

1982

## AN INVESTIGATION OF THE POTENTIAL OF SOY CELLULOSE AS A TABLET DISINTEGRANT

Divyakant S. Desai  
*University of Rhode Island*

Follow this and additional works at: <https://digitalcommons.uri.edu/theses>

Terms of Use

All rights reserved under copyright.

---

### Recommended Citation

Desai, Divyakant S., "AN INVESTIGATION OF THE POTENTIAL OF SOY CELLULOSE AS A TABLET DISINTEGRANT" (1982). *Open Access Master's Theses*. Paper 190.  
<https://digitalcommons.uri.edu/theses/190>

This Thesis is brought to you by the University of Rhode Island. It has been accepted for inclusion in Open Access Master's Theses by an authorized administrator of DigitalCommons@URI. For more information, please contact [digitalcommons-group@uri.edu](mailto:digitalcommons-group@uri.edu). For permission to reuse copyrighted content, contact the author directly.

AN INVESTIGATION OF THE  
POTENTIAL OF SOY CELLULOSE  
AS A TABLET DISINTEGRANT

BY

DIVYAKANT S. DESAI

A THESIS SUBMITTED IN PARTIAL  
FULFILLMENT OF THE REQUIREMENTS  
FOR THE DEGREE OF  
MASTER OF SCIENCE  
IN  
PHARMACY

UNIVERSITY OF RHODE ISLAND

1982

SOY CELLULOSE AS A TABLET DISINTEGRANT

MASTER OF SCIENCE THESIS

OF

DIVYAKANT S. DESAI

Approved:

Thesis Committee

Major Professor

*CRM*

*Thomas A. Campbell*

*John M. ...*

*CRM*

*Arithon N. Paruta*

*A. A. Michel*

Dean of the Graduate School

UNIVERSITY OF RHODE ISLAND  
1982

ABSTRACT

Scanning Electron Micrographs of soy cellulose (Emcosoy) particles were studied to further the understanding of the morphology of the substance and thereby to increase the comprehension of how the excipient functions in tablet systems.

Bulk swelling and water uptake were studied for Emcosoy, CLD II, Corn Starch, and Ac-Di-Sol. Emcosoy had maximum swelling about 56%; followed by CLD II 49% and Ac-Di-Sol 9%. No swelling was observed in Corn Starch. An ideal disintegrant should perform uniformly in pH range of gastrointestinal tract in order to disintegrate the tablet. When Emcosoy and Ac-Di-Sol were tested against various pH values, there was no significant difference in swelling of these disintegrants. Since alcohol and sodium lauryl sulfate are often used as dissolution media, the effect of different concentrations were also observed on Emcosoy and Ac-Di-Sol. There was no significant effect.

Another important preformulation test reported in this thesis is a comparative study of the viscosity of different concentrations of various disintegrants. When a tablet disintegrant reacts with water a gel may be formed. If the gel is too viscous, it impedes the water penetration in the tablet. Generally it can be said that the cellulose group of disintegrants (Ac-Di-Sol, CLD II and Emcosoy) had a viscous gel compared to starch group of disintegrants (Corn Starch, Explotab and Sta-Rx 1500). If the viscosity of the gel of starch group of disintegrants has to be arranged **indescending** order then Explotab is followed by Sta-Rx 1500 which is followed by corn starch. If the viscosity of the gel of

cellulose group of disintegrants had to be arranged in descending order then CLD II is followed by Emcosoy which is followed by Ac-Di-Sol.

Plastic deformation and elasticity of various disintegrants were studied using a Universal Instron Testing Machine. High plastic deformation of the disintegrant particles, may be advantageous since the deformed particles are "energy-rich" and that energy is released when the particles are exposed to water. The energy-rich particles probably swell more rapidly in water, unlike undeformed grains, which require more energy of swelling in order to well. Ac-Di-Sol particles have good plastic deformation compared to Emcosoy and Explotab particles. The three major mechanisms of disintegrant action are : (i) swelling (ii) capillary action (wicking) and (iii) deformation. Emcosoy particles swell considerably and they have good wicking effect, but they have poor plastic deformation.

Following the performulation study, Emcosoy (soy cellulose) was compared with other disintegrants in about 45 formulations. About 100 dissolution profiles were obtained on various vitamin formulations using either Emcompress or Endex as the tablet matrix. Emcosoy is as effective a disintegrant as Ac-Di-Sol and very much more effective than Corn Starch. The results of preformulation study and formulation study strongly indicate that Emcosoy is a powerful tablet disintegrant which deserves careful consideration by all formulators.

ACKNOWLEDGMENTS

I want to express my deep gratitude to Dr. Christopher T. Rhodes who is also guardian of my shortcomings, for his unceasing encouragement and guidance during this study and preparation of thesis.

I also want to thank my graduate committee members, Dr. Anthony N. Paruta and Dr. Norman A. Campbell, for their advice and encouragement.

Grateful acknowledgment is made of the invaluable cooperation and support of Edward Mendell Company, and special thanks to Dr. Joseph L. Kanig for his advice during the study.

Appreciation is also acknowledged of the invaluable cooperation of Dr. Joan M. Lausier in solving many problems with the tablet press.

Grateful acknowledgment is also made of the invaluable cooperation of Dr. Chong M. Lee and Dr. Martin A. Saad in conducting elasticity study on Instron Testing Machine.

Thanks are also due to Donald Scales for Scanning Electron Microscopy work, to Jack Hoblitzell for developing the negative of Scanning Electron Micrographs, to Edward M. Rudnic and Rao N. Chilamkurti for their invaluable help at all stages of the study.

I also wish to acknowledge the help of Dr. George E. Osborne for kindly reading the manuscripts. Appreciation is also acknowledged of the formidable task of Mrs. Susan Killian and Miss Robin Dexter who typed the manuscript.

## TABLE OF CONTENTS

	PAGE
ABSTRACT . . . . .	i
ACKNOWLEDGEMENTS . . . . .	iii
TABLE OF CONTENTS . . . . .	iv
LIST OF TABLES . . . . .	vi
LIST OF FIGURES . . . . .	ix
I. INTRODUCTION . . . . .	1
A. 1. Bioavailability Considerations . . . . .	2
2. Disintegration of a Tablet and Disintegrants . . . . .	7
3. Mechanism of Disintegrant Action . . . . .	13
B. 1. Objective and Justification for the Present Study . . . . .	17
II. EXPERIMENTAL . . . . .	19
A. Equipment . . . . .	19
B. Materials . . . . .	21
C. Methods . . . . .	23
1. Scanning Electron Micrographs . . . . .	23
2. Bulk Swelling and Water Uptake . . . . .	25
3. Water Penetration of Disintegrants . . . . .	28
4. Energy of Wetting . . . . .	29
5. Wetting and Drying of Disintegrants . . . . .	31
6. Aqueous Dispersion . . . . .	31
7. Comparative Study of the Viscosity of Different Strength Solutions on Various Disintegrants . . . . .	32
8. Maximum Hydration Capacity of Various Disintegrants . . . . .	34
9. Emcosoy - Sieve Analysis . . . . .	35
10. Plastic Deformation and Elasticity of Various Disintegrants . . . . .	35



	PAGE
12. Tablet Productions . . . . .	43
a. Weight . . . . .	44
b. Thickness . . . . .	45
c. Hardness . . . . .	45
d. Friability . . . . .	46
e. Disintegration . . . . .	46
f. Dissolution . . . . .	46
III. RESULTS AND DISCUSSIONS . . . . .	50
1. Scanning Electron Mircographs . . . . .	50
2. Bulk Swelling and Water Uptake . . . . .	50
3. Water Penetration of Disintegrants . . . . .	67
4. Energy of Wetting . . . . .	72
5. Wetting and Drying of Disintegrants . . . . .	72
6. Aqueous Dispersion . . . . .	79
7. Comparative Study of the Viscosity of Different Strength Solutions of Various Disintegrants . . . . .	79
8. Maximum Hydration Capacity of Various Disintegrants .	89
9. Emcosoy: Sieve Analysis . . . . .	89
10. Plastic Deformation and Elasticity of Various Disintegrants . . . . .	91
11. Tablet Production . . . . .	99
IV. CONCLUSION	176
V. SUGGESTIONS FOR FUTURE WORK	178
VI. REFERENCES	179
VII. BIBLIOGRAPHY	183
VIII. APPENDIX	187

## LIST OF TABLES

TABLE	PAGE
I. Swelling of Different Disintegrants in Water. . . . .	60
II. Swelling of Emcosoy in Citrate-Phosphate-Borate/Hcl Buffer. . . . .	60
III. t-Test for Buffer Reading . . . . .	60
IV. Swelling of Ac-Di-Sol in Distilled Water. . . . .	61
V. Swelling of Emcosoy in Alcohol. . . . .	62
VI. t-Test for Alcohol Reading with Emcosoy as Disintegrant	63
VII. Swelling of Emcosoy in Sodium Lauryl Sulfate. . . . .	63
VIII. t-Test for Sodium Lauryl Sulfate Reading with Emcosoy as a Disintegrant . . . . .	64
IX. Swelling of Ac-Di-Sol in Citrate-Phosphate-Burase/Hcl Buffer. . . . .	64
X. t-Test for Citrate-Phosphate-Borak/Hcl Buffer Reading with Ac-Di-Sol as Disintegrant. . . . .	65
XI. Swelling of Ac-Di-Sol in Sodium Lauryl Sulfate. . . . .	65
XII. t-Test for Sodium Lauryl Sulfate Reading. . . . .	65-a
XIII. Swelling of Ac-Di-Sol in Alcohol. . . . .	66
XIV. t-Test for Ac-Di-Sol Swelling in Alcohol. . . . .	66
XV. Water Penetration in (cm) <sup>2</sup> in Emcosoy Powder Bed at 22 ± 1°C . . . . .	68
XVI. Water Penetration in (cm) <sup>2</sup> in Ac-Di-Sol Powder Bed at 22 ± 1°C . . . . .	68
XVII. Water Penetration in (cm) <sup>2</sup> in CLD II Powder Bed at 22 ± 1°C . . . . .	68

XVIII.	Water Penetration in $(\text{cm})^2$ in Explotab Powder Bed at $22 \pm 1^\circ\text{C}$ . . . . .	60
XIX.	Energy of Wetting of Emcosoy Particles . . . . .	74
XX.	Comparison of Swelling of Original Emcosoy and Modified Emcosoy. . . . .	77
XXI.	Comparison of the Physical Properties of the Tablets Made by Using Original Emcosoy and Modified Emcosoy. . .	78
XXII.	Viscosity Study of Gel Formed by Various Disintegrants .	80
XXIII.	Viscosity Study of Gel Formed by Various Disintegrants .	81
XXIV.	Emcosoy (2%) Dispersion in Water and Dilute Acid (Hcl 1 in 100). . . . .	90
XXV.	Emcosoy -- Sieve Analysis. . . . .	90
XXVI.	Modulus of Elasticity of Various Disintegrants . . . . .	92
XXVII.	Percentage Plastic Deformation of Various Disinte- grants . . . . .	93
XXVIII.	Multivitamin Formulations with Emcompress as a Matrix. .	102
XXIX.	Multivitamin Formulations with Emdex as a Matrix . . . .	103
XXX.	Pyridoxine Hydrochloride Formulations with Emcompress as a Matrix. . . . .	104
XXXI.	Pyridoxine Hydrochloride Formulations with Emdex as a Matrix. . . . .	105
XXXII.	Ascorbic Acid Formulation With Emcompress as a Matrix. . . . .	106
XXXIII.	Dissolution Data of Multivitamin Formulations with Emcompress as a Matrix. . . . .	107
XXXIV.	Dissolution Data of Multivitamin Formulations with Emdex as a Matrix . . . . .	119

XXXV.	Dissolution Data of Pyridoxine Hydrochloride Formulation with Emcompress as a Matrix. . . . .	131
XXXVI.	Dissolution Data of Pyridoxine Hydrochloride Formulation with Emdex as a Matrix . . . . .	144
XXXVII.	Dissolution Data of Ascorbic Acid Formulation with Emcompress as a Matrix. . . . .	156

## LIST OF FIGURES

FIGURE	PAGE
1. Working Principle of Scanning Electron Microscope. . .	24
2. U-tube Apparatus . . . . .	26
3. The U.S.P. Disintegration Apparatus In Use . . . . .	47
4. Scanning Electron Micrographs of Emcosoy Particles . .	51
5. Scanning Electron Micrographs of Emcosoy Particles . .	52
6. Percentage Swelling of Ac-Di-Sol as Function of Different Weights of Disintegrants . . . . .	54
7. Log of Percentage Swelling of Ac-Di-Sol as Function of Different Weights of Disintegrants . . . . .	55
8. Milliliter of Water Travelled in Ac-Di-Sol as Function of Different Weights . . . . .	56
9. Water Absorption Test in Ac-Di-Sol . . . . .	57
10. Percentage Swelling of Ac-Di-Sol as a Function Dif- ferent Concentration Sodium Lauryl Sulfate Solution. .	58
11. Percentage Swelling of Ac-Di-Sol as a Function of Different Concentrations of Alcohol. . . . .	59
12. The Penetration of Water Into Various Disintegrants. .	70
13. Energy of Wetting of Emcosoy Particles . . . . .	73
14. Dilatant, Newtonian, and Pseudo Plastic Profiles . . .	84
15. Rheological Profiles of Explotab, Emcosoy, and Ac-Di- Sol -- All at 10% Concentration. . . . .	85
16. Rheological Profile of Corn Starch and Sta-Rx 1500 at 14% Concentration . . . . .	86
17. Rheological Profile of Poliplasdone XL at 14% Con- centration of the Disintegrant . . . . .	87
18. Rheological Profile of CLD II Gel at 4% Concentra- tion of the Disintegrant . . . . .	88

19.	Dissolution Profiles of Multivitamin Formulation, Emcompress as a Matrix, Emcosoy, Ac-Di-Sol and Explotab at 0.5% Concentration. . . . .	108
20.	Three Dissolution Profiles of Multivitamin Formulations, Emcompress as a Matrix, and Emcosoy as a Disintegrant at 0.5% Concentration. . . . .	109
21.	Three Dissolution Profiles of Multivitamin Formulations Containing 0.5% Ac-Di-Sol, Emcompress as a Matrix. . . . .	110
22.	Three Dissolution Profiles of Multivitamin Formulations Containing 0.5% Explotab, Emcompress as a Matrix . . . . .	111
23.	Dissolution Profiles of Multivitamin Formulations, Emcompress as a Matrix, Ac-Di-Sol and Explotab at 1.0% Concentration. . . . .	112
24.	Three Dissolution Profiles of Multivitamin Formulations Containing 1% of Emcosoy, Emcompress as a Matrix . . . . .	113
25.	Three Dissolution Profiles of Multivitamin Formulations Containing 1% of Ac-Di-Sol, Emcompress as a Matrix . . . . .	114
26.	Three Dissolution Profiles of Multivitamin Formulations Containing 1% of Explotab, Emcompress as a Matrix. . . . .	115
27.	Dissolution Profiles of Multivitamin Formulations, Emcompress as a Matrix, Emcosoy and Corn Starch at 3.0% Concentration. . . . .	116
28.	Two Dissolution Profiles of Multivitamin Formulations Containing 3% of Emcosoy, Emcompress as a Matrix . . . . .	117
29.	Two Dissolution Profiles of Multivitamin Formulations Containing 3% of Corn Starch, Emcompress as a Matrix . . . . .	118
30.	Dissolution Profiles of Multivitamin Formulations Containing 0.5% of Emcosoy, Ac-Di-Sol, and Explotab, Emdex as a Matrix. . . . .	120
31.	Three Dissolution Profiles of Multivitamin Formulations Containing 0.5% of Emcosoy, Emdex as a Matrix. . . . .	121
32.	Three Dissolution Profiles of Multivitamin Formulations Containing 0.5% of Ac-Di-Sol, Emdex as a Matrix. . . . .	122

33.	Three Dissolution Profiles of Multivitamin Formulations Containing 0.5% of Explotab, Emdex as a Matrix. . . . .	123
34.	Dissolution Profiles of Multivitamin Formulations Containing 1.0% of Emcosoy, Ac-Di-Sol, and Explotab, Emdex as a Matrix . . . . .	124
35.	Three Dissolution Profiles of Multivitamin Formulations Containing 1.0% of Emcosoy, Emdex as a Matrix . . . . .	125
36.	Three Dissolution Profiles of Multivitamin Formulations Containing 1.0% of Ac-Di-Sol, Emdex as a Matrix . . . . .	126
37.	Three Dissolution Profiles of Multivitamin Formulations Containing 1.0% of Explotab, Emdex as a Matrix. . . . .	127
38.	Dissolution Profiles of Multivitamin Formulations Containing 3.0% of Emcosoy and Corn Starch, Emdex as a Matrix. . . . .	128
39.	Two Dissolution Profiles of Multivitamin Formulations Containing 3% of Emcosoy, Emdex as a Matrix. . . . .	129
40.	Two Dissolution Profiles of Multivitamin Formulations Containing 3% of Corn Starch, Emdex as a Matrix. . . . .	130
41.	Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 0.5% of Emcosoy, Explotab, and Ac-Di-Sol. Emcompress as a Matrix. . . . .	132
42.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 0.5% Emcosoy, Emcompress as a Matrix . .	133
43.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 0.5% Ac-Di-Sol, Emcompress as a Matrix .	134
44.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 0.5% Explotab, Emcompress as a Matrix. .	135
45.	Dissolution Profiles of Hyridoxine Hydrochloride Formulations Containing 1.0% of Emcosoy, Ac-Di-Sol and Explotab, Emcompress as a Matrix. . . . .	136
46.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containg 1.0% of Emcosoy, Emcompress as a Matrix. .	137

47.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 1.0% of Ac-Di-Sol, Emcompress as a Matrix. . . . .	138
48.	Two Dissolution Profiles of Pyridoxine Hydrochloride Containing 1% of Explotab, Emcompress as a Matrix. . . . .	139
49.	Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 2.0% of Emcosoy, Ac-Di-Sol, Explotab, and Corn Starch; Emcompress as a Matrix . . . . .	140
50.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 2.0% Emcosoy, Emcompress as a Matrix . .	141
51.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 2% of Ac-Di-Sol, Emcompress as a Matrix.	142
52.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 2% of Explotab, Emcompress as a Matrix .	143
53.	Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 0.5% of Emcosoy, Ac-Di-Sol and Explotab; Emdex as a Matrix. . . . .	145
54.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 0.5% of Emcosoy, Emdex as a Matrix . . .	146
55.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 0.5% of Ac-Di-Sol, Emdex as a Matrix . .	147
56.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 0.5% of Explotab, Emdex as a Matrix. . .	148
57.	Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 1% of Emcosoy, Ac-Di-Sol and Explotab; Emdex as a Matrix . . . . .	149
58.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 1% of Emcosoy, Emdex as a Matrix . . . .	150
59.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 1% of Ac-Di-Sol, Emdex as a Matrix . . .	151



60.	Two Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 1% of Explotab, Emdex as a Matrix. . . .	152
61.	Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 0.5% of Emcosoy and Corn Starch, Emdex as a Matrix. . . . .	153
62.	Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 3% of Emcosoy, Emdex as a Matrix . . . . .	154
63.	Dissolution Profiles of Pyridoxine Hydrochloride Formulations Containing 3% of Corn Starch, Emdex as a Matrix . . . .	155
64.	Dissolution Profiles of Ascorbic Acid Formulations Containing 0.5% of Ac-Di-Sol and Emcosoy, Emcompress as a Matrix . .	157
65.	Two Dissolution Profiles of Ascorbic Acid Formulations Containing 0.5% of Emcosoy, Emcompress as a Matrix . . . . .	158
66.	Two Dissolution Profiles of Ascorbic Acid Formulations Containing 0.5% of Ac-Di-Sol, Emcompress as a Matrix . . . . .	159
67.	Dissolution Profiles of Ascorbic Acid Formulations Containing 1.0% of Emcosoy, Ac-Di-Sol and Explotab, Emcompress as a Matrix. . . . .	160
68.	Two Dissolution Profiles of Ascorbic Acid Formulations Containing 1.0% of Emcosoy, Emcompress as a Matrix. . . . .	161
69.	Two Dissolution Profiles of Ascorbic Acid Formulations Containing 1% of Ac-Di-Sol, Emcompress as a Matrix . . . . .	162
70.	Two Dissolution Profiles of Ascorbic Acid Formulations Containing 1.0% of Explotab, Emcompress as a Matrix. . . . .	163
71.	Dissolution profiles of Ascorbic Acid Formulations Containing 2.0% of Emcosoy, Ac-Di-Sol, Explotab and Corn Starch, Emcompress as a Matrix. . . . .	164
72.	Dissolution Profiles of Ascorbic Acid Formulations Containing 2.0% of Emcosoy, Replicate 1, Emcompress as a Matrix. . . .	165
73.	Dissolution Profiles of Ascorbic Acid Formulations Containing 2.0% of Emcosoy, Replicate 2, Emcompress as a Matrix. . . . .	166

74. Dissolution Profiles of Ascorbic Acid Formulations Contain-  
ing 2.0% of Ac-Di-Sol, Replicate 1, Emcompress as a Matrix. . . 167
75. Dissolution Profiles of Ascorbic Acid Formulations Contain-  
ing 2.0% of Ac-Di-Sol, Replicate 2, Emcompress as a Matrix. . . 168
76. Dissolution Profiles of Ascorbic Acid Formulations Contain-  
ing 2.0% of Explotab, Replicate 1, Emcompress as a Matrix . . . 169
77. Dissolution Profiles of Ascorbic Acid Formulations Contain-  
ing 2.0% of Explotab, Replicate 2, Emcompress as a Matrix . . . 170
78. Dissolution Profiles of Ascorbic Acid Formulations Contain-  
ing 2.0% of Corn Starch, Replicate 1, Emcompress as a Matrix. . 171
79. Dissolution Profiles of Ascorbic Acid Formulations Contain-  
ing 2.0% of Corn Starch, Replicate 2, Emcompress as a Matrix. . 172
80. Dissolution Profiles of Ascorbic Acid Formulations Contain-  
ing 3.0% of Corn Starch and Emcosoy, Emcompress as a Matrix . . 173
81. Two Dissolution Profiles of Ascorbic Acid Formulations Con-  
taining 3% of Emcosoy, Emcompress as a Matrix . . . . . 174
82. Two Dissolution Profiles of Ascorbic Acid Formulations Con-  
taining 3% of Corn Starch, Emcompress as a Matrix . . . . . 175

## I. INTRODUCTION

There are two classes of drugs administered orally in tablet dosage form. These are: (1) insoluble drugs intended to exert a local effect in the gastrointestinal tract (such as many antacids and absorbents), and (2) soluble drugs intended to exert a systemic drug effect following their dissolution in the gut and subsequent absorption.

In the case of drug products intended to exert a systemic effect, the design of a dosage form which rapidly disintegrates and dissolves may or may not be critical, depending on whether the drug is absorbed in the upper gastrointestinal tract or more generally throughout the intestinal tract. Because dosage forms must be designed to disintegrate or dissolve to release the drug in an available form at or above the region of absorption in the gut, the design must also be based on the solubility properties of the drug at or above its absorption site.

The present study **covers** the bioavailability consideration of the tablet as a dosage form, the disintegration process in general, and the information about a few widely accepted new disintegrants. The mechanism of disintegrant action will be discussed with the help of numerous references on the subject.

1. Bioavailability Considerations:

Bioavailability has been defined as the measurement of both the relative amount of an administered dose that reaches the general circulation (i.e., the extent of absorption of a given dose) and the rate at which this occurs (1).

Some of the most impressive biopharmaceutical research with practical implications, published during the middle nineteen sixties, was that which demonstrated that different products of the same drug, although chemically identical in terms of drug content, could produce significantly different blood levels. For example, a study of the blood levels produced by different brands of tetracycline, all containing the same amount of drug, showed that the best brand gave blood levels ten times as high as the worst (2).

Glazko, et al, showed that four different brands of 250 mg chloramphenicol capsules had different in vitro and in vivo bioavailability (3). Levy, et al, studied absorption and dissolution of two brands of phenytoin capsules and found that there was a significant difference in the bioavailability of the drug because each product differed from the other in disintegration and deaggregation (4). Bioavailability has been a major problem for pharmaceutical dosage formulators. The subject has drawn considerable attention since the nineteen fifties. Proctor, in 1862, stated: "The reason why

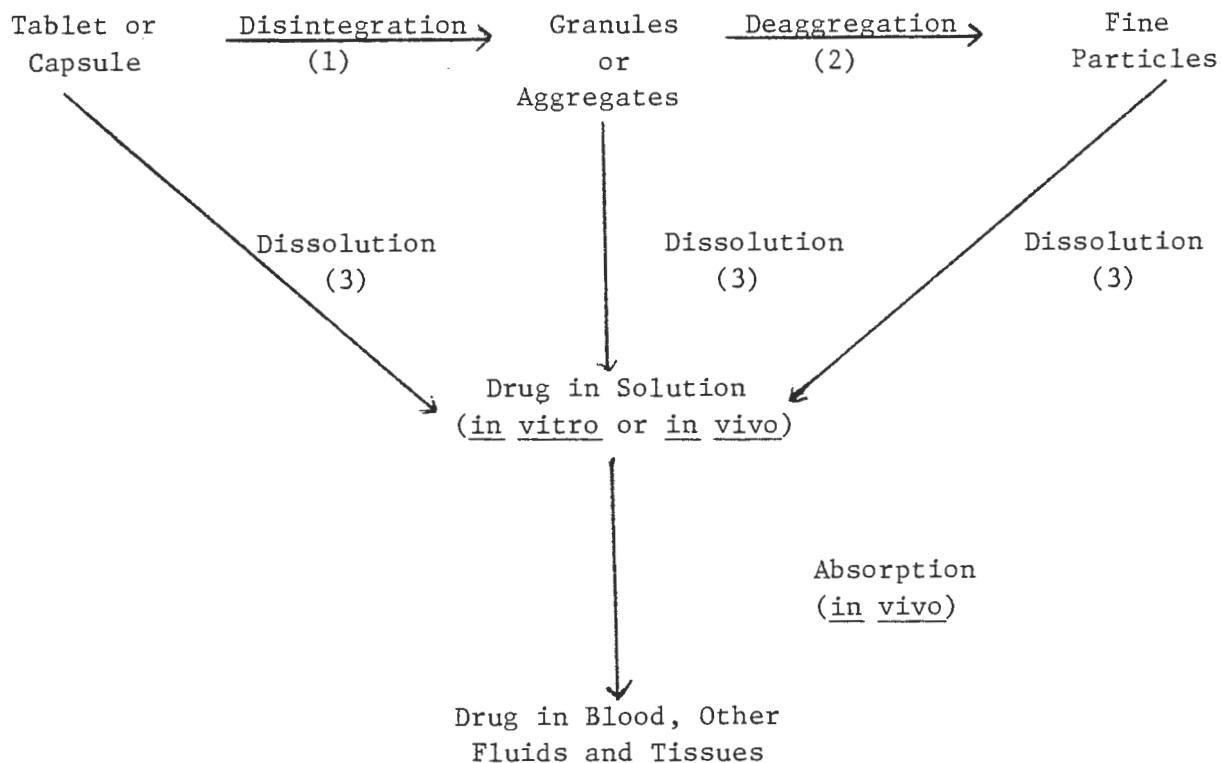
pills occasionally pass through the stomach undissolved must be sought for in the state of the patient or in the composition of the pill, rather than in the nature of the coating." (5). Proctor was not discussing enteric-coated pills, since the first enteric coating (Keratin) was described by Unna in 1884 -- 22 years later. Today, we realize Proctor's statement is only partially true, and that all three factors he discussed, the state of the patient, the composition and the nature of the coating, may determine in vivo disintegration time and rate of dissolution and absorption of the drug.

Unfortunately, Proctor's concern was not shared by other scientists. The need to quantify tablet disintegration by official standards was not recognized until just prior to World War II, and specifications for tablet disintegration did not appear in the USP until the fourteenth revision which was published in 1950. During the course of subsequent disintegration testing experiments, it **was theorized and eventually** proven that drug dissolution must follow tablet disintegration if bioavailability is to occur (6).

The topic of bioavailability has now become of interest to government; the FDA (Food and Drug Administration) has already issued specific regulations on bioavailability.

When a drug in a dosage form is administered to a patient, the first steps in the sequence whereby the drug reaches the site of action are commonly disintegration and dissolution. The importance of these processes is

understood when one realizes that solid oral dosage forms (mainly compressed tablets or hard gelatin capsules) are by far the most important dosage form class in terms of either doses administered per year or dollar value of drug dispensed (7). The process of disintegration, dissolution, and absorption can be outlined as follows (8):



Before drugs can effectively pass through the gastrointestinal wall, they must be in solution. Drugs which are only sparingly soluble in the gastrointestinal content at or above their absorption site can have, as the controlling process affecting their absorption, the rate of drug solution in these fluids. In this type of system, the drug goes into solution at a slow rate, absorption occurs almost immediately and is

not, therefore, the rate-limiting step. In one study, Nelson (9) correlated the blood level concentration of various theophylline salts with their dissolution rates.

Drugs which exert a systemic effect must dissolve as a prerequisite to effective drug absorption. The various processes of tablet making, including the aggregation of drug into granular particles, the use of binders, and the compaction of the system into a dense tablet, are all factors which militate against a rapid drug dissolution and absorption in the gastrointestinal tract. In considering, in a general manner, the availability of drugs from various classes of dosage forms, drugs administered in solution will usually produce the most available drug product, provided that the drug does not precipitate in the stomach or is not deactivated there. The second most available form of a therapeutic agent would be drug dispersed in a fine suspension, followed by micronized drug in capsule form, followed by uncoated tablets, with coated tablets being the least bioavailable drug product in general. In formulating and designing drug products, as well as in considering methods of manufacture, the fact that the tablet dosage form is one of the least bioavailable forms (all other factors being equal) should be kept in mind (10).

Many factors can affect drug dissolution rates from tablets; hence, possibly drug bioavailability, including the crystal size of the drug, tablet disintegration mechanisms and rates, the method of granulation,

type and amount of granulating agent employed, type, amount and method of incorporation of disintegrants and lubricants, and other formulations and processing factors.

