 15 313 238.4
90200 0 323 83.3
90300 3 323 120.0
90400 6 323 157.9
90500 9 323 203.9
90600 12 323 242.1
90700 15 323 285.0
90800 +

```

```

91000 GOPTIONS NOTEXT82 DEVICE=QMS1200E NOPOLYBUNFILL
100000      COLURS=(BLACK) NOCIRCLEARC NOCHARACTERS
110000      NODISPLAY NOPROMPT AUTOFEED NOFILL
120000      NODASH HANDSHAKE=XONXOFF GSFLEN=132
130000      GEND='8080A'X GSFNAME=AUUI GSFMODE=APPEND;
140000 TITLE1 J=C H=2 F=SIMPLEX C=BLACK
150000      'IBUPROFEN/ CD PH 2';
160000 TITLE2 J=C H=1.5 F=SIMPLEX C=BLACK
170000      '
231000 PROC SORT ; BY TEMP CD;
232000 PROC PRINT;
233000 PROC G3GRID OUT=STATS;
234000      GRID TEMP*CD=LUNLN /
235001      AXIS2=0 TO 5 BY 1
236000      AXIS1=293 TO 323 BY 1.5;
237000 PROC G3D DATA=STATS;
238000      PLOT TEMP*CD=LUNLN / SIDE GRID
239000      ROTATE      =60
239100      TILT       =60
239200      ZMAX        =1800
239300      ZMIN        =0
240000 /*
250000 //
END OF DATA

```

```

21000  GOPTIONS  NOTEXT82  DEVICE=QMS1200E  NOPOLYGONFILL
100000      COLORS=(BLACK)  NOCIRCLEARC  NOCHARACTERS
110000      NODISPLAY  NOPROMPT  AUTOFEED  NOFILL
120000      NODASH  HANDSHAKE=XONXOFF  GSFLEN=132
130000      GEND='B08A'X  GSFNAME=A001  GSFMODE=APPEND
140000  TITLE1  J=C  H=2  F=SIMPLEX  C=BLACK
150000      'DIAZEPAM/  CH  FH 6'
160000  TITLE2  J=C  H=1.5  F=SIMPLEX  C=BLACK
170000      /  /
231000  PROC SURF ; BY TEMP CD;
232000  PROC PRINT;
233000  PROC G3GRID OUT=STATS;
234000  GRID TEMP*CD=CONCN /
235000  AXIS2=0 TO 15 BY 1
236000  AXIS1=293 TO 323 BY 1.5;
237000  PROC G3D DATA=STATS;
238000  PLOT TEMP*CD=CONCN / SIDE GRID
239000  ROTATE  =60
239100  TILT  =60
239200  ZMAX  =600
239300  ZMIN  =0
240000  /*
250000  //
END OF DATA

```

```

10000 //LJV101A JOB (LJV101),'MENARD',TIME=2,NUTIFY=LJV101
20000 /*JOBPARM L=20
30000 // EXEC SAS
40000 //GU.F111F001 DD SYSUUT=A,HOLD=YES
50000 //GU.F112F001 DD SYSUUT=F,HOLD=YES
60000 //GU.A001 DD SYSUUT=A,DEST=QMS1
70000 //GU.SYSIN DD *
80000 DATA FILE1;
81000      INPUT CD TEMP CONDN;
82000      CARDS;
83000 0 293 412.3
84000 3 293 665.54
85000 6 293 856.56
86000 9 293 1104.9
87000 12 293 1303.5
88000 15 293 1478.5
89000 0 303 925.0
89100 3 303 1091.0
89200 6 303 1326.4
89300 9 303 1595.3
89400 12 303 1869.0
89500 15 303 1817.0
89600 0 313 1301.0
89700 3 313 1568.5
89800 6 313 1774.9
89900 9 313 2026.5
90000 12 313 2128.5
90100 15 313 2518.5
90200 0 323 2277.3
90300 3 323 2505.5
90400 6 323 2671.5
90500 9 323 2864.0
90600 12 323 3033.5
90700 15 323 3186.8
90800 ;

```

```

E
10000 //LJV101A JOB (LJV101),'MENARD',TIME=2,NOTIFY=LJV101
20000 /*JOBPARM L=20
30000 // EXEC SAS
40000 //GO.F11F001 DD SYSOUT=A,HOLD=YES
50000 //GO.F12F001 DD SYSOUT=A,HOLD=YES
60000 //GO.AOUT DD SYSOUT=A,DEST=RMS1
70000 //GO.SYSIN DD *
80000 DATA FILE1;
81000 INPUT CD TEMP CONCEN;
82000 CARDS;
83000 0 293 39.4
84000 1 293 186.2
85000 2 293 316.4
86000 3 293 471.0
87000 4 293 582.8
88000 5 293 730.6
89000 0 303 39.87
89100 1 303 179.2
89200 2 303 327.0
89300 3 303 468.0
89400 4 303 609.2
89500 5 303 752.9
89600 0 313 72.6
89700 1 313 218.2
89800 2 313 373.6
89900 3 313 523.0
90000 4 313 666.0
90100 5 313 837.8
90200 0 323 124.6
90300 1 323 258.8
90400 2 323 423.8
90500 3 323 562.4
90600 4 323 706.2
90700 5 323 891.8
90800 ;

```

FIGURE 1
 $\text{PHE NYTORJ}/\rho$ PH 5

FIGURE 2
 $\text{PHENYLOIN}/\rho_{\text{CD}} \text{ PH} /$

FIGURE 3
PHENYTOIN λ_{CD} PH 8

FIGURE 4
 PHENYTOIN/ ρ CD PH 9

FIGURE 5
IBUPROFEN/ β -CD PH 2

FIGURE 6
IBUPROFEN/ ρ CD PH 4

FIGURE 7
ISUPROFEN, CD PH 5

FIGURE 8
IBUPROFEN/CO PH 6

FIGURE 9
DIA. TELAM/βCO PH 2

FIGURE 10
DIAZETAM/ β CD PH 3

FIGURE 11
DIA. EPAM, β CD PH 4

FIGURE 12
 $\text{DIA:TPAM}/\beta$ CD WITH 6

FIGURE 13
HYDRAULIC ROTATION OF ρ_{sed} FOR 5

FIGURE 14
HYDROCHLOROTHIAZIDE / β CD PH 8

FIGURE 15
HYDROCHLOROTHIAZIDE/ β -CD, PH 9

APPENDIX H

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