Levy, et al (11), showed the effect of granule size upon the dissolution rate of salicylic acid. Salicylic acid of two mesh ranges containing 300 mg of aspirin and 60 mg of starch, were compressed at  $715 \text{ kg cm}^{-2}$ . The 60 to 80 mesh granules had better vitro bioavailability than 40 to 60 mesh granules.

Lachman, et al (12), studied the effect of crystal size and granule size on a delayed-action matrix using tripeleennamine hydrochloride. He notes that while granule and crystal size both affected release rate, in this instance the crystal size played a greater role than granule size in dissolution rate.

Paul, et al (13) showed that with nitrofurantoin there was an optimal average crystal size of about 150 mesh, which resulted in adequate drug excretion (hence, absorption and efficacy), but minimized emesis. This exemplifies a situation in which too rapid drug dissolution in the stomach may produce nausea and emesis; an intermediate release rate reduced this effect while achieving adequate bioavailability.

Numerous accounts of the effect of particle size on dissolution rate of steroids have been reported. In one study, Campagna, et al (14), showed that, in spite of good disintegration, therapeutic inefficacy of prednisone tablets could occur.



As discussed above, bioavailability of the drug depends on many factors. The ensuing discussion will focus on the disintegration process, disintegrants available, and the mechanism action of disintegrants.

## 2. Disintegration of a Tablet and Disintegrants

Complete tablet disintegration is defined by NF XIII (15) as: ". . . that state in which any residue of the tablet, except fragments of insoluble coating, remaining on the screen is a soft mass having no palpably firm core." This often makes tablet disintegration a necessary first step to achieve rapid availability of the active ingredient(s). The importance of tablet disintegration was recognized as early as 1879, when a patent recommended that pills be perforated to admit gastric juice for better disintegration (16).

### Reasons for Measuring Disintegration Times and Rates of Dissolution

- (i) For research purposes to elucidate the mechanism involved in the processes and to determine the relative importance of the various variables involved in the process of disintegration, deaggregation, and dissolution.
- (ii) For developmental purposes to guide the pharmaceutical formulator in the preparation of optimum dosage forms of drugs for clinical trial.

- (iii) For control purposes to ensure that a given pharmaceutical product is essentially uniform from lot to lot.
- (iv) For predictive purposes so that one may estimate rate(s) of absorption in man from measurement of disintegration time and/or rates of dissolution in vitro. Such predicting requires careful correlation of in vitro and in vivo results.

Our discussion of disintegration will be confined to uncoated tablets designed to release all the active ingredient(s) rapidly.

The disintegration of a tablet depends on compression force, tablet hardness, properties of fillers and active ingredients, properties of binders, properties and concentration of disintegrant and the properties of lubricants. All these factors play an important role in tablet disintegration. If we keep other factors identical (compressive force, fillers, active ingredient binders, and lubricant) in two formulations but select two different disintegrants at the same concentration for these two formulations, the in vitro bioavailability will entirely depend on the quality of the disintegrants. The drug will be released quickly from a tablet which disintegrates faster because it has better disintegrants. Thus, the disintegrants play a role in the disintegration process. The ensuing discussion will focus on different disintegrants.

Disintegrant is a term applied to substance added to a tablet granulation for the purpose of causing the compressed tablet to break apart when placed into an aqueous environment. The disintegrant in a tablet formula may be considered as a dispersing agent for the dry compacted tablet mass in the gastric milieu. Ideally, it should cause the tablet to disrupt not only into the granulated form which it was compressed, but also into the powder particles from which the granulation was prepared.

The function of the disintegrant is, in effect, to counteract the action of the tablet binders and the physical forces of compression necessary to form the tablet. The stronger the effect of the binder, the more efficient must be the disrupting effect of the disintegrant in order to release the active ingredient in the gastrointestinal tract (17).

There are two methods used for incorporating disintegrating agents into tablets. These methods are called external addition and internal addition. The most common method is the external addition method in which the disintegrant is added to the sized granulation with mixing just prior to compression. In the internal addition method, the disintegrant is mixed with other powders before wetting the powder mixture with the granulating solution. Thus, the disintegrant is incorporated within the granule. When this method is used, part of the disintegrant is added internally and part by external addition. Many experts believe that use of the two-step method usually produces better and more complete disintegration than the usual method of adding the dis-

integrant to the granulation surface only.

Six basic categories of disintegrants have been described: starches, clays, celluloses, algin, gums, and miscellaneous. Many disintegrants have also been shown to possess binder or adhesive properties. Since disintegration is the opposite operation to granulation (agglomeration) and the subsequent formation of strong compacts, one must carefully weigh these two phenomena when designing a tablet (18).

Formulators have tried a number of materials as tablet disintegrants with varying degrees of success. Lowenthal tabulated all the disintegrants with their pertinent references in a review article of "Disintegration of Tablet," in Journal of Pharmaceutical Sciences (19).

In the past six to seven years, exciting developments have occurred in the area of tablet disintegrants. A number of new disintegrants have been marketed in this period. Attention will be focused on these "super disintegrants," as they are called, because of their remarkable quality as disintegrants.

(i) Direct-Compression Starch:

One of the most significant modifications of starch for direct-compression tableting came with the advent of starch 1500 (Colorion, Inc., West Point, Pennsylvania, originally marketed by A.E. Staley Co. as Sta-Rx 1500). The material is prepared by subjecting ordinary corn starch to physical compression or shear in high moisture conditions; that preparation causes both an increase in temperature and a partial

gelatinization of some of the starch. Direct compression starch retains its disintegrating characteristics despite its partial pregelatinization. While plain starch acts as a disintegrant primarily through wicking action and the elastic recovery of compression-deformed grains when they are wetted, Starch 1500 also acts by truly swelling in the presence of moisture.

(ii) Sodium Carboxymethyl Starch:

Another of the modifications of starch aimed at improving its disintegration properties involves chemical changes that form a low-substituted sodium carboxymethyl starch. The addition of the carboxymethyl groups makes the starch grains more hydrophilic -- but not completely water soluble. The process also causes a two- to three-fold increase in the size of the starch grains. A minimum of 3% to 4% sodium carboxymethyl starch is required for most formulations. The sodium carboxymethyl starch is marketed as Primojel HV (Generichem Corp., Little Falls, New Jersey, and Explotab, Edward Mendell Co., Carmel, New York).

(iii) Cross-Linked Sodium Carboxymethyl Cellulose:

Internally cross-linked carboxymethyl cellulose fibers have been introduced commercially -- as Ac-Di-Sol (FMC Corp., Philadelphia, Pennsylvania), and as CLD Cellulose (Buckeye

Cellulose Corp., Memphis, Tennessee), for use as tablet disintegrants. The cross-linking greatly reduces water solubility while still permitting the material to swell and absorb many times its weight in water without losing individual fiber integrity. Cross-linked sodium carboxymethyl cellulose may be used as a tablet disintegrant in concentrations as low as 1%, although 2% is recommended for tablets made by direct compression, and a 3% level for those made by wet granulation.

(iv) Cross-Linked Polyvinylpyrrolidone:

Cross-linked polyvinylpyrrolidone (PVP) is a homopolymer of N-vinyl-d-pyrrolidene and has been marketed as Polyplasdone-XL (GAF Corp., New York, New York). Because of its high molecular weight and cross-linked structure, it is insoluble in water but is still very hydrophilic. The particles of PVP are porous in nature. As water is absorbed into the porous structure of the agglomerates, the lattice structure of polymer expands, causing high stress on surrounding tablet components. The porous nature of the particles provides intraparticulate wicking of water. Like cross-linked carboxymethyl cellulose, cross-linked PVP has the ability to shrink to its original particle size when dried and then expand again when rewetted. Both cross-linked carboxymethyl cellulose and cross-linked PVP would thus appear to be effective disintegrants in tablets made by wet granulation, as well as in those produced by direct

compression.

Shangraw, et al, discussed the properties of these disintegrants in detail with their scanning electron micrographs in the Pharmaceutical Technology article (20).

### 3. Mechanism of Disintegrant Action:

For various disintegrants, various theories relating to their disintegration actions have been proposed; but none has truly explained the disintegrant properties of all the agents. The three major mechanisms of action are (i) swelling, (ii) porosity and capillary action (wicking), and (iii) deformation.

#### (i) Swelling:

Rank orders of decrease in swelling of various starches under different conditions were described as: wheat, corn, potato, arrowroot, and rice (21); potato, corn and wheat (22); and potato, wheat, and rice (23).

Tablets made with low pressure have high porosity, and hence, too much space. When starch swells, no pressure is exerted; therefore, disintegration is slow. Medium pressure allows just enough space so that when the starch swells, it exerts pressure on the granules to cause disintegration. High pressure, producing low porosity, decreases the ability of fluid to enter; so, disintegration is again slow (24, 25). Starch swelling was claimed to be dependent upon amylose and

amylopectin content; the amylopectin expands, and the amylose gives osmotic pressure (26).

Borzenou and Nesmiyan (27) reported that the degree of swelling of acid carboxymethyl cellulose was 220%; of sodium carboxymethyl cellulose, 400%; of sodium alginate, 450%; and of ultra-amylopectin, 1100%. A mixture of starch with 0.1% ultra-amylopectin gave maximum swelling, while acid carboxymethyl cellulose - starch mixture (3:97) gave the **second largest swelling**. A mixture of ultraamylopectin-starch (2:97) as 10% of a formulation gave the best disintegration times, while acid carboxymethyl cellulose - starch (3:97) gave the second best disintegrant times. It was suggested that starch forms the capillaries, and the ultra-amylopectin acts as a swelling agent (28).

Many substances swell to a greater degree than the starches, but are poorer disintegrants. Amylose does not swell but has been stated to cause good disintegration (29). Although starch grains swell in water according to some views which I do not share, the rate and extent of swelling is still debated (30-32).

Whenever swelling of disintegrant particles take place with great force, it overcomes the adhesiveness of other ingredients in a tablet and causes the tablet to fall to powder.

(ii) Porosity and Capillary Action (Wicking):

Porosity of tablets has been studied with increasing fre-



quency in recent years, using a variety of materials and numerous experimental methods. Porosity, void space, and pore size decrease as pressure increases (33-46). Potato and corn starches increase mean pore diameters and porosity (47-50). Pore size and volume decrease as the moisture content of starch increases.

The rate of penetration of fluids into a tablet is proportional to mean pore diameter or porosity (51-52); corn and starches increase penetration of fluids into tablets. Permeability of tablet decreases as pressure increases. The effect of starch on porosity may be due to its poor ability to bond and compress (52). In 1955, Curlin reported that although aspirin tablets containing starch disintegrated in 15 seconds, starch grains were not swollen; nevertheless, a drop of dye solution placed on the tablet penetrated rapidly. He suggested that the disintegrating action of starch was due to capillary action, rather than to swelling (53).

Wicking is due to capillarity of fibers. Stiff fibers of uniform structure and resistance to collapse are required for good wicking. The fibers should have zero contact angle and should not swell (54). This would appear to rule out any wicking effect due to starch or cellulose fibers.

The existence of pores or capillaries is not the complete answer to the mechanism of action of disintegrants, because semipolar and nonpolar fluids penetrate into tablets (55), yet do not cause the tablets to break. Also, tablets do disintegrate with minimum porosity (56).

(iii) Deformation:

Plastic deformation of starch grains under high pressure has been reported by a number of investigators (57). Starch grains are generally thought to be elastic; therefore, any grains that are deformed under pressure tend to return to their original shape and size when the pressure is removed. However, it has been suggested that compression may cause more permanent deformation -- that the deformed starch grains are energy rich and that this energy is released when the grains are exposed to water (58). The energy-rich starch grains swell rapidly in water, unlike undeformed grains, which require more heat in order to swell.

The various mechanism of disintegration action discussed under a separate heading, Interrelationships, probably occur in almost all tablet formulations.

B. 1. Objective and Justification for the Present Study:

As discussed earlier, tablet disintegration has been increasingly viewed as an important factor in formulating pharmaceutical systems. Mechanisms by which tablet disintegrants function have been investigated.

The current concern about bioavailability of drug products has made formulators very selective in the use of disintegrants. The United States Pharmacopoeia is presently considering a radical extension of dissolution test requirements to most conventional tablets and capsules, and thus a number of formulators are now re-evaluating their formulations to see whether it is now appropriate to alter the identity or quantity of disintegrant (59).

The pharmaceutical industry has been using corn starch and guar gum extensively as tablet disintegrants for many years. Both disintegrants are natural source materials, but their function as a tablet disintegrant are not satisfactory. There is, however, a demand from certain segments of the pharmaceutical industry, particularly those concerned with vitamin formulations, which require a powerful disintegrant of natural origin.

Soy cellulose (Emcosoy), is an all-natural source material, derived from defatted soy beans by a special process. It contains no sugar or starch, and it has been given GRAS status (Generally Regarded

As Safe) by the United States Food and Drug Administration (60). Many formulators are not aware of the potential of this material because of the lack of authentic and extensive study. It is hoped that the present study will provide a reliable evaluation of soy cellulose (Emcosoy) as an effective substitute for corn starch and guar gum.

## II. EXPERIMENTAL

A. Equipment:

The following items were used:

Mettler K5 Balance, Mettler Instrument Corp., Hightstown, NJ.

Mettler H3 Balance, Mettler Instrument Corp., Hightstown, NJ.

Sieve Shaker, Fisher Wheeler, New Britian, CT.

Brookfield Synchro Lectic Viscometer, Model RVT, Stoughton, MA.  
(Food Science and Nutrition Department, URI).

Instron Universal Testing Instrument, Instron Corporation,  
Canton, MA. (Department of Mechanical Engineering,  
Wales, URI).

Scanning Electron Microscope, Cambridge Instruments, Cambridge,  
England (Graduate School of Oceanography, URI).

Twin Shell Dry Blender, Patterson Kelly, East Strousburg, PA.

U-tube type water uptake apparatus (Two water reservoirs are  
joined by rubber tube; one is to measure the swelling of  
disintegrant, and the other to measure the water uptake.  
(Fig. ).

Stokes Rotary B2 Tablet Press, Pennwault Corporation, Warminster,  
PA.

Erweka Hardness Tester, Erweka G. m. b. H. Frankfurt, West  
Germany.

Roche Friabilator, Erweka G. m. b. H. Frankfurt, West Germany.

U.S.P. Disintegration Apparatus, Scientific Glass Apparatus Co.,  
Bloomfield, NJ.

U.S.P. Basket Dissolution Apparatus, Precision Scientific Co.,  
Chicago, IL.

Dissograph (programmable automated dissolution test analyzer),  
Hanson Research Corp., Northridge, CA.

Perkin Elmer Hitachi 200 Spectrophotometer with recorder,  
Perkin Elmer Corporation, Norwalk, CT.

Hewlett Packard Calculator with plotter, Hewlett Packard  
Calculator Products Division, Loveland, CO.

Computer, Intel As/5, running IBM OS/MVT with HASP II, Ver-  
sion 3. (URI Academic Computer Center, Tyler Hall, URI,  
Kingston, RI 02881).

B. Materials:

The following chemicals were obtained from Amend Drug and Chemical Company, Irvington, NJ:

Riboflavin USP, Lot D 14687 M 23;

Niacin USP Lot D 15492 R 19;

Ascorbic Acid USP Lot D 15128023;

Pyridoxine Hydrochloride USP Lot F 15949B24;

Corn Starch USP Lot C 712551;

Sodium Lauryl Sulfate USP Lot D 14148 I 01.

The following materials were obtained from Edward Mendell Company, Carmel, NJ:

Calcium Phosphate Dihydrate (Emcompress) Lot No. 9229;

Soy Cellulose (Emcosoy) Lot No. SY 131 X;

Emdex (90 to 92% Dextrose + 3 to 5% Maltose + Higher Glucose) V 15;

Sodium Starch Glycolate (Emplotab) Lot No. C 192 X;

Other materials used in this study included:

Carmellose (Type A) (Ac-Di-Sol) - Lot No. 6379. FMC Corp., Philadelphia, PA.

Carmellose (Type B) (CLD II) - Lot No. 1026 BP. Buckeye Cellulose Corp., Memphis, TN.

Granulated Ascorbic Acid - Lot No. 230020; Roche Laboratories, Nutley, NJ.

Magnesium Stearate - Lot No. N 49483; Ruger Chemical Company, Inc., Irvington, NJ.

Stearic Acid, Ruger Chemical Company, Inc., Irvington, NJ.

Sta-Rx 1500 (modified starch) - Colordon, Inc., West Point, PA.

Polyplasdone XL (Cross-linked polyvinylpyrrolidone), GAF Corporation, NY.

NOTE: Emcompress, Emdex, Emcosoy, Ac-Di-Sol, CLD II, Explotab, Polyplasdone XL, and Sta-Rx 1500 are trade names. Trade names are used throughout the thesis due to their familiarity and convenience.



C. Methods:

There are several tests which can be used to evaluate disintegrants before using them in actual formulations. No one test is perfect; each has its advantages and disadvantages. The behavior of disintegrants alone could be vastly different compared to their behavior in compressed tablets. Some of these tests may, however, be useful as the basis of a raw material specification designed to control lot-to-lot variation.

( 59 ).

1. Scanning Electron Micrographs:

The role of particle morphology(size, shape and composition) in the production of tablets has long been discussed. This theoretical effort has found an ally in scanning electron microscopy (SEM), which has made it possible to obtain direct photographs of tableting excipients and finished compacts. This instrument provides scanning electron micrographs of a range of disintegrating agents, lubricants, and glidants in an attempt to further the understanding of the morphology of those substances and thereby to increase the comprehension of how the excipients function in tablet systems. ( 6 ).

Fig. ( 1 ) depicts the principle of the scanning electron microscope. The entire procedure was carried out as follows:

- a. **An** adhesive was placed on a metal stub, which was coated with gold or cadmium.
- b. A few disintegrant particles were sprinkled on the stub.

Working Principle of Scanning Electron Microscope :

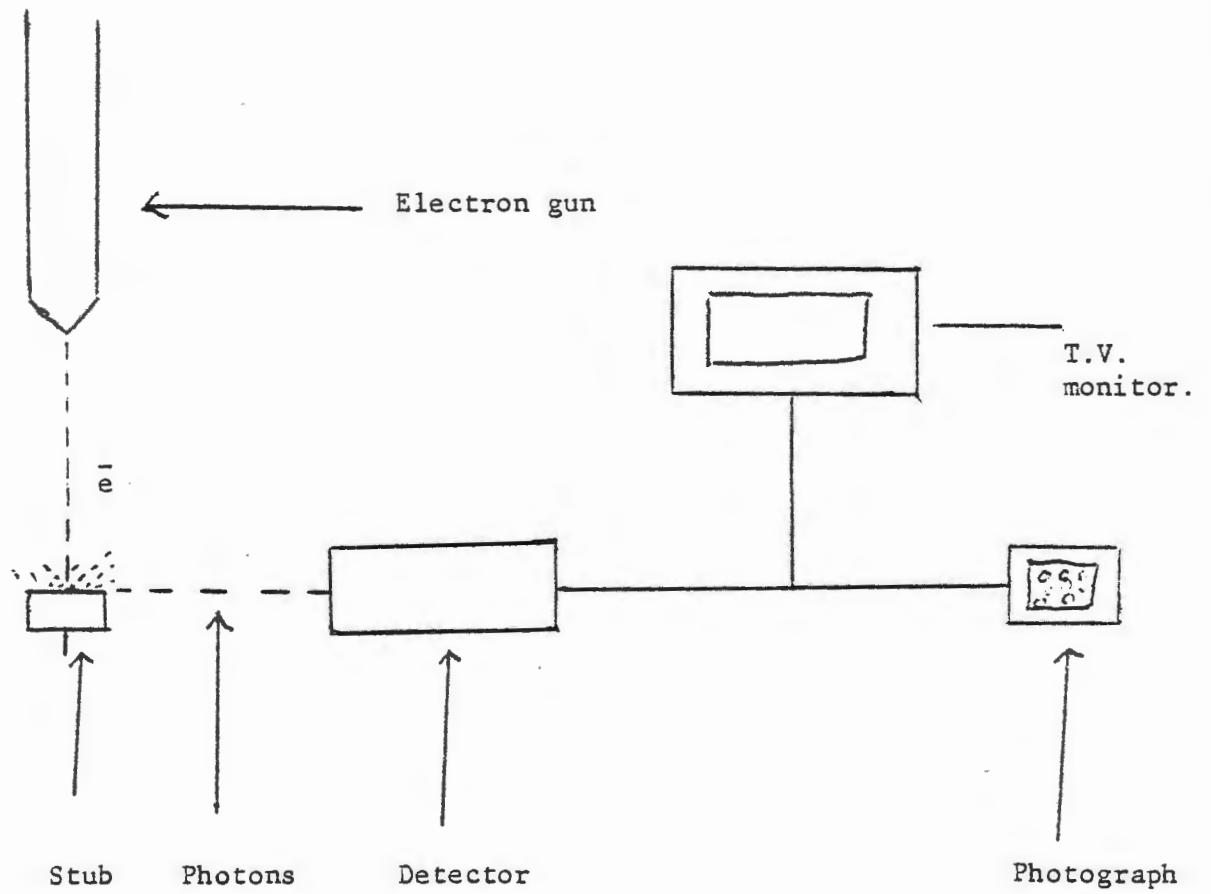


Fig. 1

c. The stub was placed in electron microscope and photographs were taken. The Emcosoy particles were magnified 200, 1070, 1100, and 2200 times to its original size.

The scanning electron microscope helps in elucidating the structure of individual disintegrant particles by magnifying them to thousands of times their original size (Fig. 4,5). (Note: The scanning electron microscopy work was performed by Mr. Donald Scales of the URI Graduate School of Oceanography).

## 2. Bulk Swelling and Water Uptake:

One relatively simple test used by many groups involved in the evaluation of tablet disintegrants is the quantification of the interaction of a bulk powder bed, composed of pure disintegrant, with water. There are several types of apparatus which can be used for this purpose. The one used in this study consisted of a calibrated glass reservoir (containing water or other appropriate fluid), connected at its base by a U-tube to a second glass reservoir which contained the disintegrant powder (Fig. 2). Values sought were the decrease in volume of water in the reservoir and the increase in size of the powder bed (the two values are not necessarily identical) as a function of time. The percentage of swelling and the percentage of water uptake

U-TUBE APPARATUS

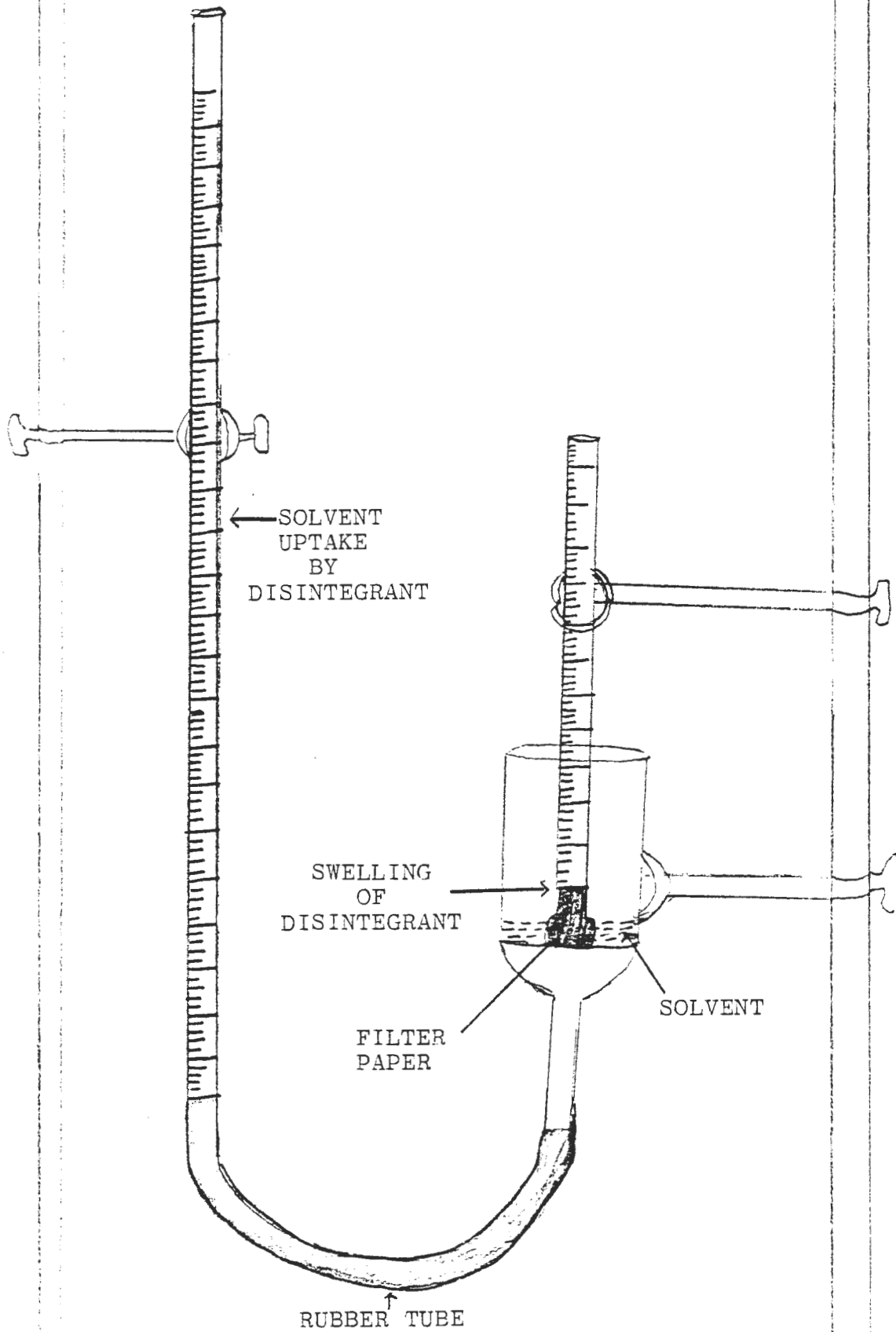


Fig. 2

$$\% \text{ Swelling} = \frac{100 ( (S_t - S_0) / S_0 )}{S_0} \quad (1)$$

where  $S_0$  is the disintegrant level in the tube at time zero, and  $S_t$  is the disintegrant level in the tube at any time  $t$ .

$$\% \text{ Water Uptake} = \frac{100 ( (V_t - V_0) / V_0 )}{V_0} \quad (2)$$

where  $V_0$  is the water level in the calibrated reservoir at time zero,

and  $V_t$  is the water level in the calibrated reservoir at time  $t$ .

Percentage of swelling and water uptake were plotted versus time. This test is helpful in determining the ease and extent of bulk powder interaction with water ( 61 ).

The disintegrants used for this test were Emcosoy, Ac-Di-Sol, corn starch, and CLD II. The solvents were distilled water, ethanol/water mixtures, and sodium lauryl sulfate solutions.

Air pressure was used to remove air bubbles from the lower funnel shape portion of the reservoir. The apparatus was left undisturbed for half an hour, so the water levels in both reservoirs remained constant. The readings were taken initially at short intervals, but later at an interval of twenty minutes.

### 3. Water Penetration of Disintegrants:

Although the mechanism of disintegration is very complicated and has not been completely determined (40), the penetration of liquid into a tablet is the first step in the process of tablet disintegration.

In this test the water penetration in various disintegrants was compared. The penetration of a liquid into a capillary is expressed by the following equation:

$$L^2 = \frac{R \gamma \cos \theta t}{2 \eta} \quad (3)$$

Where: L is the penetrated length of capillary

R is the radius of capillary

$\gamma$  is the surface tension of liquid

$\eta$  is the viscosity of solution

$\theta$  is the angle or contact

t is the time in seconds

The penetrated length of powder bed can be seen visually in a calibrated tube. Then the reading, which is in milliliters, can be converted into centimeters by multiplying it by 2.1, since 1 ml = 2.1 cm.

$$L^2 = \frac{R\gamma\cos\theta}{2\eta} \quad (4)$$

$$L^2 = kt \quad (5)$$

Where:  $k = R\gamma\cos\theta$  which is called penetrating rate constant.

$k$  varies with materials used; i.e., it depends on the average radius of void space  $R$ , and contact angle  $\theta$ .

The water temperature was about  $22 \pm 1^\circ\text{C}$ . The tube containing disintegrants was tapped ten times in all the cases. The water penetration was measured at intervals of twenty minutes.  $L^2$  vs.  $t$  was plotted.

#### 4. Energy of Wetting:

Penetration rate of water into disintegrant bed also depends on temperature of water (55). An attempt was made to find out approximate energy of wetting of Emcosoy. The water temperatures were 22, 29.2, and  $37^\circ\text{C}$  respectively. The temperature range was kept as wide as possible; temperatures included storage condition (room temperature), as well as temperatures of the mediums (for disintegration and dissolution tests).

The entire experiment was carried out the same way as discussed under water penetration of disintegrants. Nogami, et al, derived an equation from Andrade's equation:

$$\eta = Ae^{E/RT} \quad (6)$$

Where E is the activation energy of viscosity, A the constant, and R the gas constant. On the other hand, the adhesion tension,  $\gamma \cos \theta$ , depends on temperature at a constant pressure P, as:

$$\Delta H = \gamma \cos \theta - T \frac{(\delta r \cos \theta)}{(\partial T)} P \quad (7)$$

Where  $\Delta H$  is the heat wetting. If systems are not disturbed during the wetting process,  $\Delta H$  is independent of temperature. In this experiment, pressure p is considered constant; solving above equation.

$$\gamma \cos \theta = \Delta H - CT \quad (8)$$

is obtained, where C is the positive constant; combining all above-mentioned equations:

$$k = \frac{\gamma}{2A} (\Delta H - CT) e^{-E/RT} \quad (9)$$

Since generally  $\Delta H/CT$  is coming between one and two, taking logarithm of equation, approximating  $\log (\Delta H/CT - 1)$  to  $\Delta H/CT + C''$

$$\text{Log } \frac{k}{T} = \alpha - \frac{\beta}{RT} \quad (10)$$

$$\text{Where } \alpha = \text{Log } C + C'' , \beta = E - \frac{R}{C} \Delta H$$

$L^2 = kt$  we discussed under water penetration of disintegrants.

We can obtain value of k from slope of each straight line and then plotting  $\log k/T$  vs.  $\frac{1}{T}$ . Slope =  $\beta/R$  we can calculate the value of  $\beta$  -- energy of wetting by multiplying slope by R (1.987 cal. deg.<sup>-1</sup> mole<sup>-1</sup>).



#### 5. Wetting and Drying of Disintegrants:

The effect of wetting and drying was observed on Emcosoy. Emcosoy was wetted with excess of water in a petri dish (diameter 22 cm). Then the first batch was dried at room temperature (21<sup>C</sup>); second and third batches were dried inside the oven at 40<sup>C</sup> and 60<sup>C</sup> respectively. The cake-like mass that formed on drying was reduced to fine particles by using a Fitzpatrick Comminuting Machine. The sieve, #00, was placed at collecting end of machine. The particles received through the "00" sieve passed through the 100 mesh (U.S. Standard) screen.

The bulk swelling was determined by using the U-tube-type apparatus for all the batches. The batch which was deemed most promising in all respects was compressed as formulation IV. Tablets from both batches were made at identical press settings, using original Emcosoy for comparison purposes. Tablets from both batches were compared for weight, thickness, hardness, and disintegration time.

#### 6. Aqueous Dispersion:

Emcosoy (2g) was dispersed in 100 ml of dilute hydrochloric acid solution (1 in 100, pH .70), and in 100 ml water in two measuring cylinders. It was left overnight with periodic stirring initially. The disintegrant settled at the bottom, the volume of disintegrant was measured in the beginning and at the end to find out the percentage of swelling.

The measuring cylinder provides larger room for the swelling of disintegrant particles, compared to calibrated tube used in bulk swelling study. The medium was kept acidic to simulate effect of gastric pH on the swelling of the disintegrant. In another cylinder, the effect of water on swelling of disintegrant was observed. The percentage of swelling of disintegrant in neutral and acidic medium was compared.

7. Comparative Study of the Viscosity of Different Strength Solutions on Various Disintegrants:

Determination of the viscosity of solution or gel (disintegrant and solvent interaction) is important because:

- a. a viscous gel that sometimes forms on the surface of a disintegrating tablet due to solvent disintegrant interaction, impedes the further penetration of water into the tablet and thus prolongs the process of disintegration.
- b. The contribution of disintegrant in increasing the viscosity of dissolution medium is important. Increase in viscosity increases the diffusion layer thickness value (Diffusion Layer Model) and thus slows down the dissolution rate. According to Noyes-Whitney Law:

$$\frac{dc}{dt} = \frac{KDS}{Vh} (C_s - C) \quad (11)$$

Where:  $\frac{dc}{dt}$  is the rate at which drug is dissolving

K is the Constant

D is the Diffusion Coefficient

S is the Surface Area of Particle

V is the Volume of Solution

h is the Thickness of Diffusion Layer

( $C_s - C$ ) is the Conc. Gradient

The equation can be further simplified:

$$\frac{dc}{dt} = K's (C_s - C)$$

$$\frac{dc}{dt} = K's C_s$$

$$\text{Where } K = \frac{KD}{Vh}$$

Where C is very small

Particle  $C_s \xrightarrow{h} C$

Stirring reduces the value of h. If we have two solutions, A and B, A has a higher viscosity than B; then stirring will reduce the value of h in B easily and dissolution process will be enhanced. Thus, viscosity plays an important role in dissolution and ultimately in the bioavailability of the product. Although the overall impact may be small, it still may be an important factor.

Solutions of different disintegrants were prepared from 2% to 16% by dispersing the disintegrant in water at 37°C. The resulting solutions were stirred with glass rod to mix the disintegrant and water thoroughly. The viscosity of the solution was determined by using a Brookfield viscometer.

$$\text{Shear rate } (\gamma) : \frac{2\pi R \cdot \text{rps}}{\delta} \quad (12)$$

$$\text{rps} = \text{rpm}/60$$

$$\text{Shear Stress } (\tau) : \frac{F\%M}{R^2 \cdot 2\pi \cdot L} \quad (13)$$

$$\text{Viscosity} = \tau/\gamma \text{ (Poise)}$$

R is the radius in the spindle.

$\delta$  is the portion of the solution which is subjected to stress.

F is 7,187 dyne-cm

L is the height of spindle in a solution.

% is the torque reading.

Spindle: Big to small (1 to 7)

Twenty, fifty, and a hundred rpm were used. The dial reading was taken when the pointer had completed one full return.

The easiest method of calculating viscosity is to use the Brookfield Viscometer Scale, which is applicable if the experimental conditions comply with those mentioned by the manufacturer. In this study, the scale was used because the experimental conditions were well within the realm of those mentioned by the manufacturer.

The rheological pattern of the different disintegrant solution was studied. Their viscosities were compared.

#### 8. Maximum Hydration Capacity of Various Disintegrants:

The concentration of disintegrant in a hundred milliliter was increased to a point where there was not enough water to wet all

the disintegrant particles. The solution was stirred with a glass rod to disperse the disintegrant. Maximum hydration capacity of Ac-Di-Sol, Emcosoy and CLD II was compared.

9. Emcosoy-Sieve Analysis:

Sieving is a commonly used method for measuring particle size distribution because it is inexpensive, simple, and rapid with little variation between operators, provided that similar procedure are followed. The procedure involves the mechanical shaking of a sample through a series of successive smaller sieves, and the weighing of the portion of the sample retained on each sieve. The type of motion influences sieving vibratory motion is most efficient, followed successively by side-top motion, bottom-top motion, rotary motion with top, and rotary motion.

One hundred gram powder was placed on the top sieve, and the mechanical shaking was continued for 30 minutes. The portion of the sample retained on each sieve was weighed. The results obtained were tabulated (Table XXV), and the average diameter of Emcosoy particle was calculated as shown in the table.

10. Plastic Deformation and Elasticity of Various Disintegrants:

Plastic deformation of starch grains under high pressure has been reported by a number of investigators (62, 63). Starch grains are generally thought to be elastic; in fact, most materials are elastic up to a certain extent. Elastic grains deformed under

pressure tend to return to their original shape and size when the deforming pressure is removed. However, it has been suggested that compression may cause more permanent deformation, which is termed, "plastic deformation." These deformed starch grains are energy rich and this energy is released when the grains are exposed to water ( 64 ). The energy-rich starch grains swell rapidly in water, unlike undeformed grains, which require more heat in order to swell. Recently published electron photomicrographs of potassium chloride and aspirin tablets clearly show deformed starch grains both on the external surfaces and in cross-sections of tablets ( 63). These grains returned to their original shape when the tablets were exposed to moisture.

We can classify various disintegrant particles into two categories: those permanently deformed by the compression (in other words, these particles are nonelastic particles); and those much less permanently deformed by the compression (elastic particles).

In the past, the scanning electron microscope was used for gaining insight into the behavior of solids under compression. It is a powerful tool; however, it does not express elasticity in any arithmetic numbers, so it is almost impossible to compare plastic deformation elasticities of various disintegrants.

The Instron Universal Testing Instrument, which is widely used in food science to study textural properties of cooked comminuted fish muscle ( 65 ), was used. This equipment is also used in textile industries, packaging industries, and rubber industries. It evaluates properties in terms of shear strength, compressive strength, modules of elasticity, resilience, and puncture strength of the material.

This machine was used in the present study for determining deformation of the disintegrants, as well as compressive force needed to achieve those deformations. Modules of elasticity, percentage of plastic deformations and percentage of recovery were also calculated. A few accessories were designed to use the machine for handling powder-like materials.

The disintegrant **was** placed in a cylinder which had one end left open; the other end was closed by a removable bottom. These removable bottoms were designed to facilitate the removal of compressed powder. The piston, which was attached to a load cell, could move upward or downward freely, into the cylinder through the open end. The load cell has a capacity to apply compressive force up to 20,000 lbs. The load cell was connected to controlling panels which had a digital dial that showed the distance traveled by the load cell. It also had switches which regulated upward or downward movement of a loading cell, as well as the distance that would be traveled by the loading cell. There

was a dial which could be rotated to regulate the compressive force of the loading cell. The movement of the loading cell with the piston could be synchronized with the movement of the chart paper.

The chart paper had ten big divisions, and each big division was divided into ten small divisions; so the total number of small divisions was one hundred. A mobile recorder pen moved along the base line of the paper, with the movement of piston. When the piston applied pressure on the disintegrant particles, the pen moved on the calibrated chart paper; so by counting the small divisions, one can determine the pressure applied by the piston.

The piston can move down and up once or more times; that makes it possible to apply single compression or cyclic compression.

The data was interpreted as follows:

$$\begin{aligned} \text{Deformation distance} &= \text{Load Cell} \div \text{Chart} \\ &= 5 \text{ in/min.} \times 1/2 \text{ min/in.} \\ &= .25 \text{ load cell/chart} \end{aligned}$$

$$\begin{aligned} \text{Modulus of Elasticity} &= \frac{\text{Compressive Stress}}{\text{Corresponding Strain}} && (15) \\ &= \frac{F/A}{\Delta L/L} \end{aligned}$$



Where F is Compressive Force

A is the area of the piston which comes in contact with powder.

$\Delta L$  is the reduction in the height of disintegrant.

L is the original height of disintegrant in a cylinder.

Now, for example, if pen travelled 9.2 divisions on chart paper:

$$25 \times \frac{9.8}{10} = 0.23 \text{ inch } (\Delta L)$$

$$\frac{\Delta L}{L} = \frac{.23}{2.25} = .102$$

$$\begin{aligned} \frac{F}{A} &= \frac{\text{Compressive Force}}{\text{Area of Piston ( } E^2 \text{)}} \\ &= \frac{2.85 \times 10^3 \text{ (From Chart)}}{3.14 \times (0.55)^2} \\ &= 2850 = 956.38 \text{ lb/in}^2 \end{aligned}$$

$$\begin{aligned} \text{Elasticity} &= \frac{F/A}{\Delta L/L} \\ &= \frac{956.38 \text{ lb/in}^2}{.1022} \\ &= 9357.93 \text{ lb/in}^2 \end{aligned}$$

How to achieve desired deformation (if you want to attempt 10% deformation).

The height of disintegrant in a cylinder is 2.25 inches.

10% of this height = 0.23 inch.

When the piston touched the powder in a cylinder, the reading was adjusted to zero. Then the distance 0.23 inch (as calculated above) would be set on the scale and the "down" switch would be pressed. The pis-

ton would travel 0.25 inch in a powder bed, and the compressive force needed to travel that distance would be recorded on a chart paper. 10%, 20%, and sometimes 40% deformations were attempted.

$$\text{Plastic Deformation:} = \frac{D_3 - D_1}{\text{Sample Height}}$$

$D_1$  is the distance travelled by piston during cycle one.

$D_3$  is the distance travelled by piston during third cycle.

If  $D_1 = 0$

and  $D_3 = 11$  div. on chart,

$$D_3 = 11 \times \frac{.25''}{10 \text{ div.}} = .275$$

$$\begin{aligned} \% \text{ Plastic deformation} &= \frac{D_3 - D_1}{\text{Sample Height}} \\ &= \frac{.275 - 0}{2.25} \times 100 \\ &= 12.22\% \end{aligned}$$

$\% \text{ Recovery} = 20 - 12.22$

$$= 7.78\%$$

The height of the disintegrants were adjusted to 2.25 inch after tapping. Modulus of elasticity values at various attempted deformations and the compressive force needed to achieve that deformation were compared for various disintegrants. Percentages of plastic deformation and percentages of recovery were also compared for various disintegrants.

11. Formulations:

Formula I:	Riboflavin	0.57%
	Pyridoxine hydrochloride	0.86%
	Niacin	5.70%
	Ascorbic acid	17.10%
	Disintegrant*	0.5% or 1.0% or 3.0%
	Stearic acid	2.5%
	Emcompress <u>or</u> Emdex ad	100%
	Tablet weight 350 mg.	
Formula II:	Pyridoxine hydrochloride	5.0%
	Disintegrant*	0.5% or 1.0% or 2.0% or 3.0%
	Magnesium stearate	0.5%
	Emcompress <u>or</u> Emdex ad	100%
	Tablet weight 200 mg	
Formula III:	Ascorbic acid	25.0%
	Disintegrant*	0.5% or 1.0% or 2.0% or 3.0%
	Stearic acid	2.5%
	Emcompress ad	100%
	Tablet weight 400 mg.	

\*Emcosoy, Explotab, Ac-Di-Sol and Corn Starch were used as disintegrants.

Formula IV:	Hydrochlorothiazide	20.0%
	Disintegrant ** A or B	1.0%
	Magnesium stearate	0.5%
	Emcompress ad	78.5%
	Tablet weight 200 mg	

\*\*Disintegrant A is original Emcosoy  
Disintegrant B is modified Emcosoy

## 12. Tablet Production

The active ingredient (or ingredients) was mixed with matrix in a twin shell dry blender for five minutes. The disintegrant was then added and mixing was continued for five minutes. After completion of this blending, stearic acid in Formulas I and II and magnesium stearate in Formula II was added, and **the mixing process was** continued for an additional five minutes. The formulations were compressed on a Stokes Rotary tablet press. A punch size of 3/8" for multivitamin and ascorbic acid formulations and punch size of 1/4" for pyridoxine formulations were used.

Disintegrants were studied in the presence of single active ingredient Formula II (pyridoxine hydrochloride) and Formula III (ascorbic acid), and more than one active ingredient (multi-vitamin formulation) Formula I, using water insoluble (Emcompress) and water soluble (Emdex) matrices. Tablets were made using various disintegrants with identical press settings. Formula I contained riboflavin, pyridoxine hydrochloride, niacin, and ascorbic acid as active ingredients. Emcosoy, Ac-Di-Sol, and Explotab were used as disintegrants at 0.5% and 1% level, and Emcosoy and corn starch at 3% level. Emcompress and Emdex were used as matrices. Stearic acid was used as lubricant at 2.5% level.

Formula II contained Pyridoxine hydrochloride as active ingredient. Emcosoy, Ac-Di-Sol, and Explotab were used as disintegrant at 0.5%, 1%, and 2% level. Corn starch and Emcosoy were used

as disintegrant at 2% and 3% level. Magnesium stearate was used as lubricant at 0.5% level. Emcompress and Emdex were used as matrices.

Formula III contained ascorbic acid as active ingredient and stearic acid as lubricant at 2.5% level. Emcosoy, Ac-Di-Sol, and Explotab were used as disintegrant at 0.5%, 1%, and 2% level. Corn starch and Emcosoy were used at 3% level. At 2% disintegration level, granulated ascorbic acid was used, while the rest of the formulation contained ascorbic acid USP.

Formulas I and III contained ascorbic acid as active ingredient; stearic acid (2.5%) was used as a lubricant, since alkaline lubricant, such as magnesium and calcium stearate and minerals; e.g., talc and hydrated silico aluminate, were shown to decrease the stability of ascorbic acid tablets (66,67).

#### Physical Tests:

Various physical tests were performed on the tablets:

- a. Weight: The weight of each individual tablet was determined by dusting each tablet off with a camel-hair brush, and placing it on an electric balance. This procedure was repeated for twenty tablets. The data from the tablets was analyzed for sample mean and Relative Standard Deviation (R.S.D.), using a computer program. This procedure is explained below. The R.S.D. value was used as

a measure of weight variation within a sample of tablets. The physical tests were run for twenty trials on each sample set. The data from the sample set was analyzed for mean value and Relative Standard Deviation (R.S.D.). A digital computer program written in Fortran was used to calculate these values, using the equation:

$$\text{Relative Standard Deviation} = \text{RSD} = 100 (S/\bar{x})$$

Where: S is the Standard Deviation of Sample, and

$\bar{x}$  is the Sample Mean

The computer program used appears in Appendix

- b. Thickness: The thickness of twenty tablets was determined by first dusting off the surface of the tablet with a brush, and then placing it in the jaws of a micrometer. The measurements were recorded and analyzed for mean value and R.S.D. as a measure of variation.
- c. Hardness: The hardness of twenty tablets was determined by placing each tablet in an electronic hardness tester (Erweka) which recorded the breaking strength of the tablet in kilograms. This procedure was repeated twenty times, and the data were analyzed for sample mean and R.S.D. as a measure of variation.

- d. Friability: This test is a measure of abrasion resistance and was determined by first weighing twenty tablets after dusting, placing them in a Roche friabilator which rotated at a fixed rate for twenty minutes. The total weight of the twenty tablets (or what was left) was recorded, and the percent friability was determined as follows:

$$\text{Friability} = 100 \cdot \frac{\text{Weight (original)} - \text{Weight (Final)}}{\text{Weight (original)}} \quad (17)$$

- e. Disintegration: Tablet disintegration was tested by using the U.S.P. apparatus with discs as described in the National Formulary XIV (68). Fig.3 schematically illustrates how this apparatus is set up. The time needed for all the fragments to pass through the mesh at the bottom of the test cage was noted visually, and was recorded. A mean value for six tablets was calculated, and the range of values was also recorded.
- f. Dissolution: Drug dissolution was measured using a U.S.P. apparatus, and according to monographs in U.S.P. XX. The exact equipment used was the U.S.P. "basket" assembly, and 5 ml samples were removed at times of 1, 2, 4, 7, 10, 20, 40, 60, minutes, etc. until 100% dissolution was observed. Plots of these readings were made to depict the dissolution process.



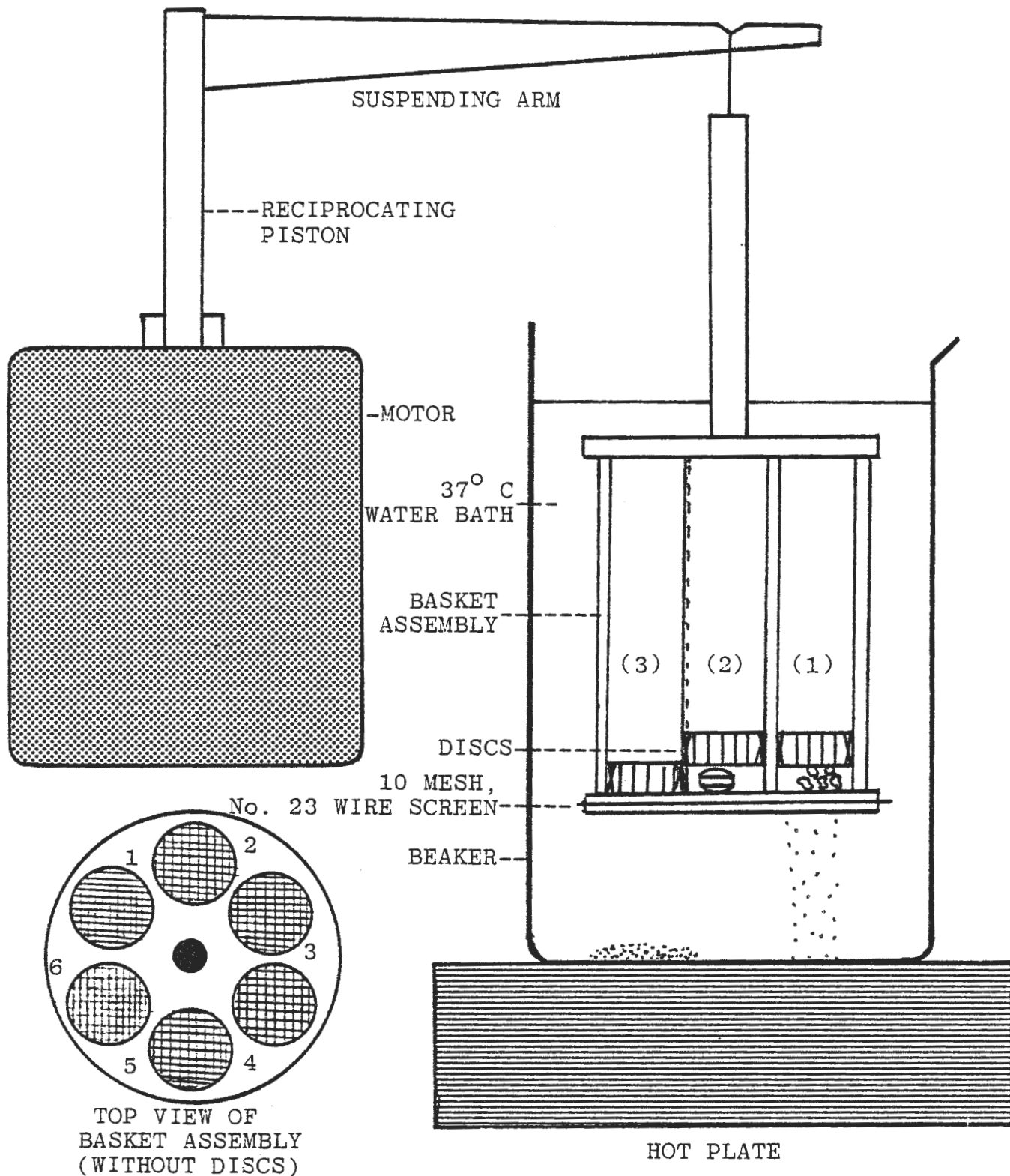


Fig.3 -The U.S.P. Disintegration Apparatus, In Use.

(2)-no disintegration, (1)-in process of disintegration, (3)-disintegration completed.

- i. Dissograph: This is a programmable dissolution test apparatus. It has the capacity to monitor the dissolution of six tablets simultaneously. Sample solution was pumped from dissolution flask to a cuvette cell. Dilution was carried out if needed. Then samples were analyzed in a spectrophotometer. Readings were recorded on the Dissograph scale. The sampling time can be programmed according to convenience. The significance of this apparatus is that all the functions are controlled automatically. The Dissograph was used to analyze the dissolution profiles of multi-vitamin formulations.
- ii. Analytical Procedures: The dissolution medium was selected as appropriate for any given drug, on the basis of stability and solubility of drug. The drug or vitamin solution was scanned using an ultra-violet spectrophotometer to find  $\lambda_{\max}$  -- the wavelength at which the solution gives maximum absorbance. Once this value was determined, the spectrophotometer was set at this wavelength, and each sample was analyzed for absorbance at this wavelength at various concentrations; the results were recorded in a Beer's plot. The drug monograph, as it appeared in the U.S.P. XX, was used as a reference for

determining the dissolution medium for the test.

The dissolution samples (five milliliters) were not returned to the dissolution bath so that there was a certain amount of drug being lost in each sample. This drug loss was reflected in subsequent samples. This loss which must be accounted for was corrected by using a digital computer program utilizing the following equations to determine the amount of drug in the five milliliter sample:

$$\text{Conc.} = (A-b)/m \quad (18)$$

Where: A is the absorbance

m is the slope of Beer's plot

b is the intercept of Beer's plot

The computer program appears in Appendix

### III. RESULTS AND DISCUSSIONS

#### 1. Scanning Electron Micrographs:

The scanning electron micrographs of Emcosoy are shown on pages 51 and 52. As can be seen in figures 4, 5 the flake-like material appears to consist of the ground cell walls of soy beans and probably contains some proteinaceous material. It swells in the presence of water, and the cellular nature of the soy polysaccharides appears to exert a wicking effect (20). This mechanism of tablet disintegration is so effective that Emcosoy can be used as a tablet disintegrant in concentrations as low as .5%.

#### 2. Bulk Swelling and Water Uptake:

Although investigators have proposed and evaluated numerous compounds as tablet disintegrants, relatively few are in common use today; only modified starches, cellulose derivatives, and some ion-exchange resins are widely used. Depending on the type of disintegrant used, many theories relating to mechanism of action of tablet disintegration have been proposed, but none has fully explained the disintegrant properties of all the agents. Lowenthal, in two review articles, had discussed in detail the different mechanisms and factors affecting tablet disintegration (19,70). The three major mechanisms of action are: (1) swelling, (2) porosity and capillary action (wicking), and (3) deformation. Even though these concepts are listed separately, interrelationships probably occur in almost all tablet disintegration.

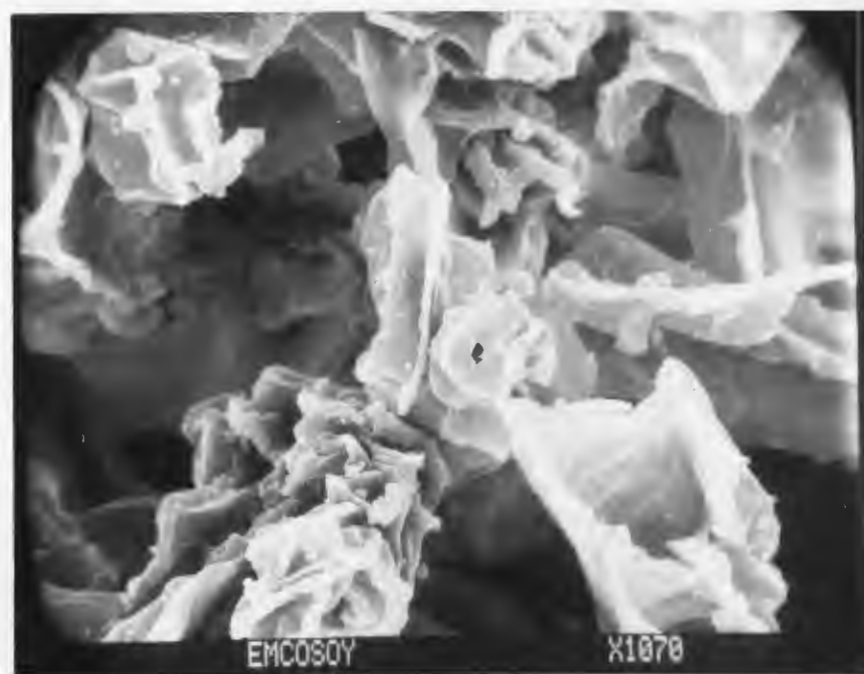
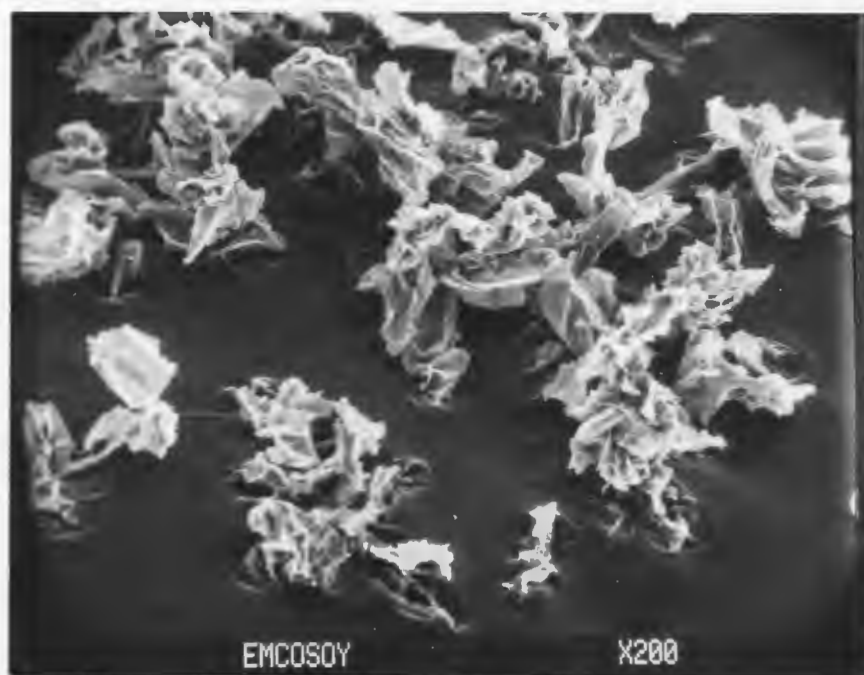


Fig 4

SCANNING ELECTRON MICROGRAPHS OF EMCOSOY PARTICLES

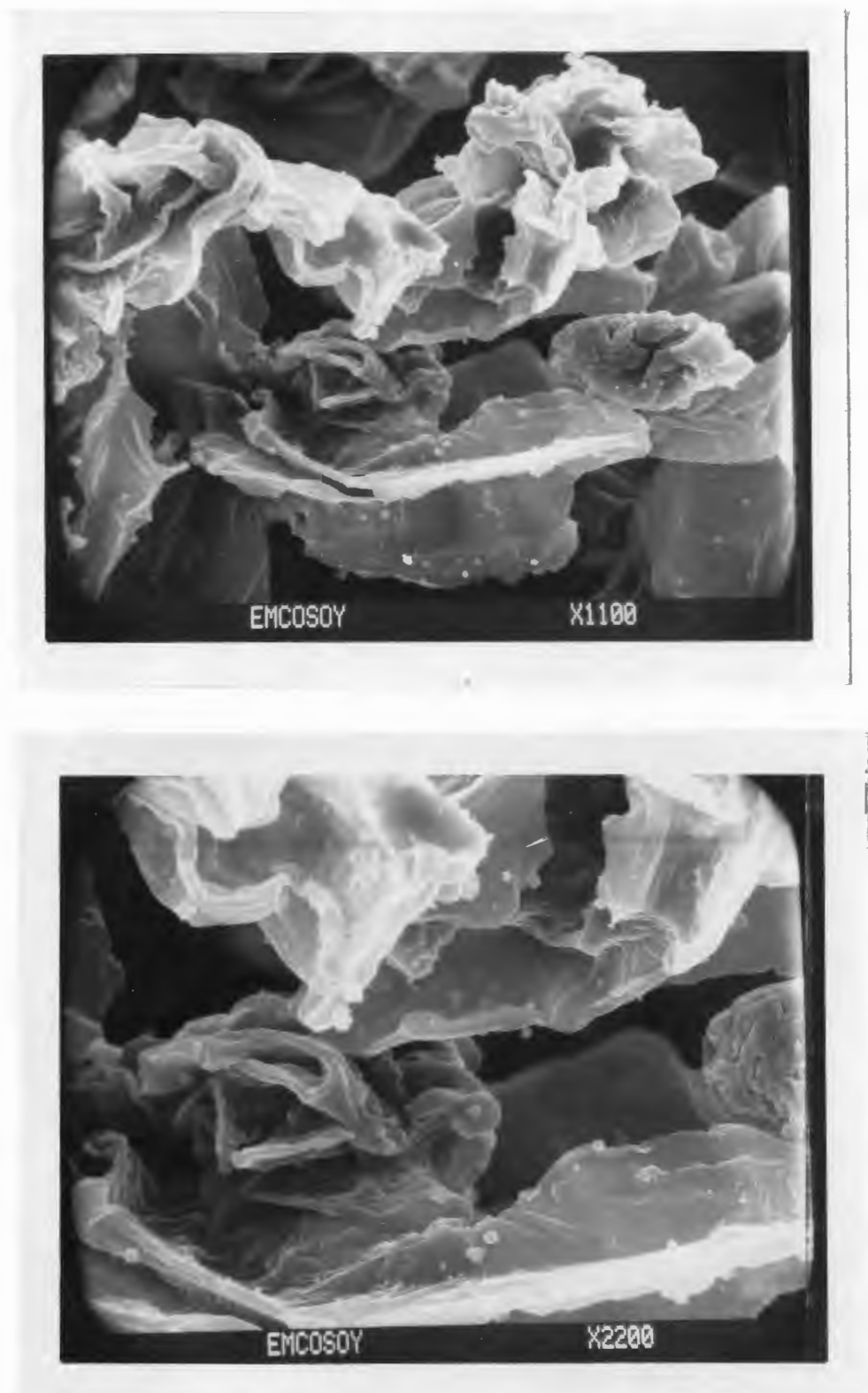


Fig. 5

As explained in the experiment section, U-tube type apparatus was used for determining the bulk swelling of various disintegrants. Bulk swelling of Emcosoy, Ac-Di-Sol, CLD and corn starch were determined. All the disintegrants had equal initial volume in the calibrated tube, since the volume difference was used in calculating percentage swelling.

For one disintegrant, different initial weight would give different percentages of swelling as found out in Ac-Di-Sol, Table IV, and graph on page 54. Logarithm of percentage swelling vs. time would give almost a straight line (Fig. 7 ). For different initial weight, the amount of water that would travel in a powder bed would vary (Fig. 8 ). The amount of water travel would reduce as the weight of the disintegrant increased.

The percentage of swelling decreased with increased initial weight of the disintegrant because the particles in the bottom portion of the tube came in contact with water of the reservoir first; they tried to swell, but their swelling was reduced by the weight of particles on the top. As the weight of the disintegrant was increasing, the bulk of particles on top was increasing; at the same time the amount of particles which were in contact with water remained the same. So, the swelling was reduced with increases in weight of the disintegrant.

Average bulk swelling of Emcosoy, Ac-Di-Sol, CLD, and corn starch were compared. Time required to reach the swelling and amount of water travelled in various disintegrants were compared (Table I ). As it can be seen from the table, Emcosoy had maximum swelling, while corn starch had no swelling. Time required to achieve that swelling

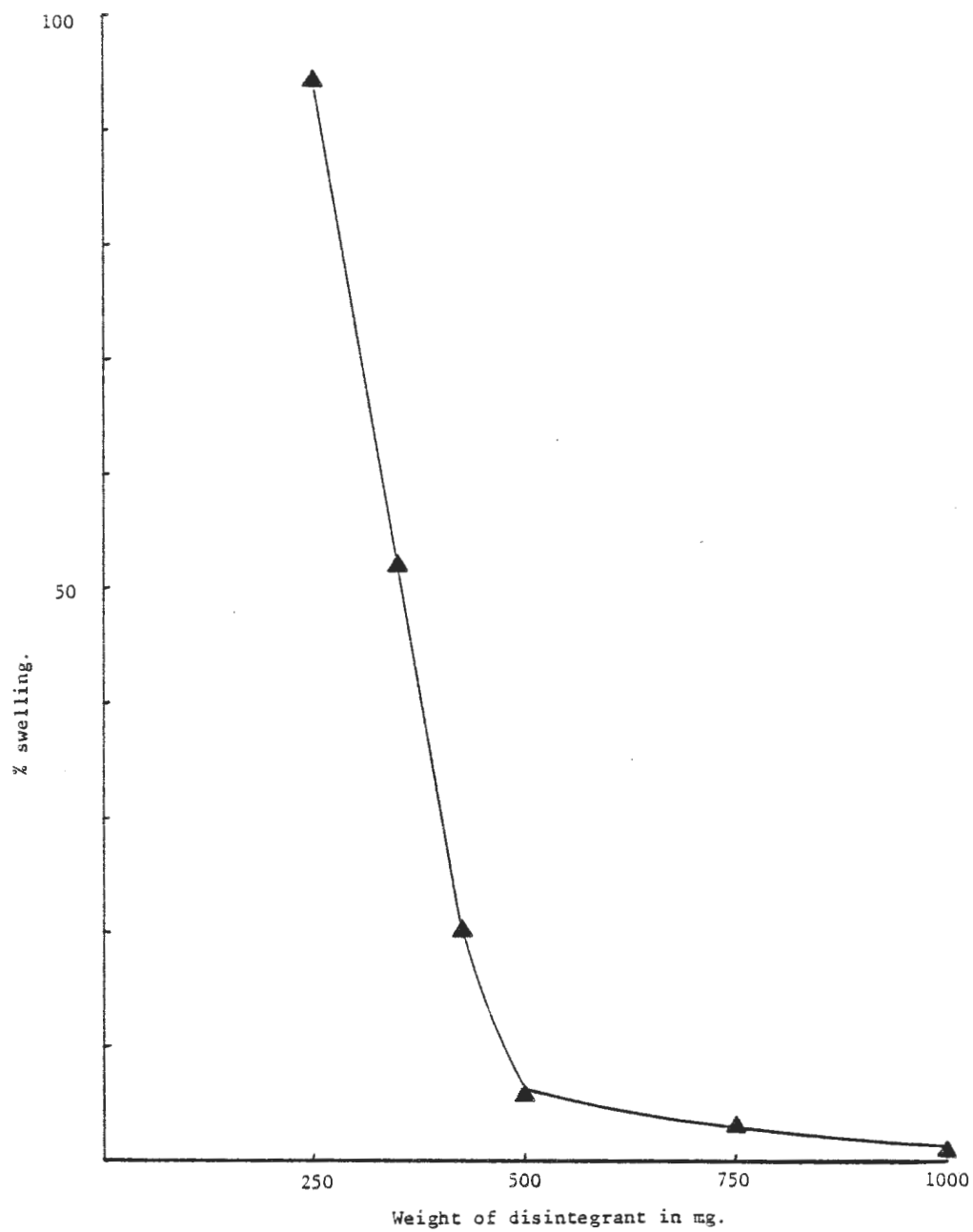


Fig. 6



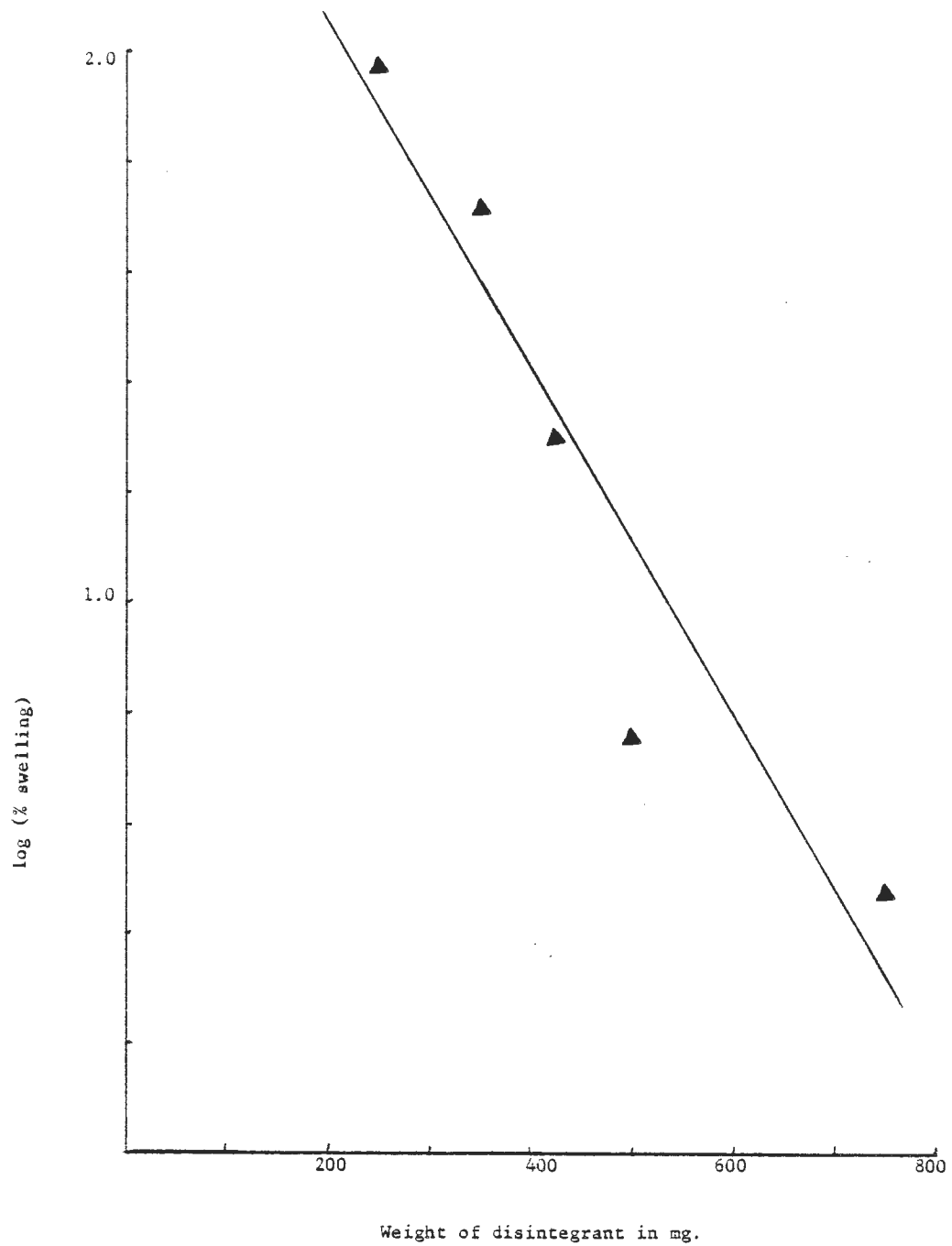


Fig. 7

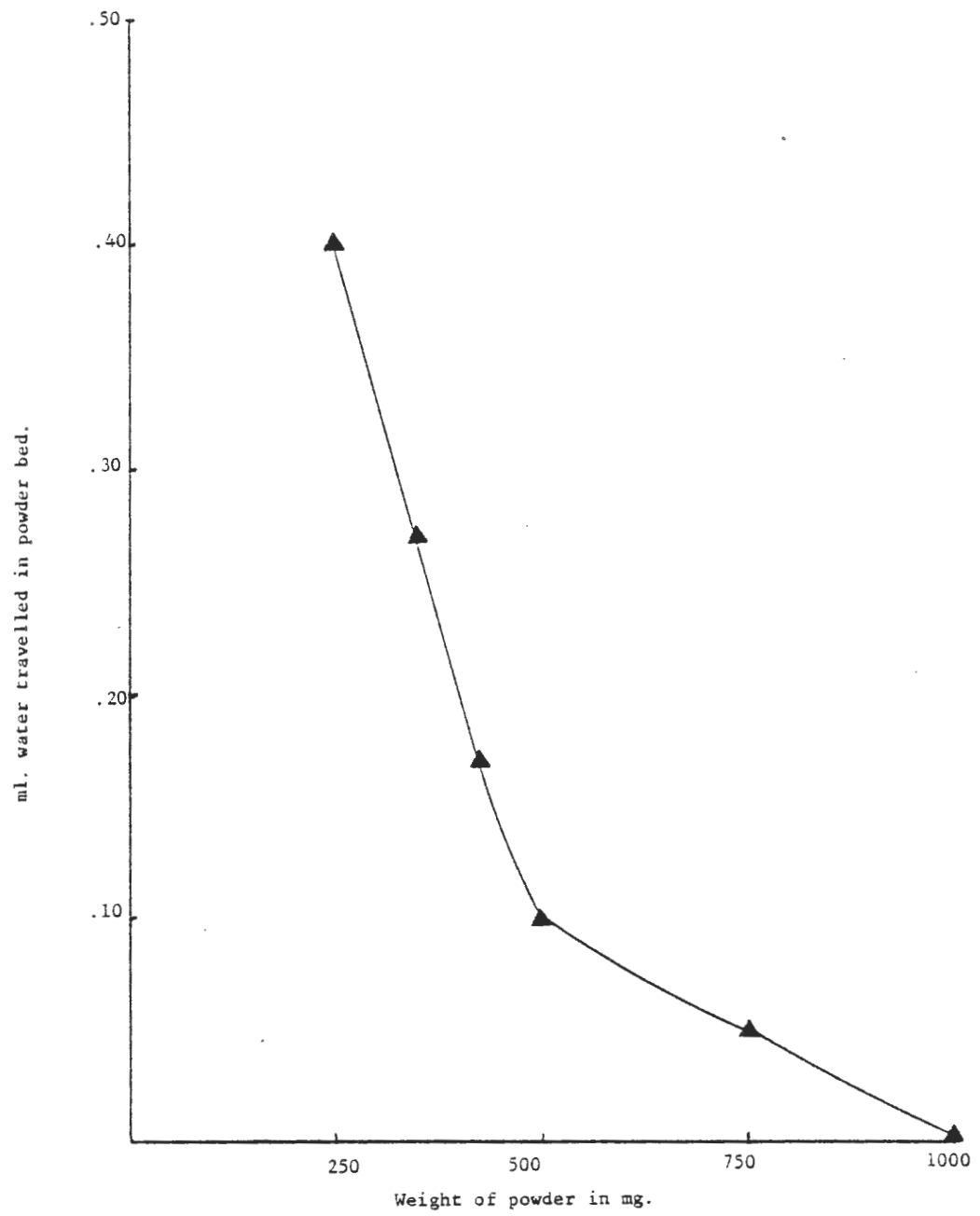


Fig. 8

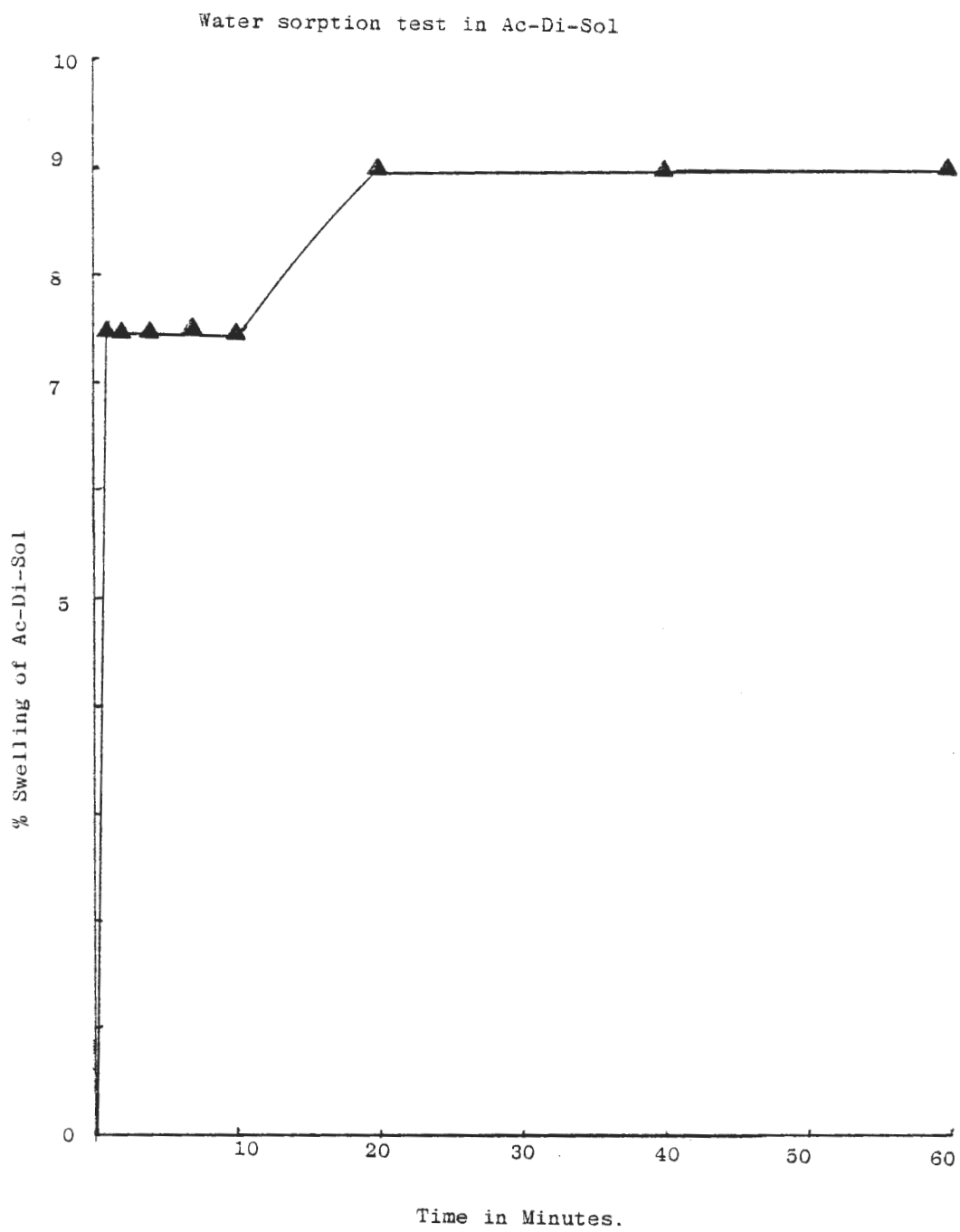


Fig. 9

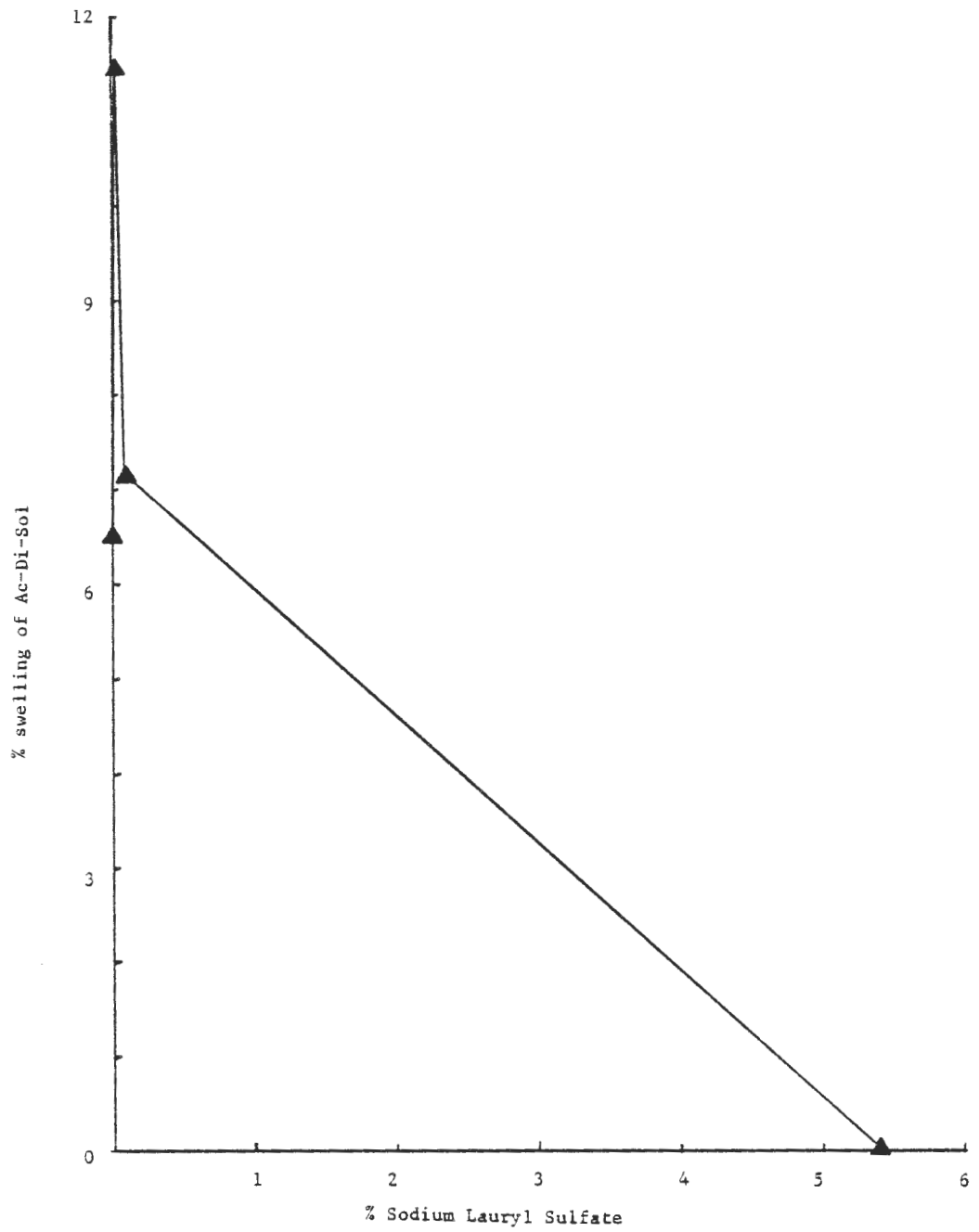


Fig. 10

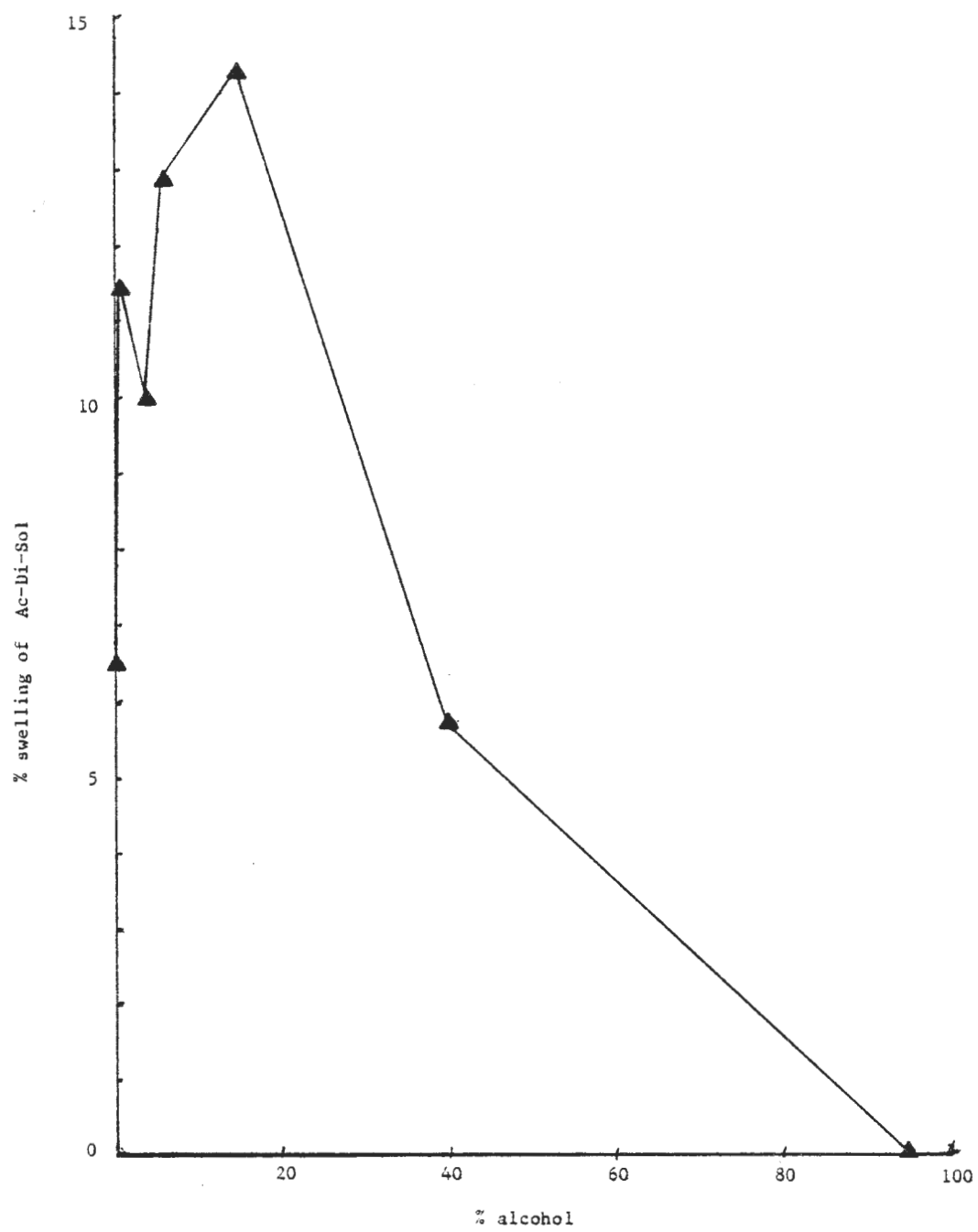


Fig. 11

TABLE I

Disintegrant	Time Required to Reach Plateau	Water Travelled in Powder Bed	% Swelling
CLD	10 min.	.3 ml.	48.57
Emcosoy	90 min.	1.04 ml.	55.71
Ac-Di-Sol	20 min.	.10 ml.	8.96
Corn Starch	6 min.	.70 ml.	nil.

TABLE II

Disintegrant: Emcosoy

Solvent: Citrate-Phosphate-Borate/Hcl buffer

PH	% Swelling of Disintegrant	Average Swelling
2	00.00	5.00
	10.00	
4	14.29	14.29
	14.29	
6	28.57	22.14
	15.71	
8	22.86	22.86
	22.86	

TABLE III

t Test for Buffer Reading

Buffer PH	5 Computer	t Table Value	Comment
2			
4	1.85	4.3	Not Significant
2			
6	2.10	4.3	Not Significant
2			
8	3.57	4.3	Not Significant

TABLE IV

<u>Disintegrant:</u>	Ac-Di-Sol	
Solvent:	Distilled Water	
Weight of the Disintegrant (mg)		% Swelling
<hr/>		
250		94.29
351		52.00
425		20.00
500		5.71
750		3.00
1000		0.77
<hr/>		

TABLE V

Disintegrant: Emcosoy

Solvent: Alcohol

Solvent	% Swelling	Average % Swell
0.0% alcohol	56 48	52
1.0% alcohol	43 43	43
4.0% alcohol	28 28	28
6.0% alcohol	30 28	29
15.0% alcohol	33 36	34.5
40.0% alcohol	23 23	23
95.0% alcohol	0.0 0.0	0.0



TABLE VI

t-test for Alcohol Reading with Emcosoy as Disintegrant

Alcohol %	t Computed	t Table Value	Comment
0.0 1.0	2.25	4.3	Not Significant
0.0 4.0	6.00	4.3	Significant
0.0 6.0	5.58	4.3	Significant
0.0 15.0	4.09	4.3	Not Significant
0.0 40.0	7.25	4.3	Significant
0.0 95.0	13.00	4.3	Significant

TABLE VII

Disintegrant: EmcosoySolvent: Sodium Lauryl Sulfate (SLS)

Solvent	% Swell	Average % Swell
0.0% SLS	56 48	52
0.01% SLS	43 38	40.5
0.10% SLS	44 58	51
5.40% SLS	14 8	11

TABLE VIII

t Test for SLS Reading with Emcosoy as a Disintegrant

SLS%	t Computed	t Table Value	Comment
0.00			
0.01	2.44	4.3	Not Significant
0.00			
0.10	0.12	4.3	Not Significant
0.00			
5.40	8.2	4.3	Significant

TABLE IX

Disintegrant: Ac-Di-SolSolvent: Citrate-Phosphate-Borate/Hcl Buffer

PH	% Swelling of Disintegrant	Average Swelling
2	17.14 21.43	19.29
4	14.29 10.00	12.15
6	21.21 15.71	18.46
8	8.57 17.14	12.86

TABLE X

t Test for Buffer Reading

Buffer PH	t Computed	t Table Value	Comment
2 4	2.35	4.3	Not Significant
2 6	.23	4.3	Not Significant
2 8	1.34	4.3	Not Significant

TABLE XI

Disintegrant: Ac-Di-SolSolvent: Sodium Lauryl Sulfate (SLS)

Solvent	% Swelling of Disintegrant	Average Swelling
00.00% SLS	5.71 7.24	6.48
00.01% SLS	12.86 10.00	11.43
00.10% SLS	5.71 8.57	7.14
05.40% SLS	0.00 0.00	0.00
10.00% SLS	0.00 0.00	0.00

TABLE XII

t Test for SLS Reading

SLS %	t Computed	t Table Value	Comment
0.00% 0.01%	3.05	4.3	Not Significant
0.00% 0.10%	0.41	4.3	Not Significant
0.00% 05.40%	8.46	4.3	Significant

TABLE XIII

Disintegrant: Ac-Di-Sol

Solvent	% Swelling of Disintegrant	Average Swelling
0% Alcohol	5.71 7.24	6.48
1% Alcohol	11.43 11.42	11.43
4% Alcohol	7.14 14.28 8.57	10.00
6% Alcohol	11.43 14.29	12.86
15% Alcohol	14.29 14.29	14.29
40% Alcohol	4.28 7.14	5.71
95% Alcohol	0.00 0.00	0.00

TABLE XIV

t Test for Alcohol Reading

Alcohol %	t Computed	t Table Value	Comment
0%			
1%	6.47	4.3	Significant
0%			
4%	1.15	4.3	Not Significant
0%			
6%	3.93	4.3	Not Significant
0%			
15%	10.21	4.3	Significant
0%			
40%	0.47	4.3	Not Significant
0%			
95%	8.46	4.3	Significant

Swelling of Ac-Di-Sol in water, alcohol, and Sodium Lauryl Sulfate has been shown graphically (Fig. 9 to 11). Emcosoy and Ac-Di-Sol swelling will not be affected significantly by the fluctuation of PH in the stomach, as well as the intestine. The most interesting observation was their swelling in Sodium Lauryl Sulfate solutions. As it appears in the table, Ac-Di-Sol particles had maximum swelling at .01% conc. of the solution, while no swelling was observed in 5.40% solution, which is recommended as a dissolution medium for drug-like Griseofulvin. The solution of Sodium Lauryl Sulfate at 5.40% conc. reduced the swelling of Emcosoy significantly.

It was observed that very dilute alcohol solutions and Sodium Lauryl Sulfate solution reduced the surface tension of water, and they also acted as a wetting agent and enhanced the absorption of water by Ac-Di-Sol; but as their concentration increased, they ceased to act as wetting agents and there was relatively small proportions of water available for disintegrant to absorb. Therefore, swelling was reduced or, in some cases, swelling did not occur.

### 3. Water Penetration of Disintegrations:

Water penetration in Explotab, Emcosoy, CLDII, and Ac-Di-Sol was measured. The square of penetrated length of powder bed versus time for various disintegrant was tabulated from Table XV to XViii. The same data was plotted on page .

TABLE XV

Disintegrant: EmcosoyTime (sec.) Water Penetration in (cm)<sup>2</sup> at 22 ± 1c

---

1500	1.02
2400	1.86
3600	2.82
5160	4.15
6000	4.41

TABLE XVI

Disintegrant: Ac-Di-SolTime (sec.) Water Penetration in (cm)<sup>2</sup> at 22 ± 1c

---

1200	0.18
2400	0.28
3600	0.32
6000	0.40

TABLE XVII

Disintegrant: CLD IITime (sec.) Water Penetration in (cm)<sup>2</sup> at 22 ± 1c

---

1200	0.18
2400	0.28
3600	0.45
4800	0.71

TABLE XVIII

Disintegrant: Explotab

Time (sec.)	Water Penetration in (cm) <sup>2</sup> at 22 ± 1c
1500	3.57
2400	6.89
3600	9.27
4800	11.29
6000	14.29
7200	16.60



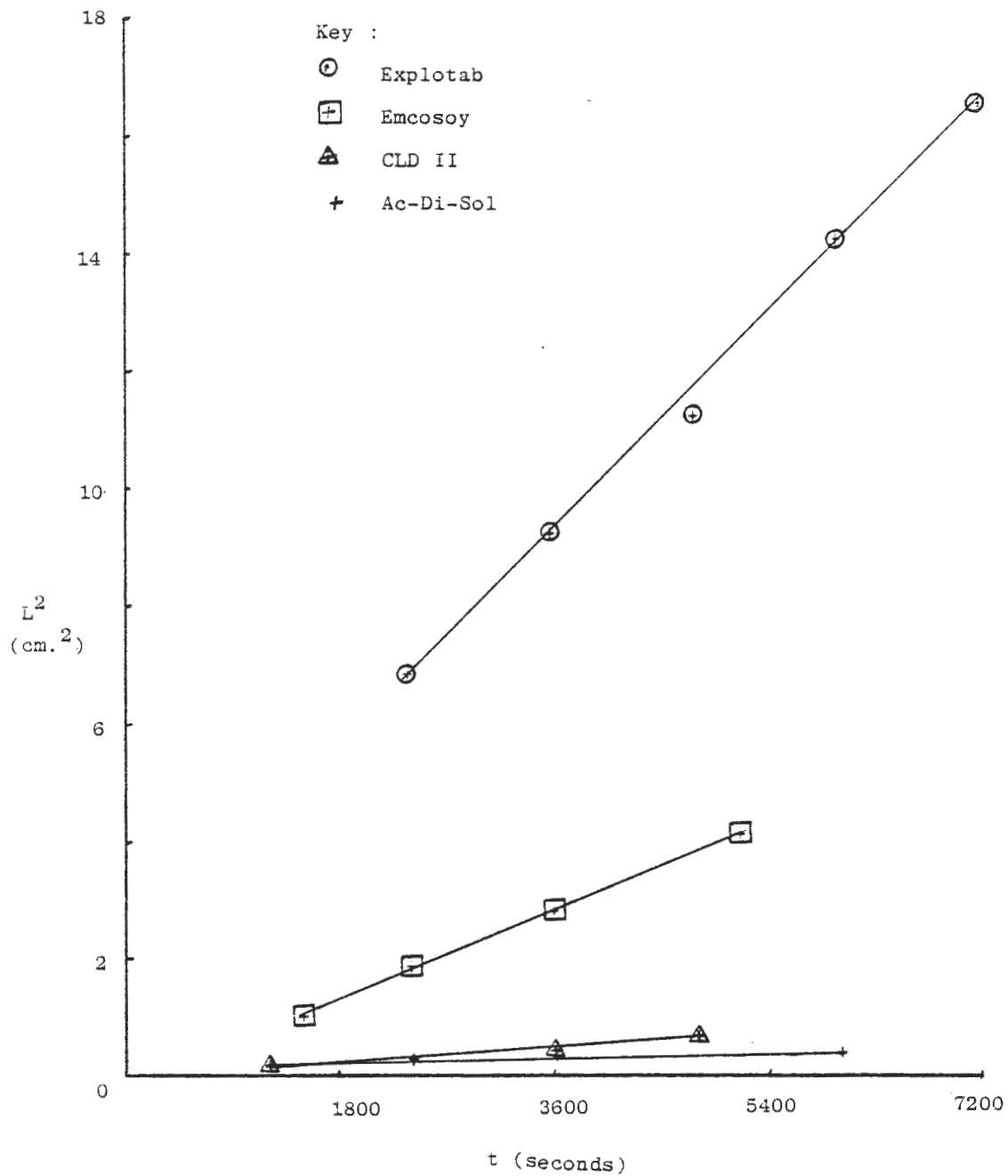
THE PENETRATION OF WATER INTO VARIOUS DISINTEGRANTS:

Fig. 12

is also very important, since in tablet disintegration it is expected that disintegrant swells considerably to rupture the tablet as quickly as possible, so rate and extent of swelling are equally important.

The penetrating rate of liquid into the disintegrant bed is expressed by Peek-McLean's equation (55):

$$\frac{dL}{dt} = \frac{r\gamma\cos\theta}{4NL} - \frac{r^2 dg}{8n}$$

Where L is the penetrating length at time t, r is the average radius of void space,  $\theta$  the contact angle between liquid and powder surface, g the acceleration constant of gravity, and  $\gamma$ , n, and d are the surface tension, viscosity, and specific gravity of liquids, respectively. The penetration rate also depends on the viscosity of gel formed by the interaction of water and disintegrant. If the gel formed is too viscous, it would impede the further penetration of water in a disintegrant bed. All these factors will be discussed in great detail separately.

In addition to water, water-alcohol mixtures of different strength, sodium lauryl sulfate solution of various concentrations and solutions of PH range 2 to 8 (Citrate - Phosphate - Borate/HCl buffer) were used to study the bulk swelling of Emcosoy and Ac-Di-Sol. Student t test was performed using Statpack computer program to find out whether the difference in swelling was significant or non-significant. All the readings and results are tabulated from Table II to XIV.

From the tables and graph, it appeared that water penetration rate was maximum in Explotab powder bed, followed by Emcosoy, CLD II, and Ac-Di-Sol under identical experiment conditions. The reason could be that average void space and contact angle between Explotab particle and water was high, compared to other disintegrants tested.

#### 4. Energy of Wetting:

The values of K for different water temperatures for Emcosoy was tabulated in Table XIX.  $\log K/T$  vs.  $\frac{1}{T}$  was plotted on page 73. The value of  $\beta$  (the energy of wetting) was calculated for Emcosoy.

The value of energy of wetting obtained by this method according to some critics is an apparent value, since they believe that the approach is too simplified to give any real values.

#### 5. Wetting and Drying of Disintegrant:

Emcosoy batches were prepared under different conditions. Bulk swelling was determined for these batches. Results are tabulated in Table XX.

As it can be seen from the Emcosoy table that was dried at room temperature and at 40°C had a bulk swelling less than the untreated (original) Emcosoy. One reason could be that these samples were dried at low temperatures, and therefore significant amounts of surface moisture was left in these samples; that could have resulted in low water uptake and less swelling.

Energy of wetting of Emcosoy particles.

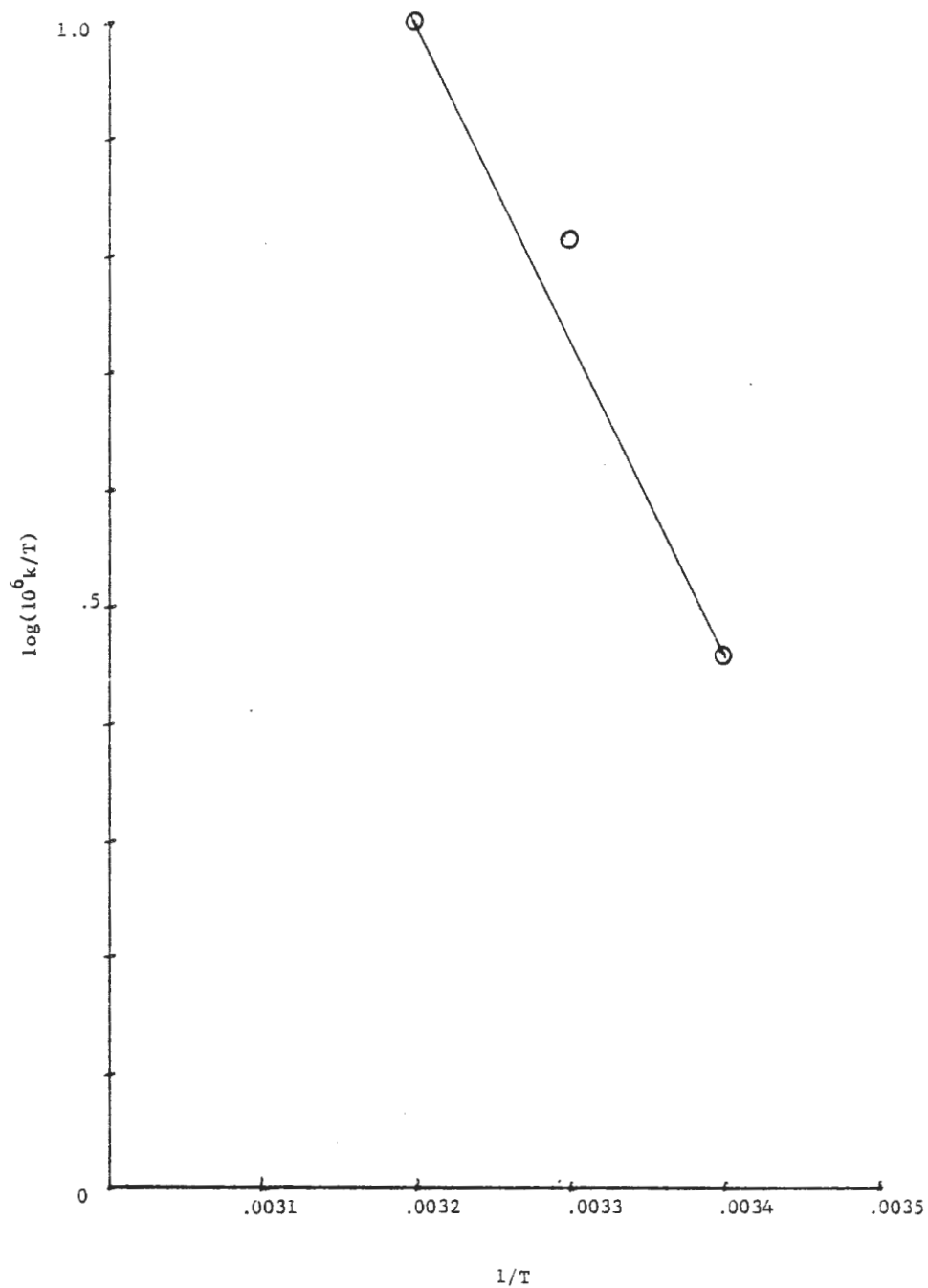


Fig. 13

TABLE XIX

Energy of wetting of Emcosoy particles :

<u>k</u>	<u>T</u>
$0.848 \times 10^{-3}$	295.0
$1.955 \times 10^{-3}$	302.2
$3.083 \times 10^{-3}$	310.0

<u><math>\log (10^6 k/T)</math></u>	<u>1/T</u>
0.4586	$3.4 \times 10^{-3}$
0.8109	$3.3 \times 10^{-3}$
0.9976	$3.2 \times 10^{-3}$

Calculation :

From graph

$$\text{slope} = - \beta/R$$

$$-2.695 \times 10^3 = - \beta/1.987 \text{ Cal.}$$

$$\beta = 5.35 \text{ K.cal.}$$

Emcosoy dried at 60<sup>c</sup> had bulk swelling more than four times that of an original Emcosoy. The result could be explained by the comparison of baking of a cake with baking (drying) of a disintegrant particle. When the cake is baked, the surface area of the cake increases enormously, and the same thing can happen in the disintegrant particles. The increase in surface area could lead to increased water uptake and increased swelling.

Emcosoy dried at 60<sup>c</sup> had an improved flow and high density compared to the original Emcosoy. The color change was from yellowish white to brown. No swelling was observed in the first two minutes in the original Emcosoy, but there was 30% to 35% swelling in the first two minutes in Emcosoy dried at 60<sup>c</sup>. Wicking effect was very much improved in the Emcosoy dried at 60<sup>c</sup> -- the time that was taken by the water to travel through the entire powder bed was 60 minutes, compared to 90 minutes for the original Emcosoy.

There was little offending odor from Emcosoy dried at 40<sup>c</sup> and 60<sup>c</sup>, and visible mold growth in Emcosoy which was dried at room temperature. Published literature on the subject indicates that molds require a water activity number of 0.75, and bacteria requires 0.88. Below these values, microbial growth cannot be supported. It has been estimated that Emcosoy with a 10% moisture content = water activity number 0.5; and with 20% moisture content, there exists a water activity of 0.88. Since original Emcosoy usually contains less than 10% moisture, it theoretically cannot promote microbial growth. However, in a

wet granulation process, this argument might not be true all the time. Prolonged storage or improper storage may lead to higher moisture content and consequently makes this disintegrant vulnerable to microbial growth.

As discussed above, that Emcosoy dried at 60<sup>C</sup> had a better physical property than the original Emcosoy; they both were compared in actual tablet formulation. Original Emcosoy was identified as "A," and the processed Emcosoy was identified as "B." In order to get unbiased evaluations of these disintegrants, the tablets were prepared and tested by my colleague. The results and other details are shown in a tabular form (Table XXI).

As it can be seen from the table and contrary to expectation, processed disintegrant tablets had higher disintegrant time; in other words, the tablets made using it took a long time to disintegrate, compared to tablets made using original Emcosoy. The reason could be poor deformation of processed disintegrant. The three major mechanisms of disintegration are: (1) swelling, (2) porosity and capillary action (wicking), and (3) deformation. As mentioned above, processed disintegrant is better than original Emcosoy in the first two categories. There is no data to compare the third one. When the tablet is compressed, the compression may cause more permanent deformation in original Emcosoy than the processed Emcosoy. When the tablet is put into a disintegrating medium, the energy-rich Emcosoy grains swell rapidly, unlike processed Emcosoy grains, which require more heat in order to swell. The overall effect leads to an early disintegration of the

TABLE XX

Details	% Swelling	Average Swelling
Emcosoy (original)	44.3	44.6
(Passed through 100 mesh)	42.9	
Emcosoy (dried at room temp.)	31.4	30.0
21 <sup>c</sup>	28.6	
Emcosoy (dried at 40 <sup>c</sup> )	34.3	32.15
	30.0	
Emcosoy (dried at 60 <sup>c</sup> )	192.7	197.1
	201.4	



TABLE XXI

	In Each Tablet	Percent	Working Formula
Hydrochlorothiazide	40.0 mg	20.0	60.0 Gm
Disintegrant <u>A or B</u> <i>A O F B</i>	2.0 mg	1.0	3.0 Gm
Magnesium Stearate	1.0 mg	0.5	1.5 Gm
Emcompress	157.0 mg	78.5	235.5 Gm

Dis.	Weight mg	R.S.D.	Thickness mm	R.S.D.	Hardness Kg	R.S.D.	Friab. %	Dis. Time
A	206.1	0.779	4.20	0.283	4.94	9.162	2.76*	6'40" M 4'55"-11'10" R  5'39" M 3'45"-9'30" R
B	208.1	0.349	4.21	0.225	5.08	12.322	2.63**	8'18" M 4'40"-11'30" R  8'48" M 7'30"-12'35" R

\* Includes caps, without caps 12.68%

\*\*Includes caps, without caps 10.72%

M = Mean

R = Range

tablet made from original Emcosoy.

#### 6. Aqueous Dispersion:

Shangraw, et al, have theorized that substances that swell to very large gelatinous volume might be excellent disintegrants in a tablet that breaks apart rapidly; however, their inclusion in a tablet that disintegrates more slowly could lead to surface gelling that would only further impede liquid penetration of the tablet matrix ( 6 ). They compared two percent aqueous dispersions of various starches, cellulose products, and of cross-linked PVP in water-diluted hydrochloric acid, (1:100) were shaken periodically, and were allowed to stand overnight. They found that all the disintegrants had different swelling; in some of the disintegrants, the swelling was enhanced or suppressed in acidic medium.

It is expected of ideal disintegrants that its swelling should remain unchanged or preferably increased in acidic medium, since most of the tablet disintegrates in acidic environments of the stomach. Emcosoy had 187.5 percent swelling in water, and 287.5 percent swelling in acidic medium. The results clearly indicated that vivo PH changes would have no adverse effect on disintegrating function of Emcosoy (Table XXIV).

#### 7. Comparative Study of the Viscosity of Different Strength Solutions of Various Disintegrants:

Viscosity data of various disintegrant solutions are given from Tables **XXII** and **XXIII**. The disintegrants are divided into two groups: one under the heading, "Cellulose Group," and the other under the heading, "Starch Group."

TABLE XXII

Viscosity Study of Gel Formed by Various Disintegrants:

Disintegrants	rpm	2%	4%	6%	8%	10%
<u>Cellulose Group:</u>						
Soy Cellulose	20	00.00	00.10	03.30	17.00	200.00
(Emcosoy)	50	00.08	00.14	02.60	16.00	120.00
	100	00.20	00.50	01.69	14.00	88.00
Cotton Cellulose	20	00.13	00.13	00.30	02.73	25.60
(Ac-Di-Sol)	50	00.22	00.35	00.40	02.20	18.80
	100	00.22	00.40	00.75	02.11	13.00
Wood Cellulose	20	05.90	224.00	1260.00	-----	-----
(CLD II)	50	04.68	146.90	664.00	-----	-----
	100	03.76	97.00	-----	-----	-----
<u>Starch Group:</u>						
Corn Starch	20	00.00	00.00	00.00	00.00	00.00
	50	00.04	00.10	00.08	00.08	00.12
	100	00.06	00.12	00.07	00.23	00.32
Sta Rx 1500	20	00.00	00.05	00.00	00.00	00.12
	50	00.00	00.16	00.12	00.14	00.14
	100	00.00	00.18	00.36	00.57	00.21
Sodium Starch	20	00.00	00.13	01.25	148.50	530.00
Glycolate	50	00.12	00.25	01.74	63.00	252.80
(Explotab)	100	00.12	00.34	02.55	39.20	136.80
Polyplasdone XL	20	00.00	00.00	00.00	00.05	00.45
(Grade c)	50	00.00	00.00	00.06	00.10	00.42
	100	00.26	00.35	00.45	00.65	00.54

Viscosity Study of Gel Formed by Various Disintegrants

Disintegrants	rpm	12%	14%	16%
<u>Cellulose Group:</u>				
Soy Cellulose (Emcosoy)	20	464.00	----	----
	50	243.20	----	----
	100	156.00	----	----
Cotton Cellulose (Ac-Di-Sol)	20	335.00	1310.00	----
	50	196.00	594.40	----
	100	102.00	370.00	----
Wood Cellulose (CLD II)	20	----	----	----
	50	----	----	----
	100	----	----	----
<u>Starch Group:</u>				
Corn Starch	20	00.00	00.10	----
	50	00.08	00.10	----
	100	00.44	00.70	----
Sta Rx 1500	20	00.10	00.40	----
	50	00.30	00.50	----
	100	00.36	00.64	----
Sodium Starch Glycolate (Explotab)	20	1980.00	----	----
	50	----	----	----
	100	----	----	----
Polyplasdne XL (Grade c)	20	02.20	07.05	----
	50	01.66	04.36	----
	100	01.30	02.70	----

Cellulose Group:

The viscosity of cellulose disintegrant solutions can be arranged in decending order:

1. Viscosity of CLD II solutions,
2. Viscosity of Emcosoy solutions, and
3. Viscosity of Ac-Di-Sol solutions.

CLD II solutions exhibited the maximum viscosity, compared to any disintegrant of either group. The gel formed by Ac-Di-Sol was less viscous compared to that of Emcosoy. If we have to justify the difference in the vitro bioavailability of two vitro formulations containing Emcosoy and Ac-Di-Sol, respectively, as disintegrants solely on the basis of viscosity, we can say that Ac-Di-Sol formulations will have a little superiority over Emcosoy formulations since the gel formed by Ac-Di-Sol will be less viscous than the gel formed by Emcosoy. As expected, cellulose derivatives form viscous gel compared to their starch counterparts.

Starch Group:

If the viscosity of starch disintegrant solutions has to be arranged in decending order, then:

1. Viscosity of Explotab solutions,
2. Viscosity of Sta-Rx 1500, and
3. Viscosity of Corn Starch.

Explotab gel has the maximum viscosity among starch disintegrants. If we have to predict the performance of Explotab and corn starch as disintegrant solely on the basis of the viscosities

of their gel, we might expect corn starch as a better disintegrant than Explotab, since the gel formed by the former is less viscous.

In practice, Explotab is much more superior to corn starch; the reason is that in addition to viscosity, there are other important factors, such as rate and extent of swelling, wicking effect, and deformation play crucial roles in disintegration mechanism.

Explotab, Emcosoy, Ac-Di-Sol, and corn starch were compared in actual tablet formulations. The viscosity of the gel of Explotab is greater than that of Emcosoy. The viscosity of Emcosoy is greater than that of Ac-Di-Sol. Vitamin formulations studied clearly indicated that Explotab is inferior to Ac-Di-Sol and Emcosoy. Viscosity study provides the answer. Explotab swells much more compared to Ac-Di-Sol and Emcosoy, and we therefore expect it to be better disintegrant compared to Ac-Di-Sol and Emcosoy. However, that is not true, because Explotab forms very viscous gel, and it will be shown later that it has poor deformation.

The disintegrant solutions were subjected to different shear rate, and the viscosity were determined. The rheological profiles of the various disintegrants were shown in Fig. 15 to 18. In theory, there were all three kinds of profiles, as shown in Figure 14. In Newtonian liquids the viscosity remains unchanged with the increase in shear rate.

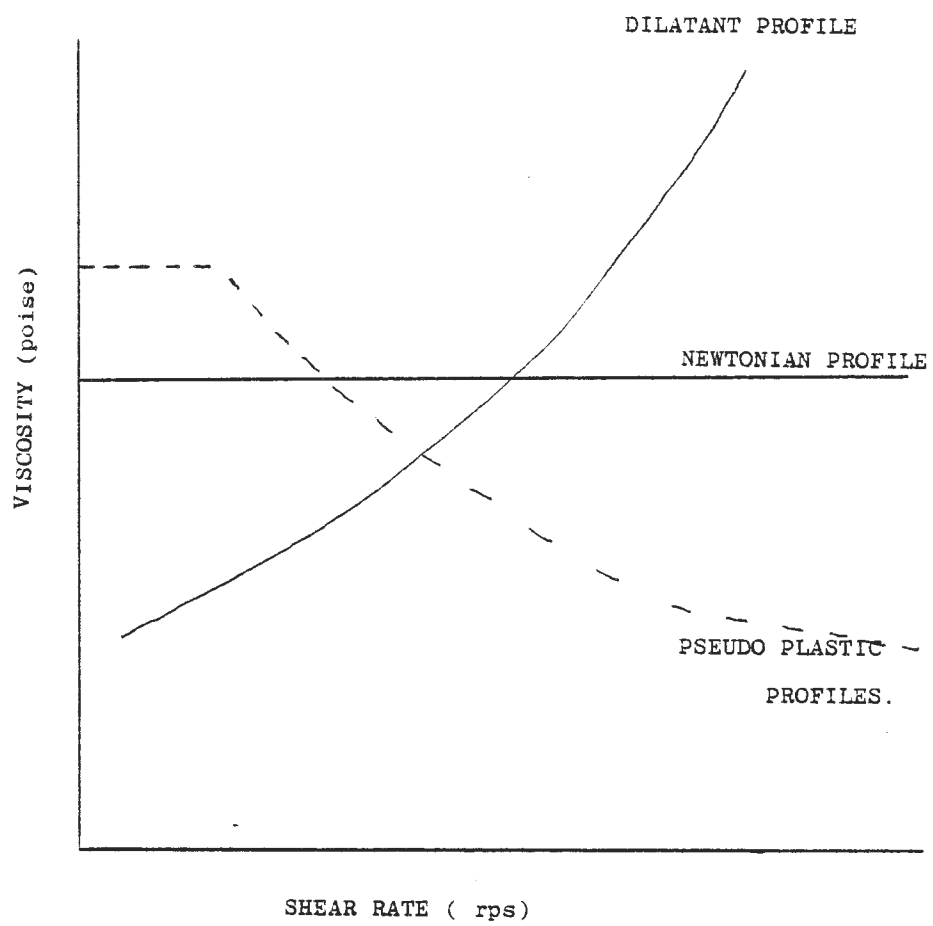


Fig. 14

RHEOLOGICAL PROFILE OF 1.Explotab - all at 10% conc.  
2.Emcosoy.  
3.Ac-Di-Sol

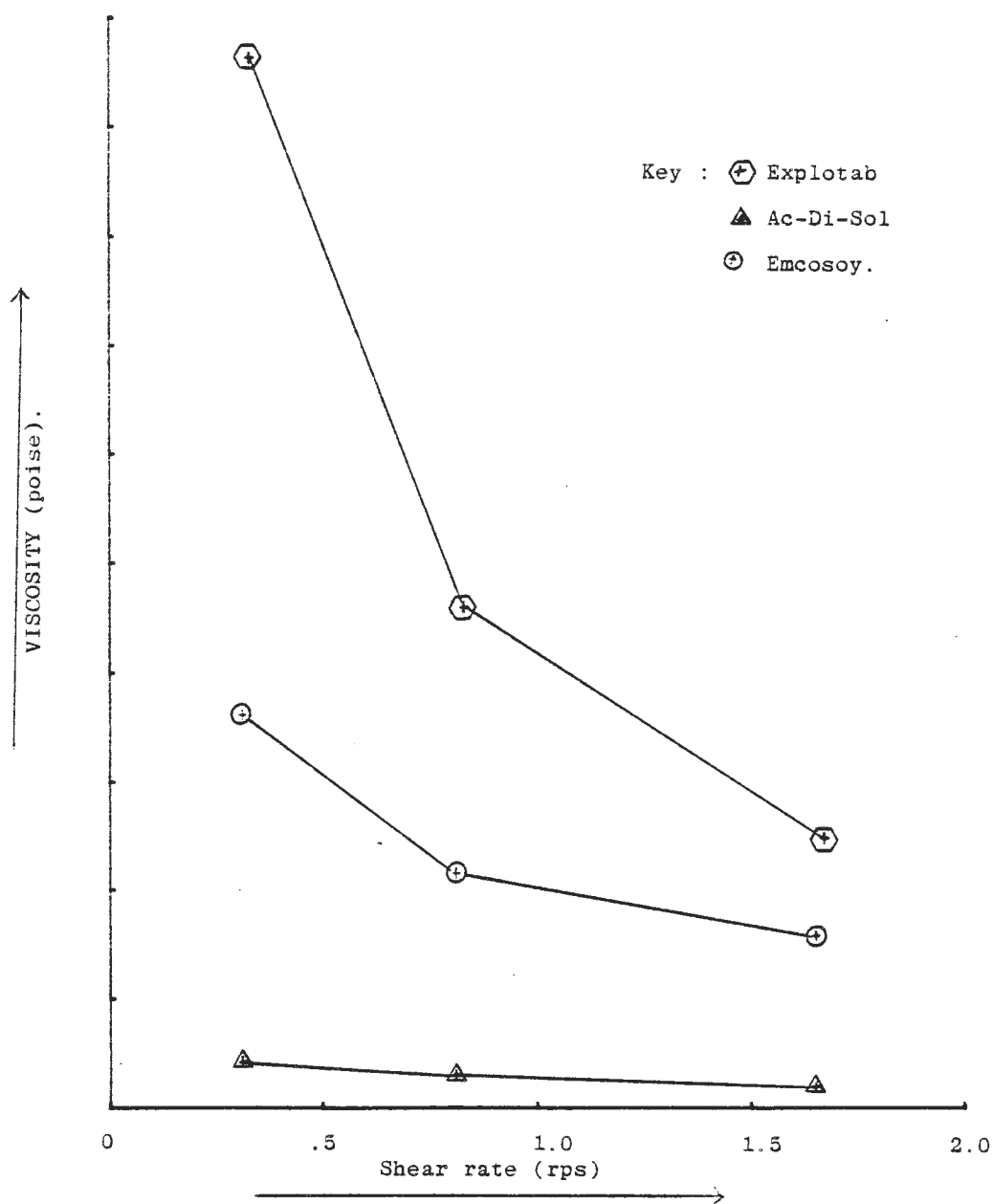


Fig. 15



RHEOLOGICAL PROFILE OF 1. CORN STARCH GEL -DISINTEGRANT CONC.14%  
2. Sta Rx 1500

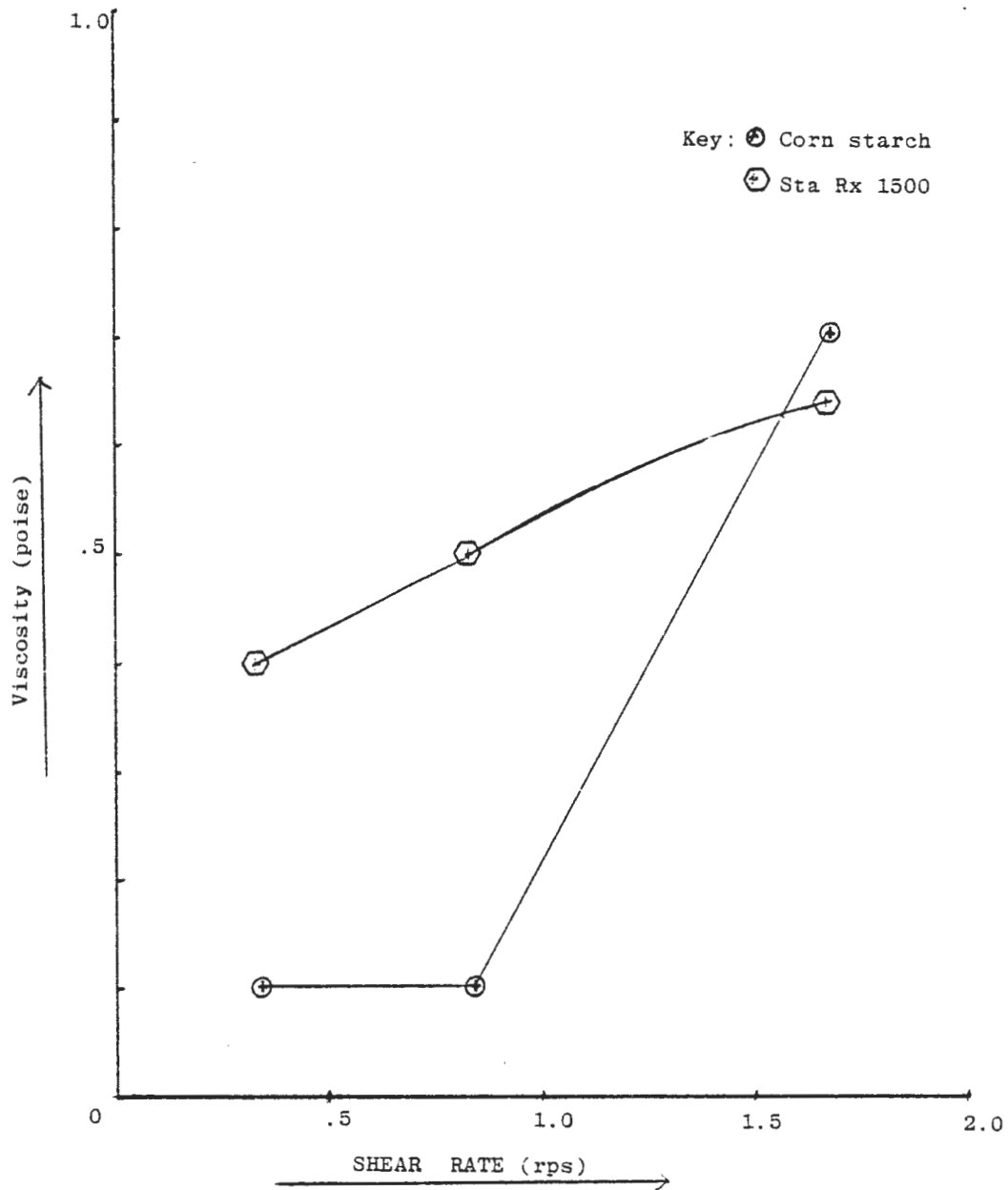


Fig. 16

RHEOLOGICAL PROFILE OF POLIPLASDONE XL AT 14% CONC. OF THE  
DISINTEGRANT.

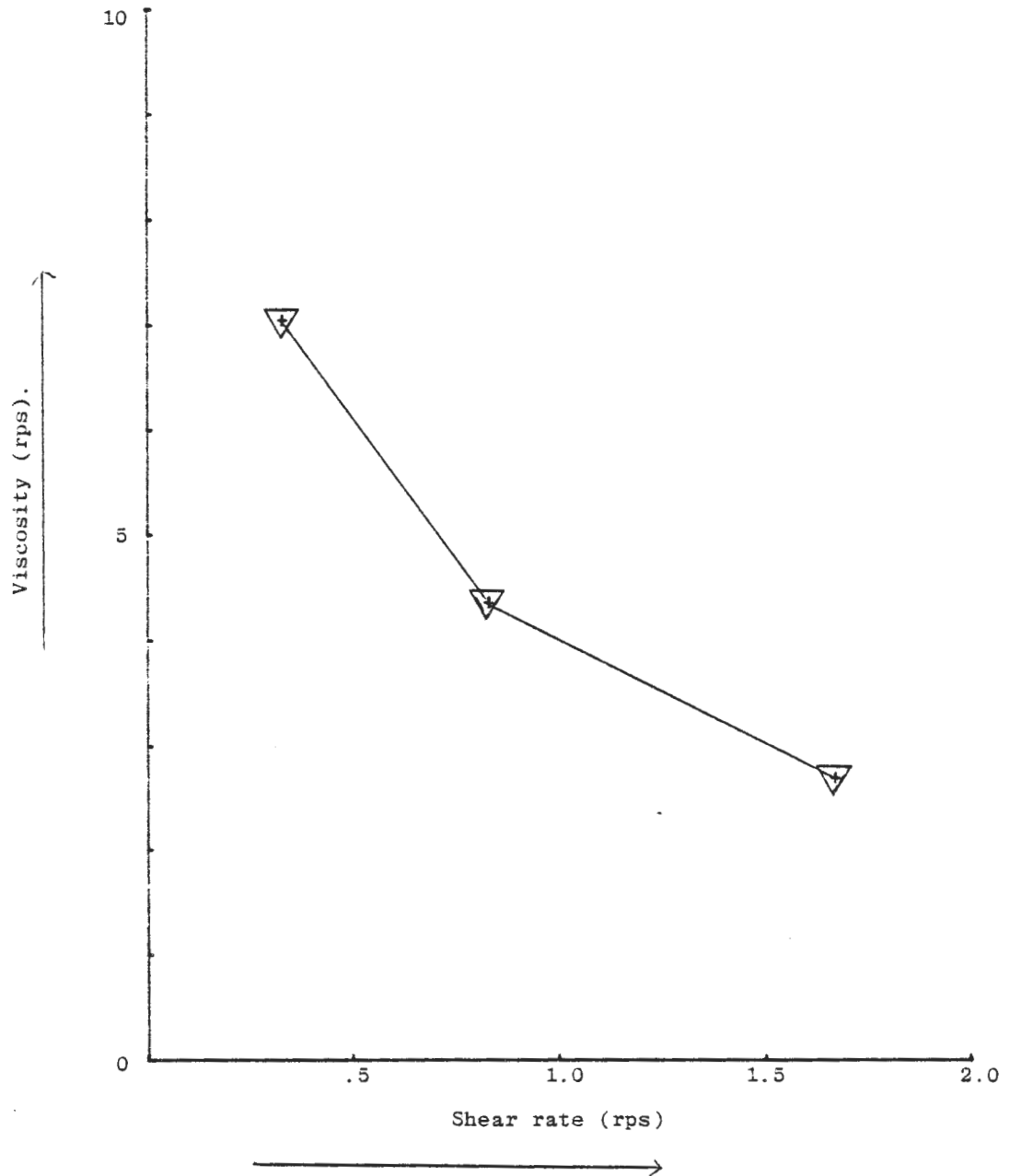


Fig. 17

RHEOLOGICAL PROFILE OF CLD II GEL AT 4% CONC. OF THE  
DISINTEGRANT.

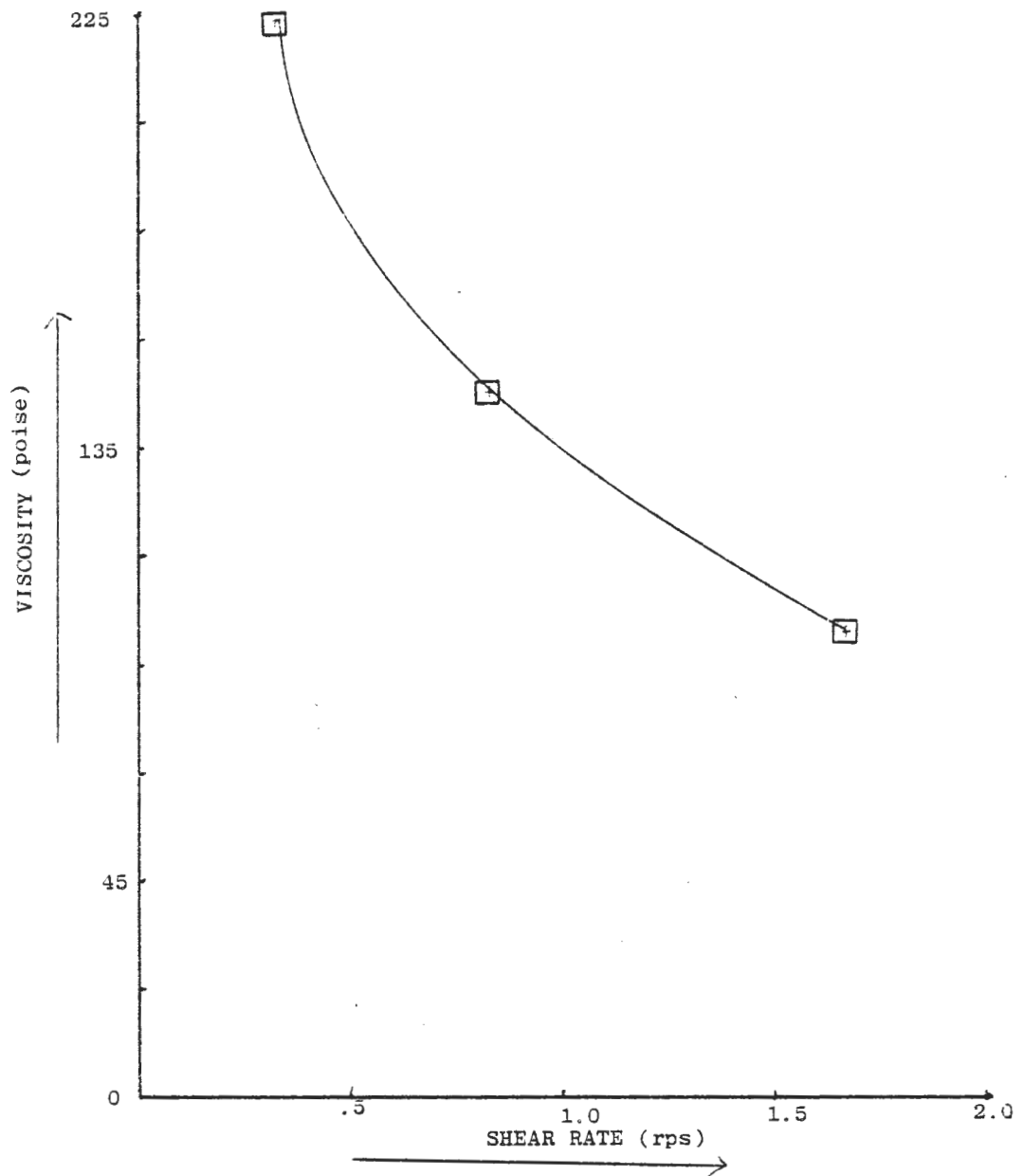


Fig. 18

In dilatant profiles, the viscosity increases with increases in shear rate. In pseudoplastic profiles, the viscosity decreases with increase in shear rate. Emcosoy, Ac-Di-Sol, CLD II, Explotab, and poliplasdone XL gave pseudoplastic profiles. Corn starch and Sta Rx 1500 solutions had dilatant profiles. Dilatancy could be one of the reasons for the poor vitro bioavailability of the formulations containing corn starch.

8. Maximum Hydration Capacity of Various Disintegrants:

Ac-Di-Sol -	16% W/V
Emcosoy -	14% W/V
CLD II -	8% W/V

High hydration capacity is desirable property of disintegrant.

High hydration capacity indicates that the disintegrant requires less amounts of water to wet itself thoroughly. It can be theorized that a disintegrant with high hydration capacity would relatively have smaller water uptake in order to achieve disintegration of the tablet, compared to a disintegrant with low hydration capacity. It can be said from the data that as the viscosity of the gel of disintegrant increases, the maximum hydration capacity decreases.

9. Sieve Analysis:

Nogami, et al, found that there should be the critical amount of starch necessary for the disintegration depending upon the particle size or specific area of ingredients. The smaller the particle size of aspirin, the more amount of starch was required for the tablet disinte-

TABLE XXIV

Emcosoy (2%) Dispersion in Water and Dilute Acid (Hcl 1 in 100):Apparatus: 100 ml. measuring cylinder.Dilute Acid (1 in 100) Hcl. pH .70Time Duration: Overnight.

Solvent	Initial Volume	End Volume	% Swelling
Water	8.0 ml.	23.0 ml.	187.5
Dilute Acid	8.0 ml.	31.0 ml.	287.5

TABLE XXV

Emcosoy - Sieve Analysis:

Sieve Number U.S. Standard (Passed/Retained)	Arithmetic Mean Size of Opening	Wt. Retained on Smaller Sieve(%)	Wt. Size
100/170	119	8.5	1011.5
170/200	81	6.8	550.8
200/230	68	11.4	775.2
230/270	58	22.8	1322.4
270/325	49	32.7	1602.3
325/400	41	8.4	344.4
400/Bottom	--	4.5	--

$$d_{ave} = \frac{5606.6}{90.6} = 62 \text{ } \mu$$

gration. The tablet of the smallest aspirin (11.9 in mean particle size) did not disintegrate by the addition of 20% of starch. Thus, the particle size of the ingredients play an important part in tablet disintegration (71).

Rudnic, et al, studied the different particle size grades of cross-linked polyvinylpyrrolidone (Polyplasdone, G.A.F. Corporation) in direct compression tablet formulations showed that increases in mean particle size enhanced powder flow, disintegration and dissolution, although hardness and friability were slightly better for tablets made from the finer grades (72). Therefore, particle size of the ingredients, as well as disintegrants, are equally important.

There is nothing like optimum particle size of disintegrant or matrix. Particle size varies for various disintegrants and tablet matrices. However, the particle size should be such that disintegrant can be mixed and distributed uniformly with the rest of the ingredients. The particle size of Emcosoy was determined using sieving. Table XXV shows the size-weight distribution of Emcosoy as measured by U.S. Standard sieves.

#### 10. Plastic Deformation and Elasticity of Various Disintegrants:

Modulus of elasticity at various attempted deformations and compressive force needed to achieve that deformation were tabulated in Table XXVI. The percentage of plastic deformation and the percentage re-

TABLE XXVI

Disintegrant	Deformation Attempted	Modules of Elasticity lb/in <sup>2</sup>	Compressive Force (lbs)
Emcosoy	10%	328.35	100
	20%	391.47	175
	40%	1063.95	1240
CLD II	10%	429.56	150
	20%	570.45	340
	40%	1677.85	2000
	50%	2013.42	3000
Ac-Di-Sol	20%	6291.95	3750
Explotab	10%	9357.93	2850
	20%	12931.26	8050
Sta-Rx 1500	20%	5706.98	3250
Polyplasdnone-XL	10%	377.41	120
	20%	503.35	300
	40%	1142.79	1400

TABLE XXVII

Disintegrant	Deformation Attempted	% Plastic Deformation	% Recovery
Emcosoy	10%	4.89	5.11
	20%	13.33	6.67
CLD II	10%	7.56	2.44
	20%	13.33	6.67
Ac-Di-Sol	10%	9.30	0.70
	20%	12.44	7.56
Explotab	10%	3.89	6.11
Sta-Rx 1500	10%	8.0	2.0
	20%	14.78	5.22
Polyplasdane-XL	10%	6.22	3.78
	20%	12.22	7.78



covery of various disintegrants were tabulated in Table XXVII.

From the elasticity data, the disintegrants were arranged in the descending order of their elasticity on page 96. The order remained unchanged for CLD II, polyplasdone-XL, and Emcosoy at 10%, 20%, and 40% attempted deformation.

Force required to achieve various percentage of deformations for various disintegrants was tabulated on page 98. The force needed to attain 20% deformation in CLD II was little more than double of the force needed to attain 10% deformation in the same disintegrant. Similar observation was made in polyplasdone-XL, and to some extent in Emcosoy. However, when the deformation was doubled from 20% to 40%, the force needed to achieve that deformation was almost five to six times than the force needed to achieve 20% of deformation. From these results, it was concluded that it was easy to deform particles initially; but as the percentages of deformation increased, the task became very difficult and greater force was needed to achieve those deformations. It was also observed that unusually high force was needed to achieve even 10% deformation in Explotab. It might be due to high degree of cross-linking among starch glycolate molecules. Cross-linked polymers of starch are more elastic, compared to their cellulose counterparts. At 20%, attempted deformation starch polymers required much more compressive force compared to cellulose polymers.

The percentages of plastic deformation and the percentages of recovery at various attempted deformations for different disintegrants were tabulated on page 93. As it could be seen from the table, at 10% attempted deformation Ac-Di-Sol had more plastic deformation than Sta-Rx 1500 and CLD II, but at 20% attempted deformation order was reversed. It was clear from the data that the initial deformation might be more in Ac-Di-Sol compared to Sta-Rx 1500 and CLD II, but the additional deformation -- the deformation after some initial range -- was less compared to Sta-Rx 1500 and CLD II, which indicated that Ac-Di-Sol particles deformed initially easily but later they resisted more deformations. The recovery was least at 10% in Ac-Di-Sol, but at 20% the recovery was more than those of Emcosoy, CLD II, and Sta-Rx 1500.

The disintegrants were arranged in descending order according to the plastic deformation (at 10% attempted deformation) on page 97 . The deformed particles are energy rich, and that energy is released when the grains are exposed to water. The energy-rich particles swell more rapidly in water, unlike undeformed grains, which require more heat in order to swell. These data provide an answer that why Ac-Di-Sol containing formulations had better disintegration and dissolution compared to formulations containing Emcosoy and Explotab.

ELASTICITY (decending order):

At 10% attempted deformation:

1. Explotab
2. CLD II
3. Polyplasdnone-XL
4. Emcosoy

At 20% attempted deformation:

1. Explotab
2. Ac-Di-Sol
3. Sta-Rx 1500
4. CLD II
5. Polyplasdnone-XL
6. Emcosoy

At 40% attempted deformation:

1. CLD II
2. Polyplasdnone-XL
3. Emcosoy

% PLASTIC DEFORMATION (decending order):at 10%

1. Ac-Di-Sol
2. Sta-Rx 1500
3. CLD II
4. Polyplasdnone-XL
5. Emcosoy
6. Explotab

at 20%

1. Sta-Rx 1500
2. CLD II, Emcosoy
3. Ac-Di-Sol
4. Polyplasdnone-XL

RECOVERY (decending order):at 10%

1. Explotab
2. Emcosoy
3. Polyplasdnone-XL
4. CLD II
5. Sta-Rx 1500
6. Ac-Di-Sol

at 20%

1. Polyplasdnone-XL
2. Ac-Di-Sol
3. Emcosoy, CLD II.
4. Sta-Rx 1500

FORCE REQUIRED FOR 10% DEFORMATION OF VARIOUS DISINTEGRANTS:

<u>DISINTEGRANT</u>	<u>COMPRESSIVE FORCE (lbs)</u>
Explotab	2850
CLD II	150
Polyplasdone-XL	120
Emcosoy	100

---

FORCE REQUIRED FOR 20% DEFORMATION OF VARIOUS DISINTEGRANTS:

<u>DISINTEGRANT</u>	<u>COMPRESSIVE FORCE (lbs)</u>
Explotab	8050
Ac-Di-Sol	3750
Sta-Rx 1500	3250
CLD II	340
Polyplasdone-XL	300
Emcosoy	175

---

FORCE REQUIRED FOR 40% DEFORMATION OF VARIOUS DISINTEGRANTS:

<u>DISINTEGRANT</u>	<u>COMPRESSIVE FORCE (lbs)</u>
CLD II	2000
polyplasdone-XL	1400
Emcosoy	1240

## 11. TABLET PRODUCTION:

Tables XXVIII through XXXII show physical properties of the tablet formulae I, II, and III.

High weight variation, thickness and high hardness variation were observed in multivitamin formulation (Formula I). Capping was observed in a formulation having Emcompress as a matrix. Emdex matrix formulation gave friability less than .90%. Disintegration time was less than 20 minutes in most of the formulation except formulation containing corn starch at 3% level in Emcompress matrix. It is obvious from the disintegration time in Tables XXVIII and XXIX that the effect of various disintegrant is much pronounced in water-insoluble matrix, such as Emcompress, compared to water-soluble matrix, such as Emdex. Dissograph was used for the dissolution study. Total optical absorbance was measured. From each formulation three dissolution profiles were studied. The results were quite reproducible (Tables XXXIII and XXXIV). Emdex matrix gave  $t_{50}$ ,  $t_{75}$ , and  $t_{90}$  within a range of two to nine minutes for various disintegrants at various concentration; while in the case of Emcompress matrix, those values differed considerably. Corn starch at 3% level gave  $t_{75}$  and  $t_{90}$  more than 30 minutes. Corn starch and Emcosoy was compared at 3% level in Emcompress as a matrix. Obviously, Emcosoy was a better disintegrant than corn starch (Fig.27).

A narrow weight variation, hardness variation, and thickness variation were observed in pyridoxine HCl formulation (Formula II). Hardness was optimum. Tablets with Emdex as a matrix were thicker than Emcompress matrix tablet. Capping was a problem in some of the formulation containing Emcompress as a matrix. The friability was less than .50% in Emdex matrix formulations. As observed in multi-vitamin formulation, the effect of various disintegrants at different concentration was more pronounced in Emcompress matrix compared to Emdex matrix formulation. At .5% disintegrant level the tablet containing Explotabs disintegrant and Emcompress as matrix failed to disintegrate within one hour, although at 1% and 2% the same disintegrant was found satisfactory. At 2% disintegrant level, all the disintegrants except corn starch were found satisfactory. Tablet containing corn starch as disintegrant at 2% level failed to disintegrate within one hour. All formulations containing Emdex as a matrix gave disintegration time less than at eight minutes at various disintegrant levels. Two dissolution profiles were studied from each formulation. At 0.5% disintegration level in Emcompress matrix Emcosoy, Ac-Di-Sol and Explotab had  $t_{90}$  more than 30 minutes; at 1% and 2% disintegrant level  $t_{50}$ ,  $t_{75}$ , and  $t_{90}$  was found satisfactory for these three disintegrants. However, at 2% level corn starch gave  $t_{90}$  more than 30 minutes compared to  $t_{90}$ 's of Emcosoy 1.5 minutes, Ac-Di-Sol three minutes, and Explotab 9.5 minutes (Fig.49). In Emdex

matrix all disintegrants gave satisfactory  $t_{50}$ ,  $t_{75}$ , and  $t_{90}$ . However, it was obvious from Tables XXXV and XXXVI that Emcosoy was superior to corn starch in both matrixes, and it competed successfully with Explotab and Ac-Di-Sol.

High weight, thickness and hardness variation were found in Ascorbic acid formulations (Formula III, Table XXXII). Ascorbic acid formulations contained only Emcompress matrix. Hardness was little less than optimum when applied pressure. Hardness increased first, and then decreased. Capping was found at low concentration level of disintegrant. Ascorbic acid is highly water soluble compared to other active ingredients used, so the disintegration time was much less except when corn starch was used as disintegrant. All formulations at 2% disintegrant level contained granulated Ascorbic acid (90%) (Roche); the rest of the formulation contained Ascorbic acid U.S.P. Granulated Ascorbic acid was less soluble compared to Ascorbic acid U.S.P.; hence, little high disintegration time was observed at 2% compared to 1% and .5% disintegrant level. Two dissolution profiles were studied from each formulation. The results were generally reproducible (Table XXXVII)  $t_{50}$ ,  $t_{75}$ , and  $t_{90}$  were satisfactory except for corn starch formulations. Corn starch at 2% and 3% gave  $t_{75}$  and  $t_{90}$  more than 30 minutes. It was very interesting to compare corn starch and Emcosoy at 3% level (Fig. 80), since both were all natural disintegrants:  $t_{90}$  for Emcosoy was two minutes, while for corn starch it was more than 30 minutes.



TABLE XXVIII  
MULTIVITAMIN FORMULATIONS WITH EMCOMPRESS AS A MATRIX

	WEIGHT		THICKNESS		HARDNESS		FRIABILITY Percent	DISINTEGRATION TIME	
	mg		mm		Kg			min-sec Mean	Range
<u>0.5% Disintegrant</u>									
Emcosoy	366.0	(1.21)	3.78	(0.98)	3.75	(57.30)	CAP*	10.10	5.40 to 19.15
Ac-Di-Sol	376.0	(1.54)	3.84	(1.31)	3.52	(48.84)	CAP	4.13	3.10 to 4.35
Explotab	354.7	(1.62)	3.84	(1.19)	3.83	(17.83)	3.14	12.11	10.03 to 16.50
<u>1.0% Disintegrant</u>									
Emcosoy	358.0	(1.23)	3.76	(0.66)	4.30	(43.26)	CAP	3.19	2.0 to 4.10
Ac-Di-Sol	373.6	(0.81)	3.83	(0.92)	3.66	(39.92)	CAP	1.56	1.10 to 2.25
Explotab	364.3	(0.55)	3.74	(0.73)	7.08	(32.51)	CAP	4.29	3.20 to 5.10
<u>3.0% Disintegrant</u>									
Emcosoy	356.1	(1.29)	3.77	(0.71)	4.73	(38.41)	CAP	1.15	0.50 to 1.50
Corn Starch	379.5	(1.13)	3.88	(1.03)	4.51	(26.62)	CAP	37.31	34.00 to 40.10

\*Capping

NOTE: In this and all subsequent tables, values in parentheses are Relative Standard Deviation.

TABLE XXIX

## MULTIVITAMIN FORMULATIONS WITH EMDEX AS A MATRIX

	WEIGHT mg	THICKNESS mm	HARDNESS Kg	FRIABILITY Percent	DISINTEGRATION TIME	
					Mean min-sec	Range
<u>0.5% Disintegrant</u>						
Emcosoy	346.2 (0.73)	4.42 (0.70)	11.53 (7.78)	0.51	3.50	3.30 to 4.05
Ac-Di-Sol	357.5 (1.29)	4.46 (0.89)	11.37 (9.58)	0.62	3.13	2.50 to 3.20
Explotab	355.8 (1.60)	4.47 (1.08)	10.37 (7.51)	0.55	3.19	3.15 to 3.20
<u>1.0% Disintegrant</u>						
Emcosoy	353.1 (1.33)	4.36 (0.98)	9.60 (16.60)	0.73	3.25	3.20 to 3.40
Ac-Di-Sol	362.1 (1.18)	4.40 (0.94)	11.26 (13.17)	0.80	4.08	3.50 to 4.20
Explotab	372.5 (1.68)	4.58 (1.39)	10.65 (10.64)	0.73	4.06	3.40 to 4.20
<u>3.0% Disintegrant</u>						
Emcosoy	342.7 (1.13)	4.48 (0.90)	9.33 (14.03)	0.59	4.18	4.00 to 4.40
Corn starch	412.4 (2.05)	5.13 (5.74)	11.51 (14.58)	0.64	5.05	4.40 to 5.10

TABLE XXX  
 PYRIDOXINE HCL FORMULATIONS WITH EMCOMPRESS AS A MATRIX

FORMULATIONS	WEIGHT	THICKNESS	HARDNESS	FRIABILITY	DISINTEGRATION TIME	
	mg	mm	Kg	Percent	min. sec. mean	range
<u>0.5% Disintegrant</u>						
Emcosoy	209.0 (0.44)	4.23 (2.35)	8.81 (20.41)	0.81	23.15	16.30-35.00
Explotab	216.7 (0.30)	4.12 (0.51)	8.04 (38.83)	CAP*	More than 1 hour	
Ac-Di-Sol	211.6 (0.50)	4.43 (3.38)	9.91 (10.26)	0.63	5.02	4.40- 5.25
<u>1.0% Disintegrant</u>						
Emcosoy	205.8 (0.61)	4.08 (3.88)	7.30 (15.74)	0.65	2.36	2.20- 2.45
Explotab	210.6 (0.41)	4.51 (5.74)	7.21 (19.48)	CAP	23.05	11.20-32.10
Ac-Di-Sol	212.1 (1.29)	4.13 (0.43)	7.36 (26.49)	CAP	4.17	3.25- 4.30
<u>2.0% Disintegrant</u>						
Explotab	211.0 (0.54)	4.69 (4.49)	7.92 (17.03)	CAP	4.10	3.10- 4.30
Ac-Di-Sol	205.3 (0.47)	3.95 (3.01)	6.45 (23.40)	CAP	1.01	0.45 - 1.20
Emcosoy	208.5 (0.76)	4.13 (0.39)	8.73 (15.16)	0.34	1.0	.50- 1.15
Corn Starch	218.5 (0.82)	4.12 (0.65)	10.93 (18.04)	1.49	—	more than 1 hr

\*CAP - CAPPING

TABLE XXXI

## PYRIDOXINE HCL FORMULATION WITH EMDEX AS A MATRIX

	WEIGHT	THICKNESS	HARDNESS	FRIABILITY	DISINTEGRATION TIME	
	mg	mm	Kg	Percent	mean <sup>min-sec</sup>	range
<u>0.5% Disintegrant</u>						
Emcosoy	202.3 (0.60)	5.36 (0.31)	12.47 (11.73)	0.29	5.10	4.50 to 5.30
Ac-Di-Sol	207.8 (0.50)	5.37 (0.37)	13.38 (11.61)	0.39	3.51	3.40 to 4.00
Explotab	205.9 (0.44)	5.34 (0.31)	12.44 (14.33)	0.35	5.09	4.40 to 6.00
<u>1.0% Disintegrant</u>						
Emcosoy	209.3 (0.47)	5.37 (0.40)	13.05 (11.47)	0.29	4.02	3.50 to 4.10
Ac-Di-Sol	213.3 (0.80)	5.39 (0.53)	12.58 (12.66)	0.45	4.05	3.50 to 4.30
Explotab	212.9 (0.52)	5.65 (2.82)	12.41 (10.02)	0.38	4.11	3.20 to 4.30
<u>3.0% Disintegrant</u>						
Emcosoy	214.5 (0.47)	5.61 (0.75)	10.88 (12.91)	0.44	3.00	2.80 to 3.20
Corn starch	226.1 (0.79)	5.73 (0.40)	13.37 (12.19)	0.33	6.00	5.10 to 7.25

TABLE XXXII

## ASCORBIC ACID FORMULATION WITH EMCOMPRESS AS A MATRIX

	WEIGHT	THICKNESS	HARDNESS	FRIABILITY	DISINTEGRATION TIME	
	mg	mm	Kg	Percent	mean mins.-sec	range
<u>0.5% Disintegrant</u>						
Emcosoy	397.9 (0.88)	4.05 (0.80)	3.50 (60.29)	CAP*	1.15	0.58 to 1.30
Ac-Di-Sol	402.2 (1.36)	4.07 (1.26)	3.35 (67.07)	CAP	0.56	0.40 to 1.20
<u>1.0% Disintegrant</u>						
Emcosoy	399.9 (5.47)	40.60(2.16)	4.81 (34.74)	CAP	0.51	0.25 to 1.25
Ac-Di-Sol	466.1 (4.67)	4.39 (3.24)	3.00 (53.32)	CAP	0.31	0.20 to 0.40
Explotab	377.4 (1.97)	3.90 (1.44)	4.61 (15.17)	2.05	2.21	1.20 to 3.25
<u>2.0% Disintegrant</u>						
Emcosoy	378.0(1.92)	4.08 (1.06)	4.25 (19.92)	1.74	0.46	0.40 to 0.55
Ac-Di-Sol	388.0(0.96)	4.13 (0.85)	4.9 (13.48)	1.67	0.46	0.40 to 0.55
Explotab	395.3 (0.79)	4.18 (0.83)	5.32 (15.46)	1.44	2.57	2.20 to 3.20
Corn Starch	408.3 (0.94)	4.20 (0.63)	5.9 (18.66)	1.67	22.18	18.10 to 24.30
<u>3.0% Disintegrant</u>						
Emcosoy	393 (3.30)	4.07 (2.24)	5.37(33.30)	1.39	0.30	0.25 to 0.40
Corn Starch	440.2 (1.58)	4.34 (1.13)	2.11 (84.24)	CAP	14.10	12.30 to 15.00

\*CAPPING

TABLE XXXIII

Formulation: Riboflavin 0.57%  
 Pyridoxine Hcl 0.86%  
 Niacin 5.70%  
 Ascorbic acid 17.10%  
 Disintegrant 0.5% or 1.0% or 3.0%  
 Stearic acid 2.5%  
 Emcompress ad 100%

## Dissolution data:

Formulation	$t_{50}$ (min)	$t_{75}$ (min)	$t_{90}$ (min)
<u>0.5% Disintegrant</u>			
Emcosoy	8.45	11.75	14.00
	8.75	12.40	14.60
	10.10	12.80	14.75
Ac-Di-Sol	3.70	4.80	6.75
	3.80	5.00	7.65
	4.20	5.30	8.40
Explotab	10.10	15.10	17.40
	10.25	16.30	18.30
	11.50	17.30	20.20
<u>1.0% Disintegrant</u>			
Emcosoy	2.60	4.10	6.80
	6.20	8.30	10.55
	9.30	13.95	16.90
Ac-Di-Sol	1.50	2.60	3.60
	1.90	2.75	3.60
	1.90	3.10	5.20
Explotab	5.10	6.25	7.15
	5.20	7.20	8.90
	6.75	8.00	10.80
<u>3.0% Disintegrant</u>			
Emcosoy	1.55	2.95	3.60
	2.00	2.95	3.60
Corn starch	18.25	> 30.00	> 30.00
	20.55	> 30.00	> 30.00

## Dissolution profiles.

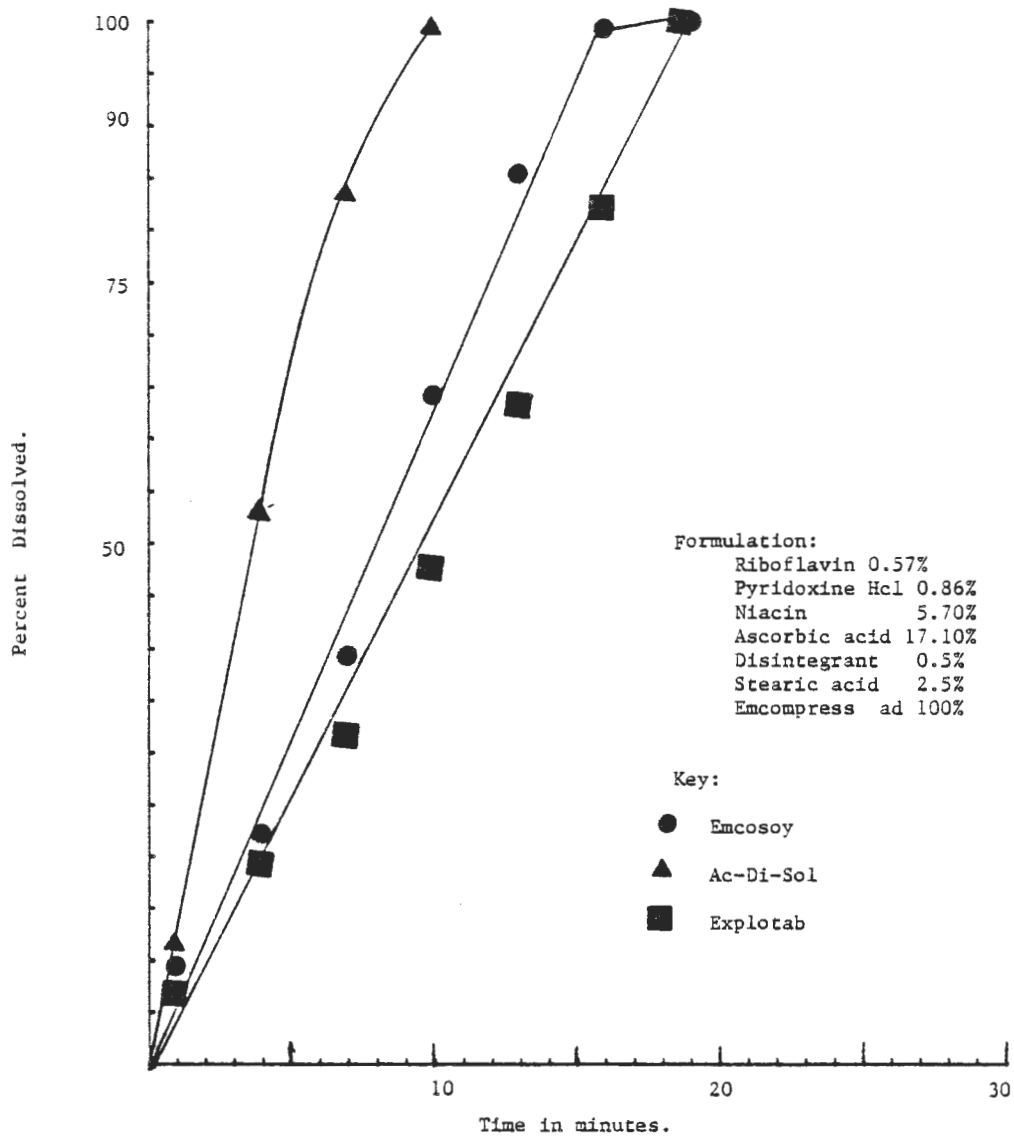
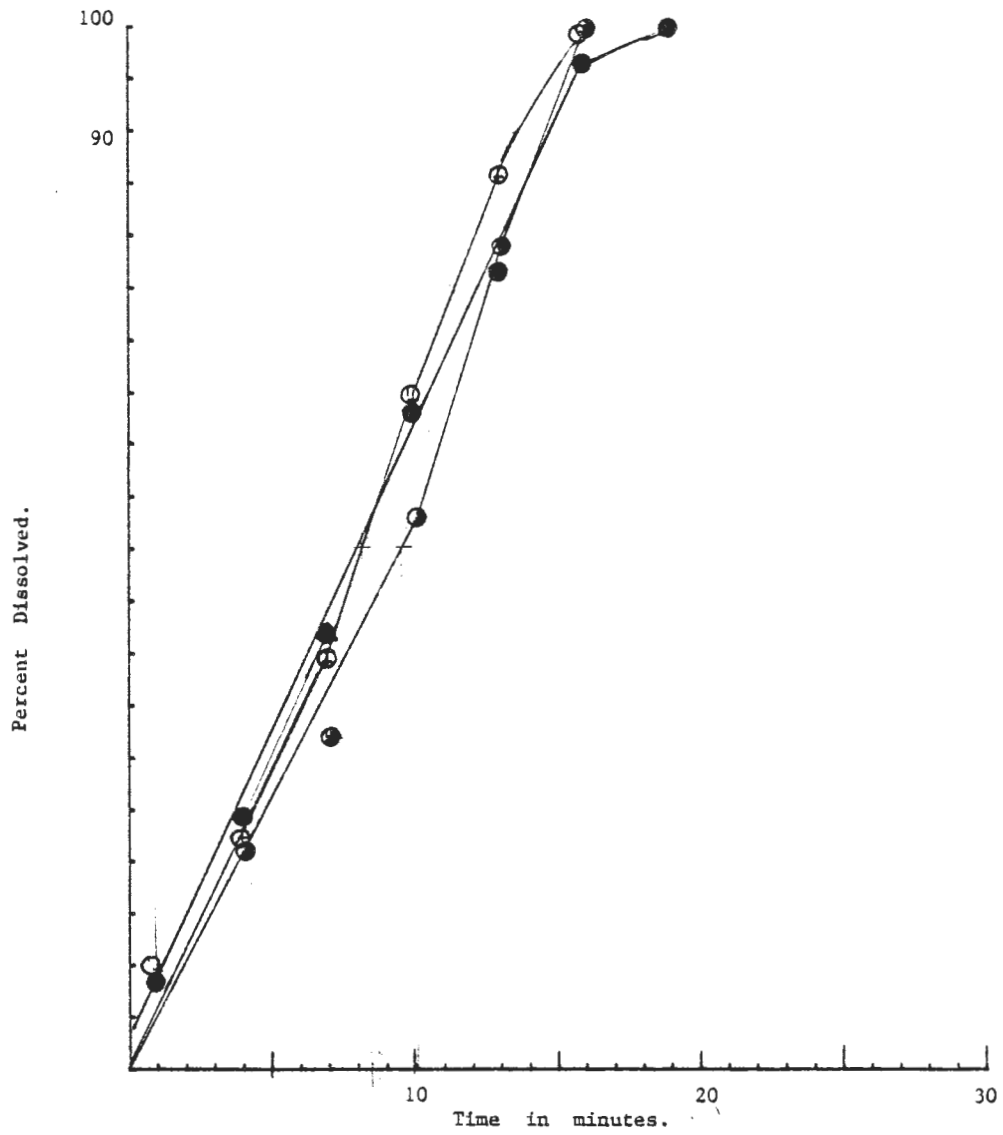


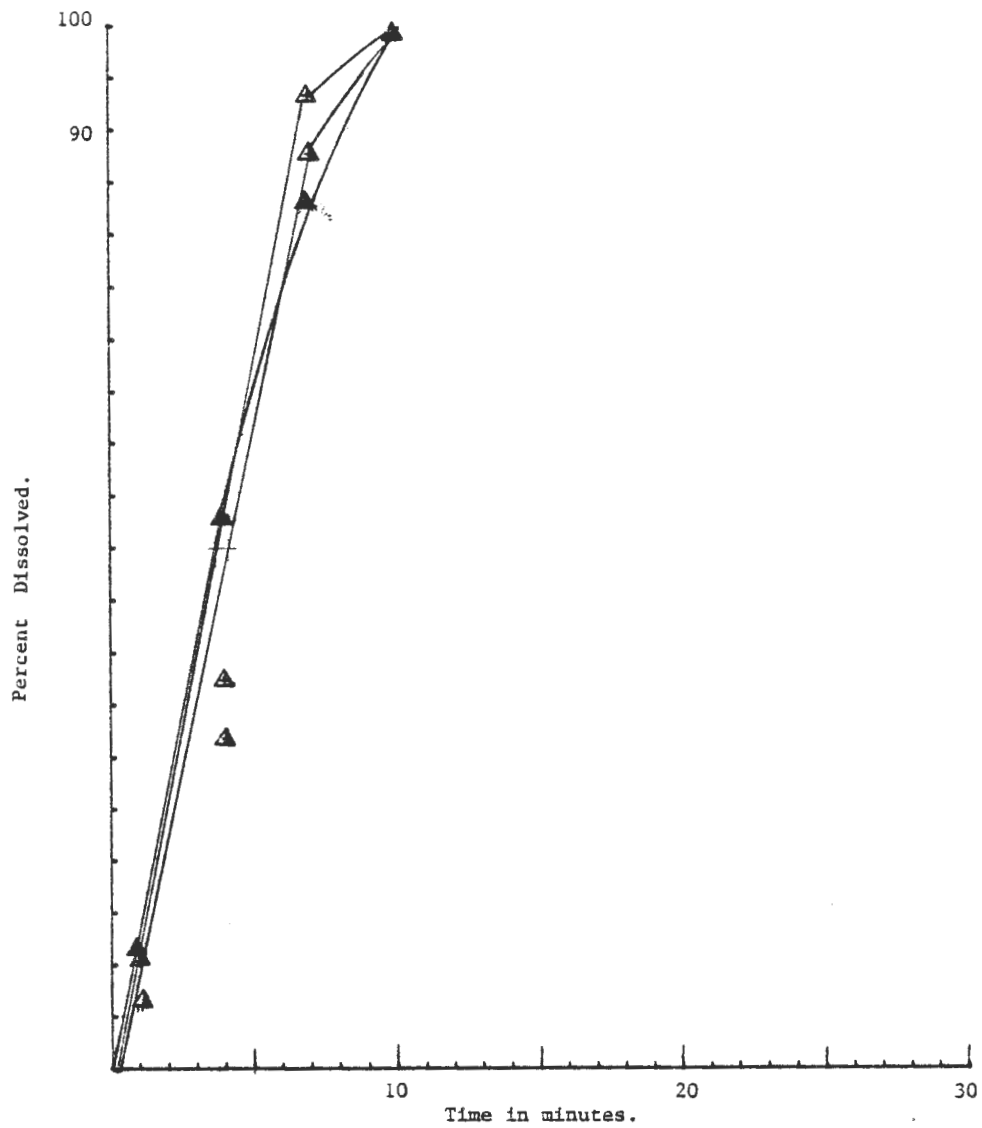
Fig. 19



Dissolution profile of multi vitamin formulation containing 0.5% of Emcosoy  
Replicate 1 ● ,Replicate 2 ⊕ ,Replicate 3 ⊙ , Emcompress as a matrix.

Fig. 20





Dissolution profile of multi vitamin formulation containing 0.5% of Ac-Di-SOL  
Replicate 1 ▲ ,Replicate 2 ▲ ,Replicate 3 ▲ Emcompress as a matrix.

Fig. 21

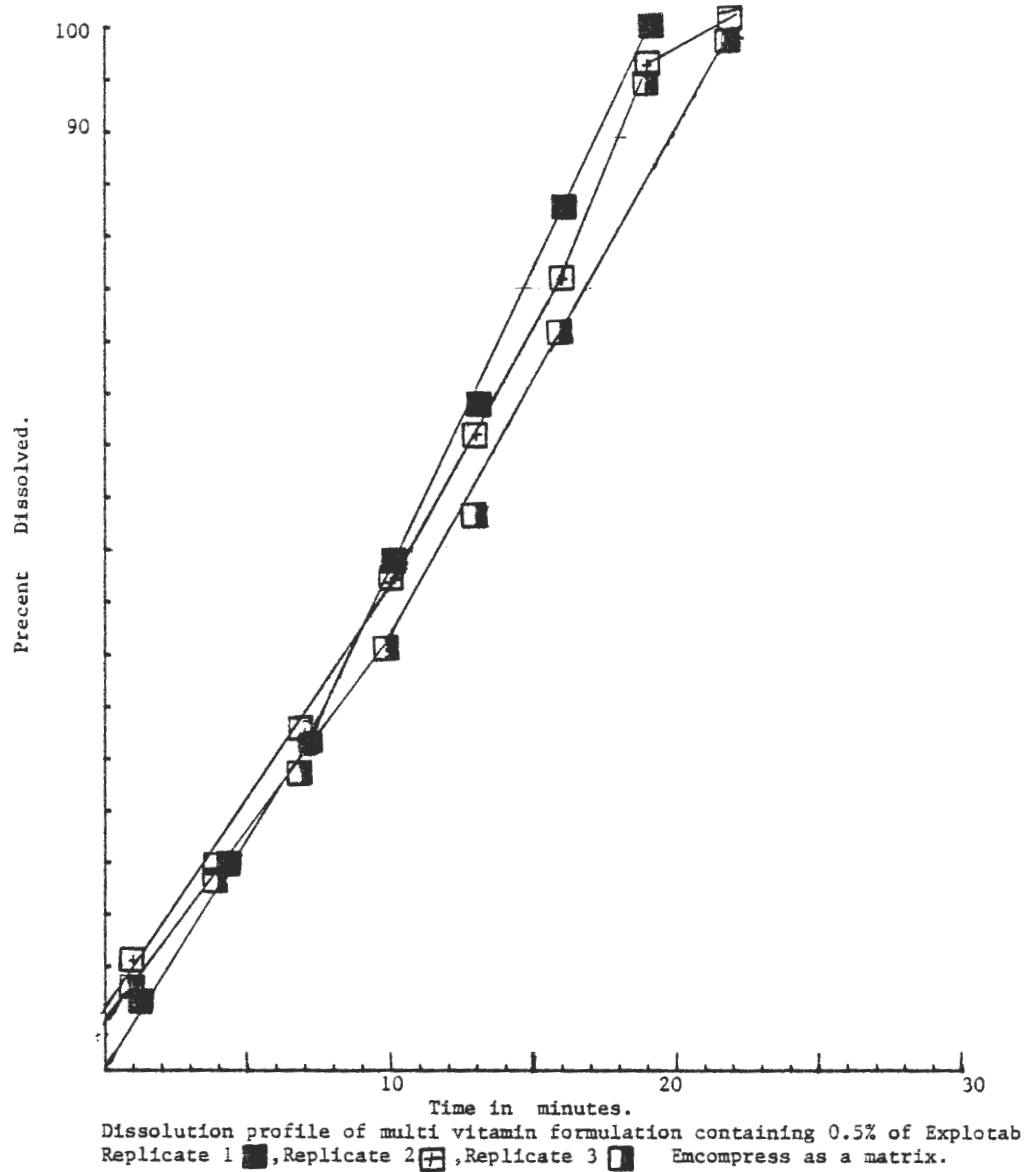


Fig. 22

## Dissolution profiles.

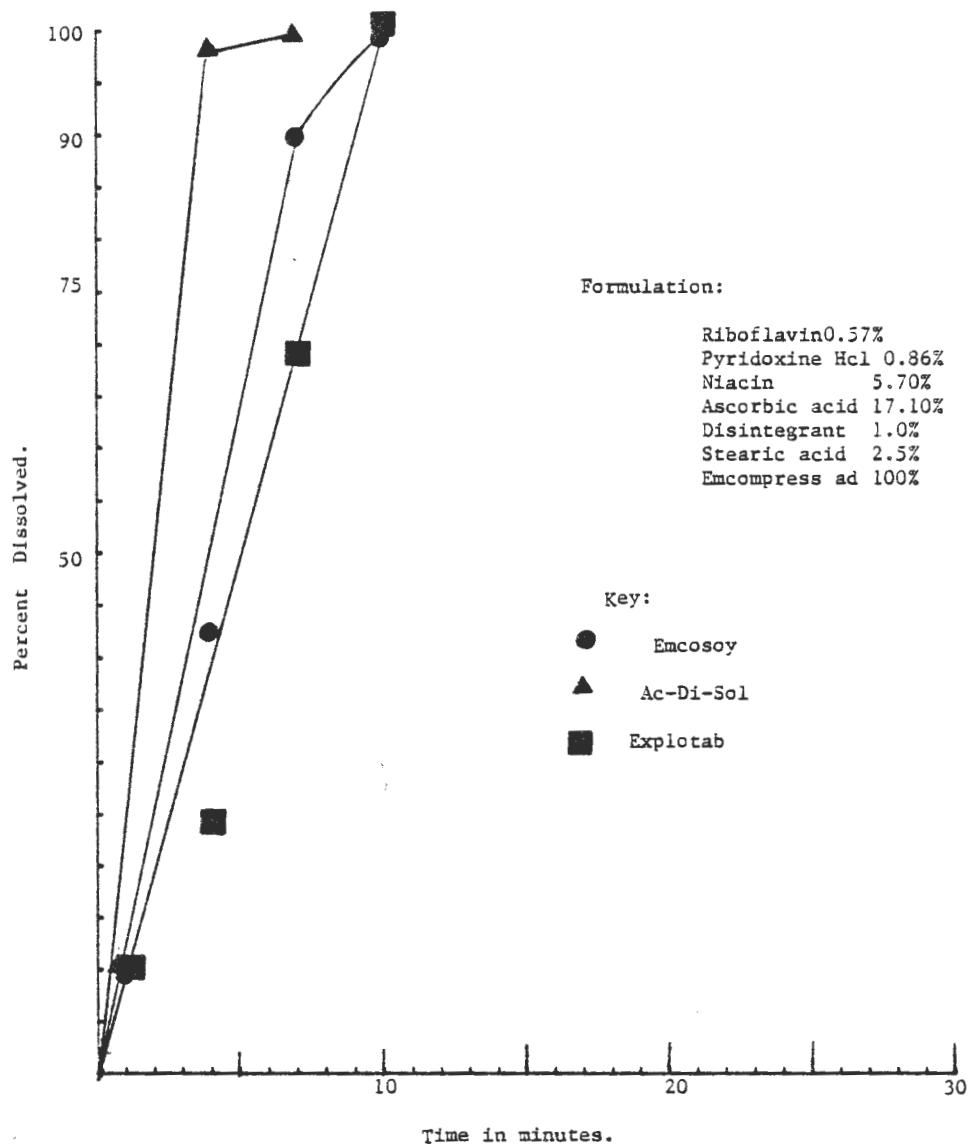


Fig. 23

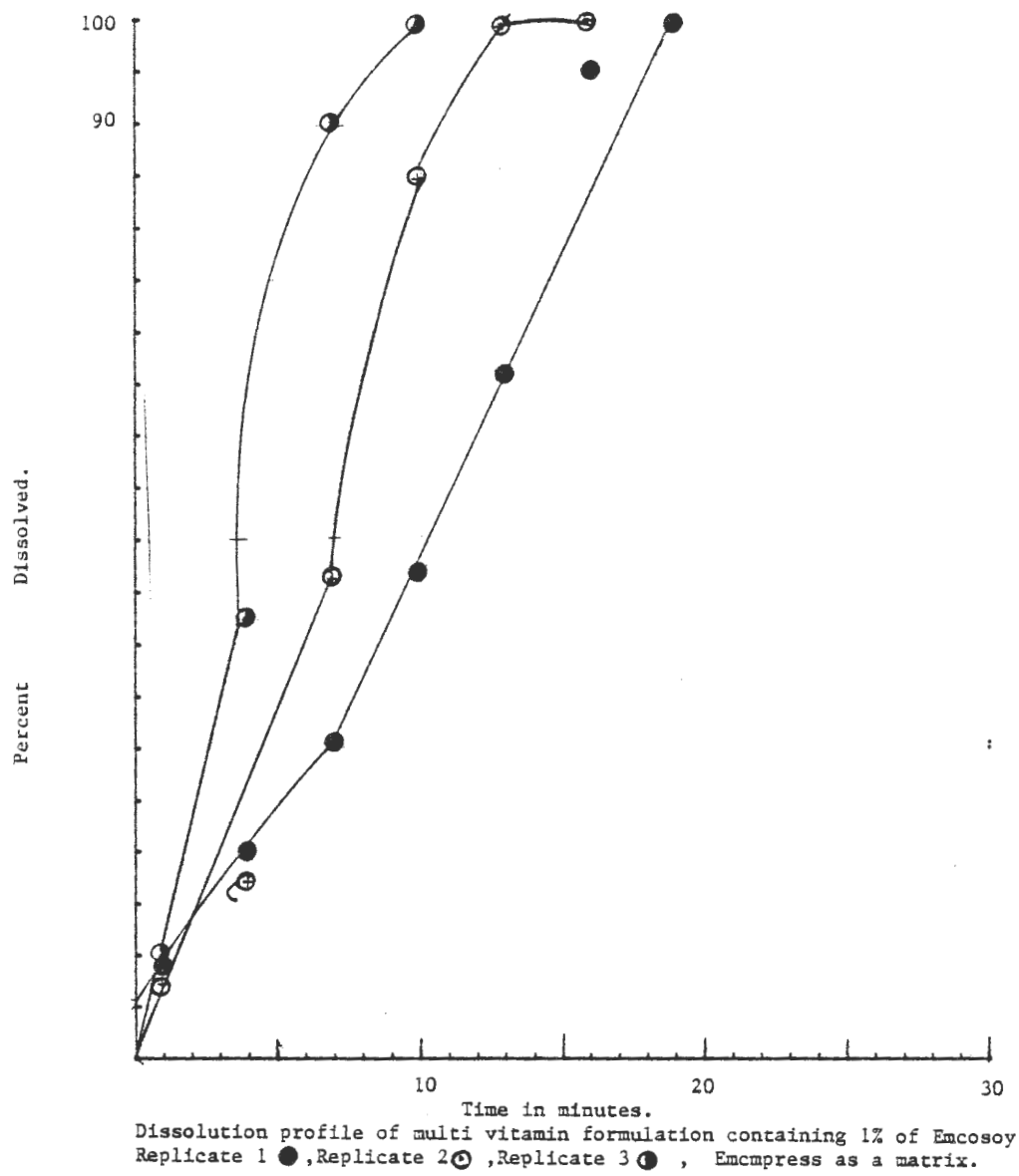


Fig. 24

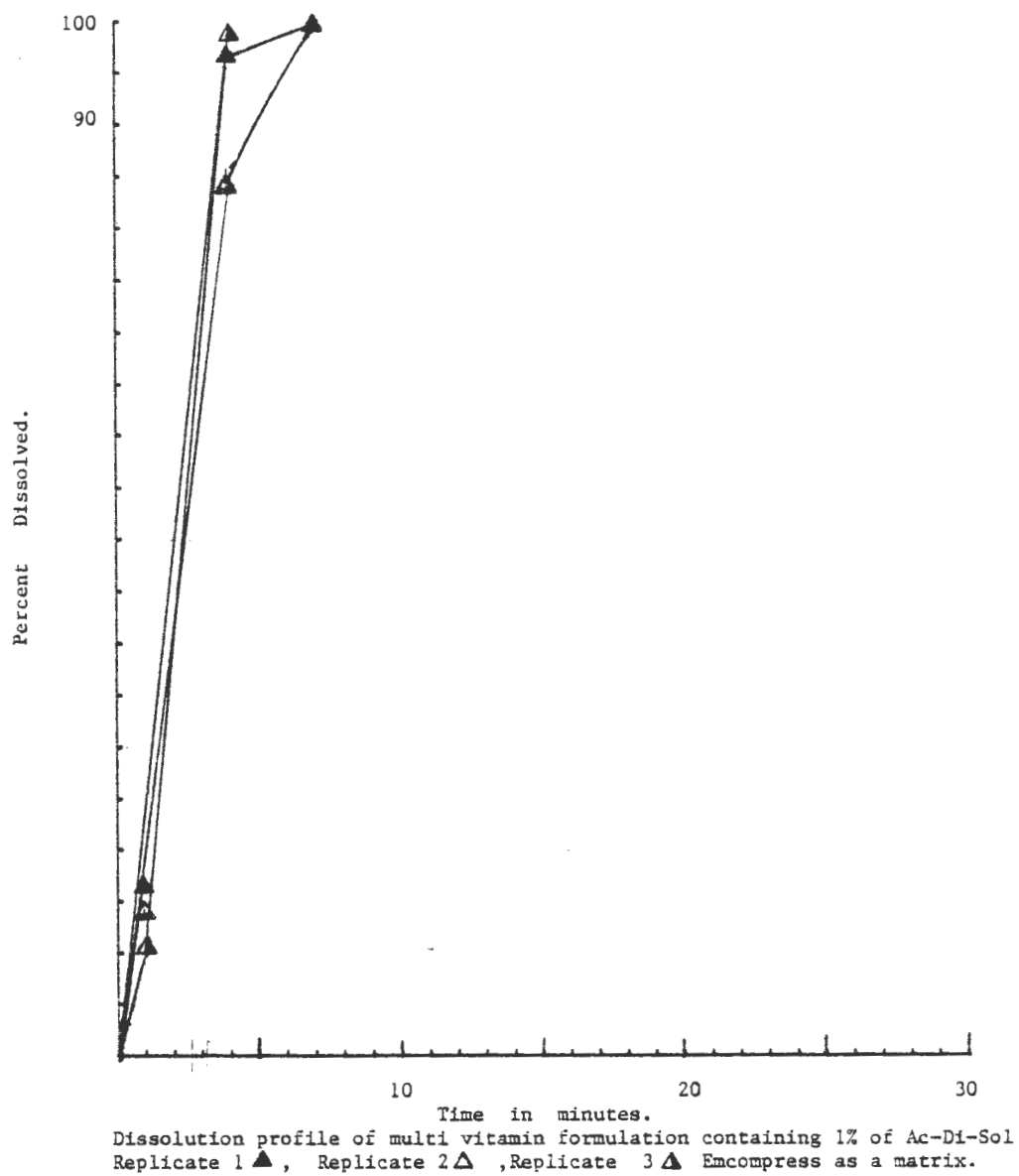


Fig. 25

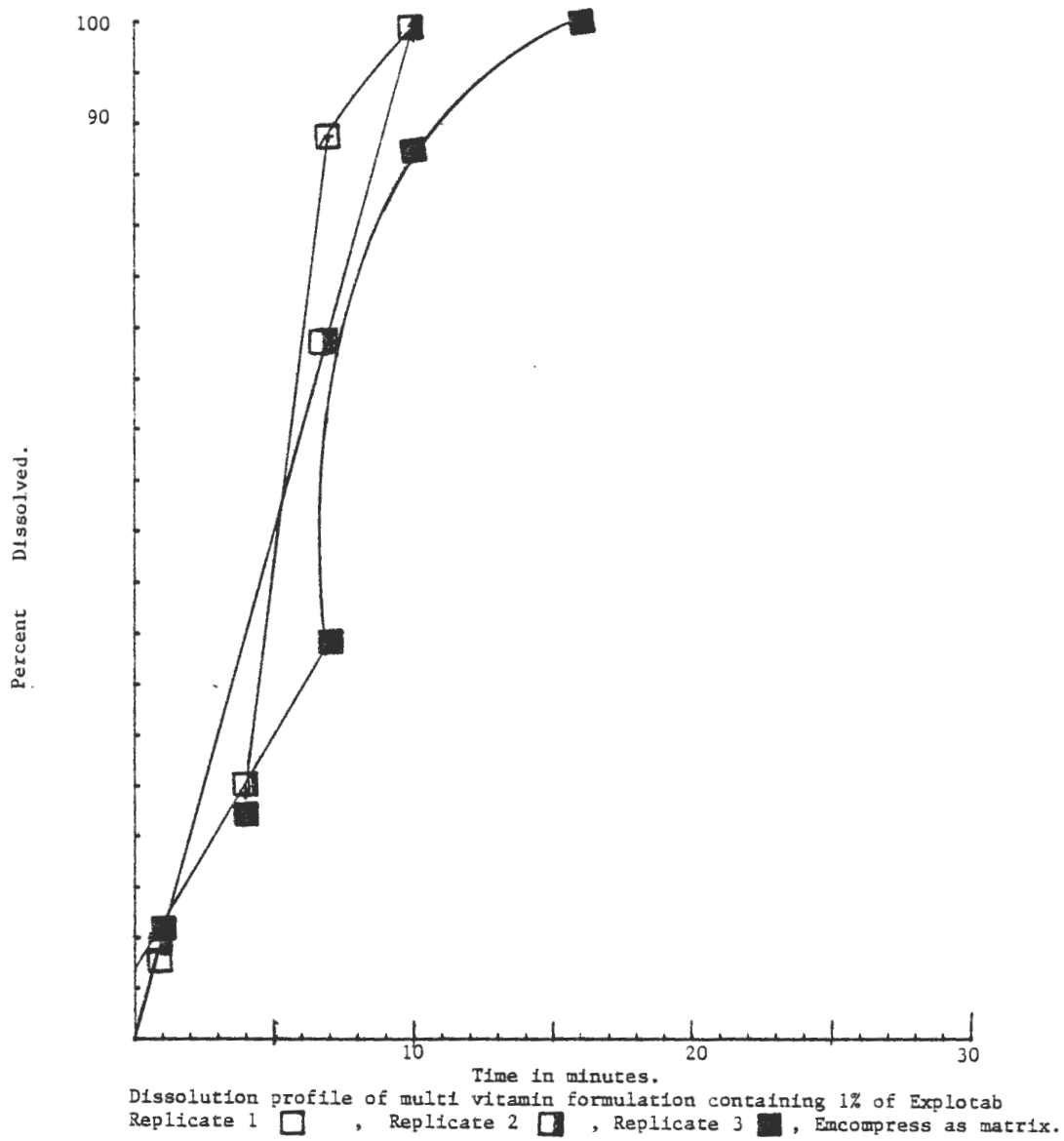


Fig. 26

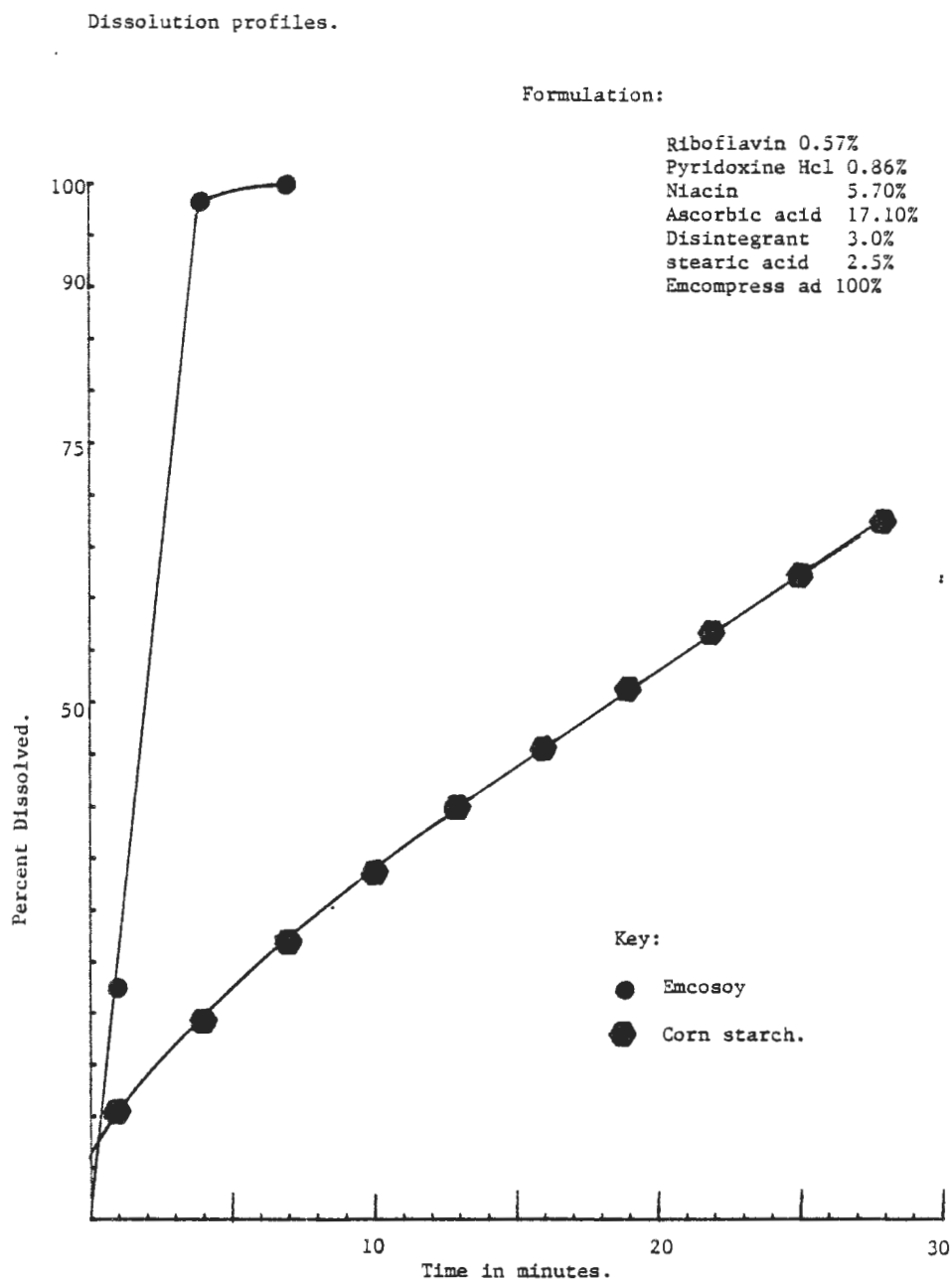


Fig. 27

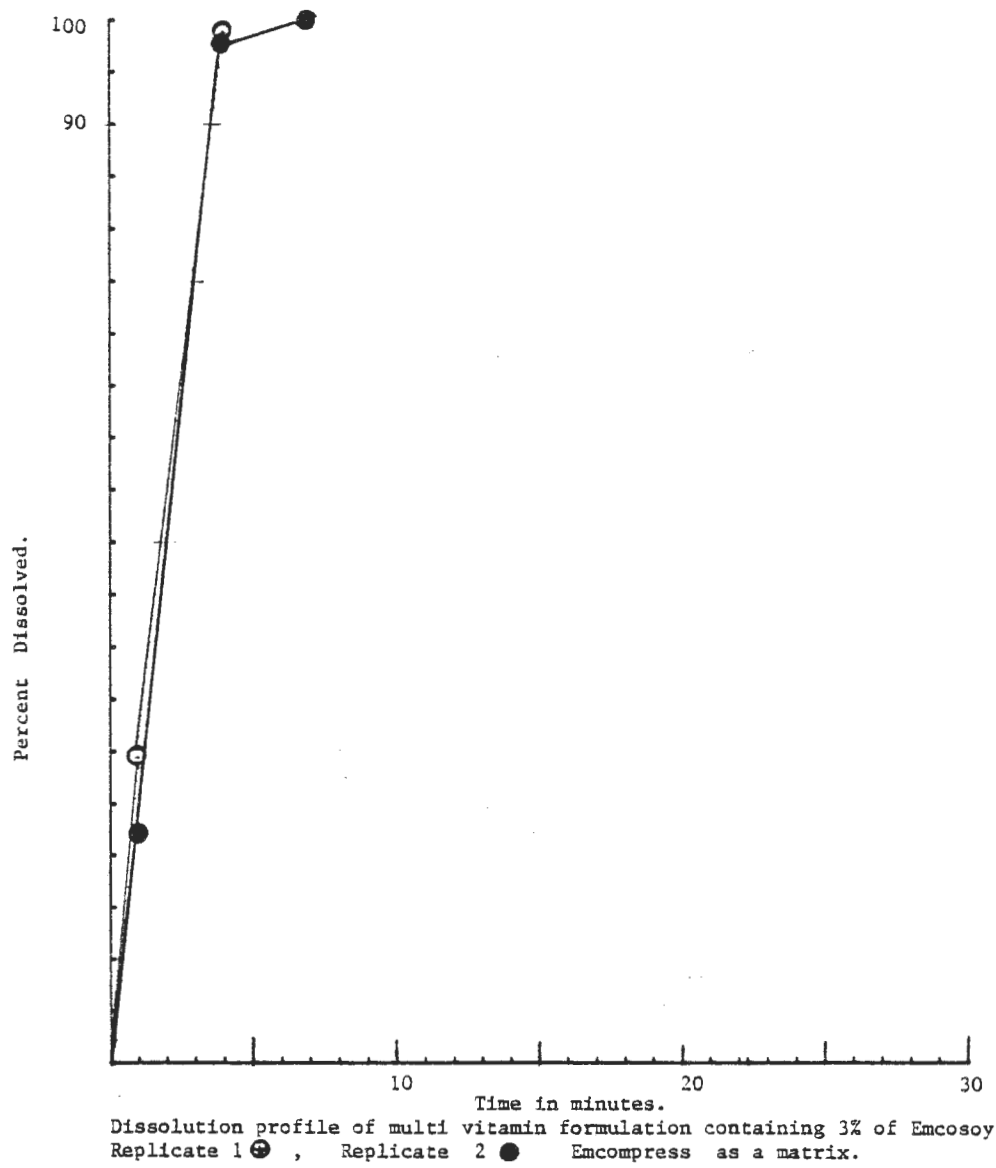
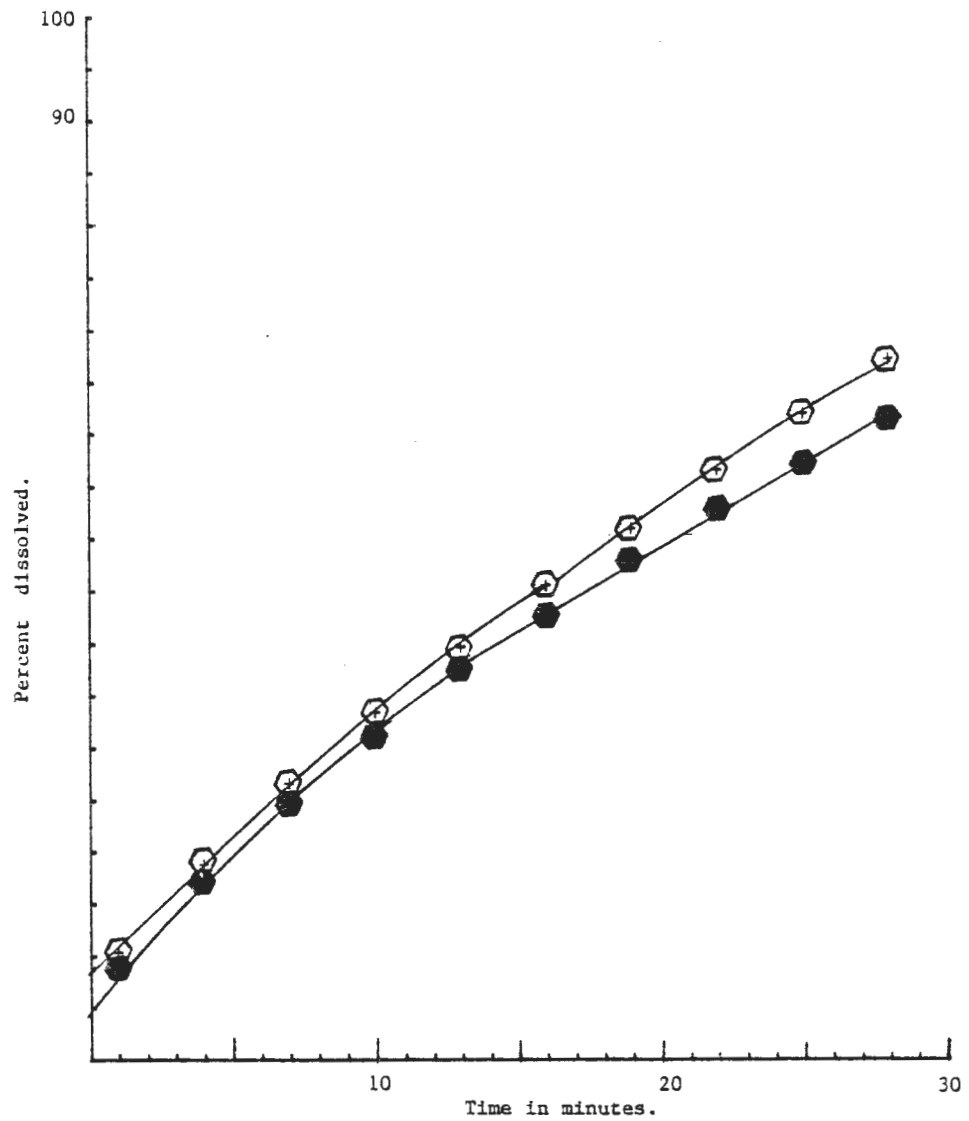


Fig. 28





Dissolution profile of multi vitamin formulation containing 3% of Corn starch  
Replicate 1 ⊕ , Replicate 2 ● Emcompress as a matrix.

Fig. 29

TABLE XXXIV

Formulation: Riboflavin 0.57%  
 Pyridoxine Hcl 0.86%  
 Niacin 5.70%  
 Ascorbic acid 17.10%  
 Disintegrant 0.5% or 1.0% or 3.0%  
 Stearic acid 2.5%  
 Emdex ad 100%

Dissolution data:

Formulation	$t_{50}$ (min)	$t_{75}$ (min)	$t_{90}$ (min)
<u>0.5% Disintegrant</u>			
Emcosoy	2.95	3.55	4.75
	2.95	3.55	4.75
	3.20	3.75	4.75
Ac-Di-Sol	3.00	4.80	6.65
	2.60	4.20	6.00
	2.40	3.90	5.30
Explotab	2.40	3.60	6.00
	2.65	4.00	5.70
	2.85	4.20	5.30
<u>1.0 % Disintegration</u>			
Emcosoy	2.80	5.20	6.75
	3.15	5.40	5.20
	3.25	5.40	6.75
Ac-Di-Sol	2.45	3.75	5.20
	2.55	3.75	5.20
	2.75	4.25	6.10
Explotab	3.55	4.20	6.45
	3.60	4.60	6.65
	3.60	4.60	6.65
<u>3.0% Disintegrant</u>			
Emcosoy	2.75	3.85	5.20
	2.75	4.05	5.40
Corn starch	3.20	5.60	8.00
	3.75	6.50	8.90

## Dissolution profiles.

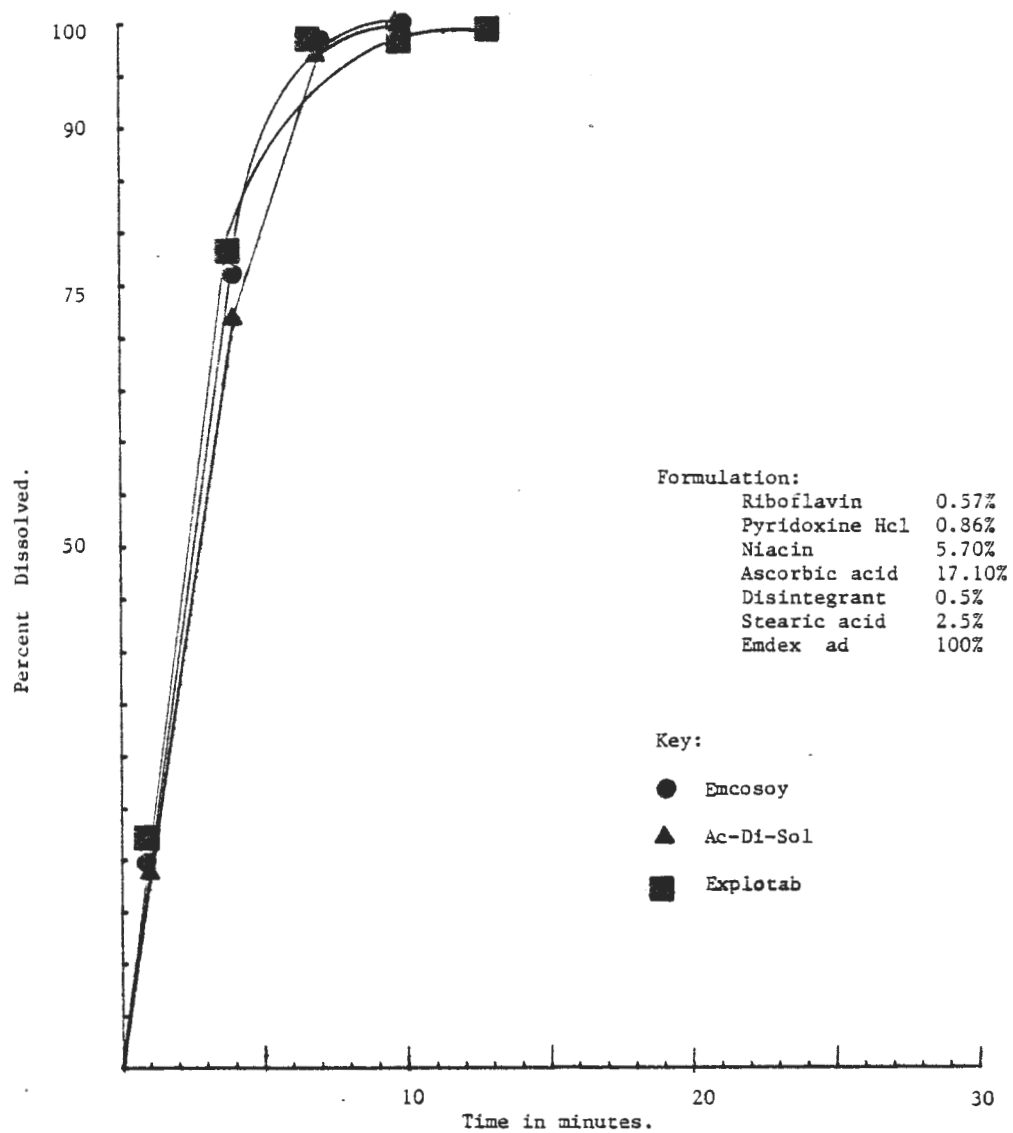


Fig. 30

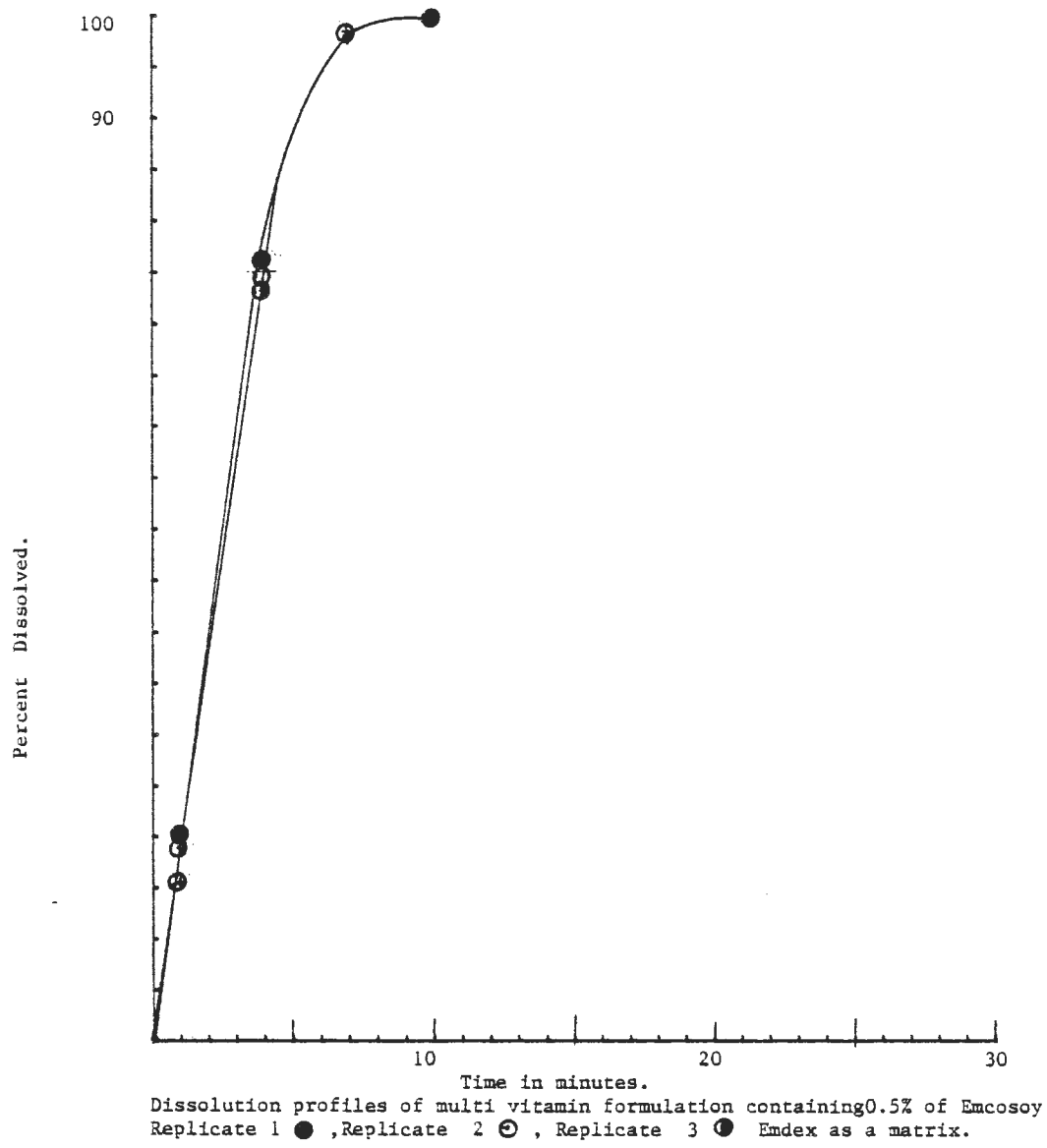


Fig. 31

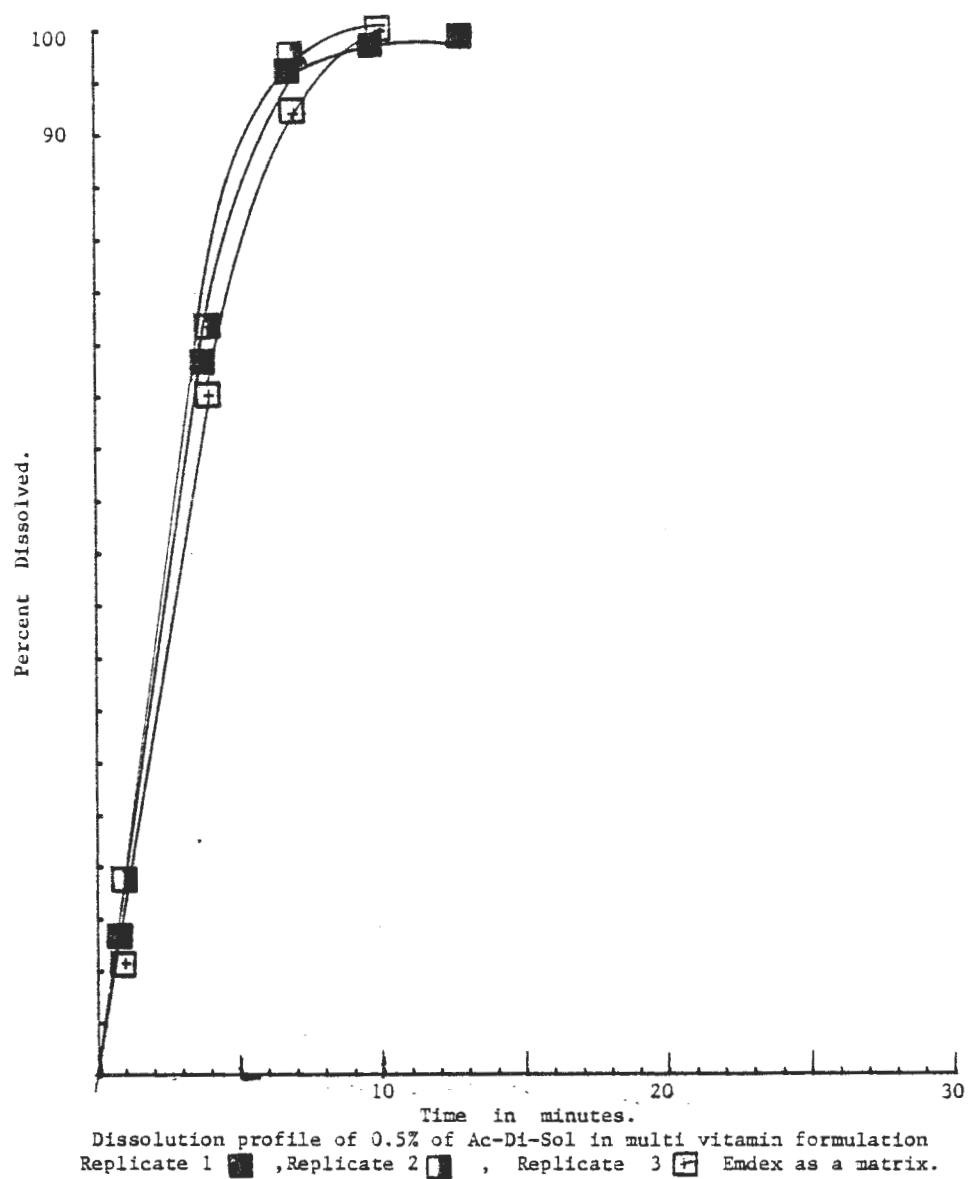


Fig. 32

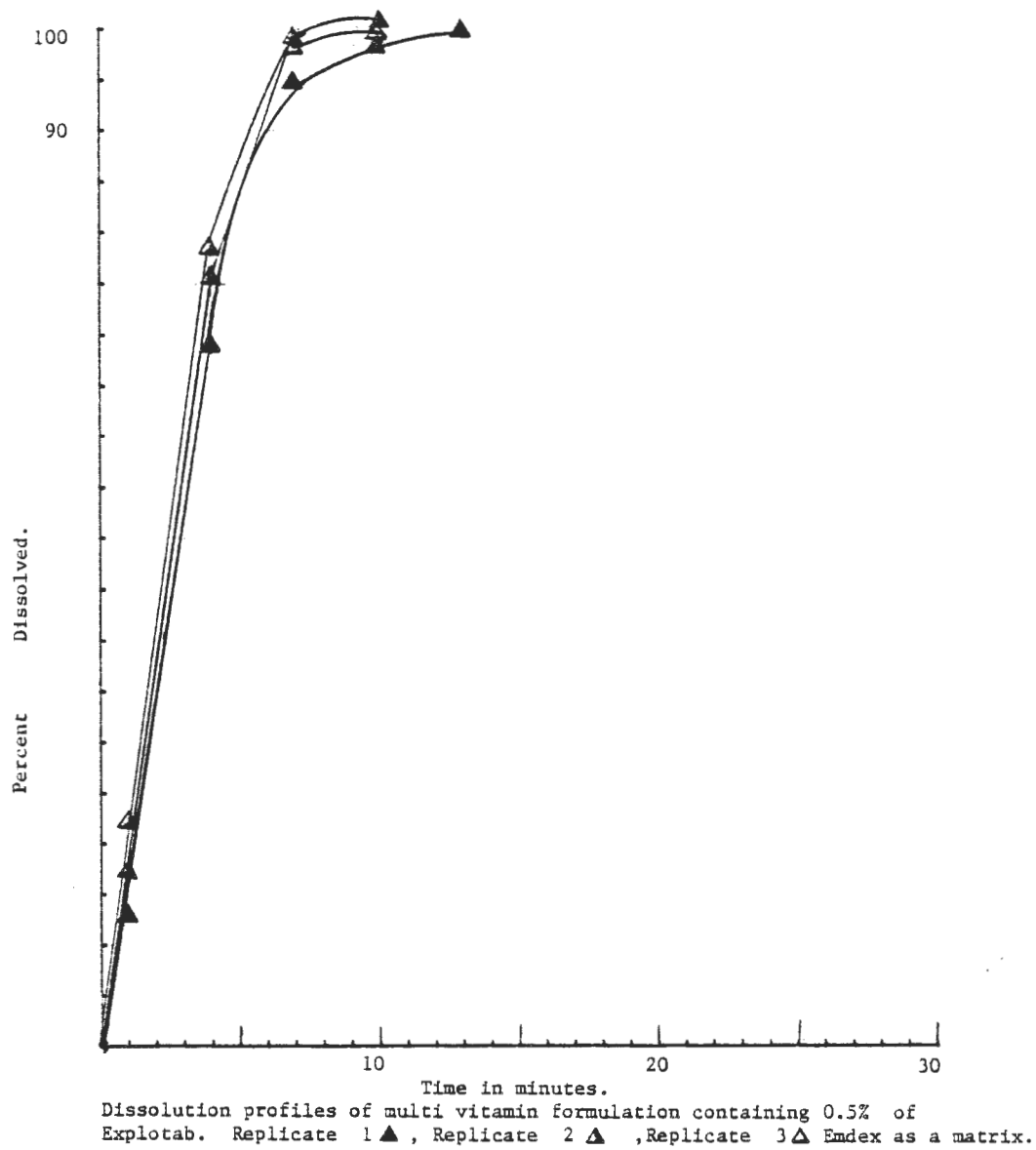


Fig. 33

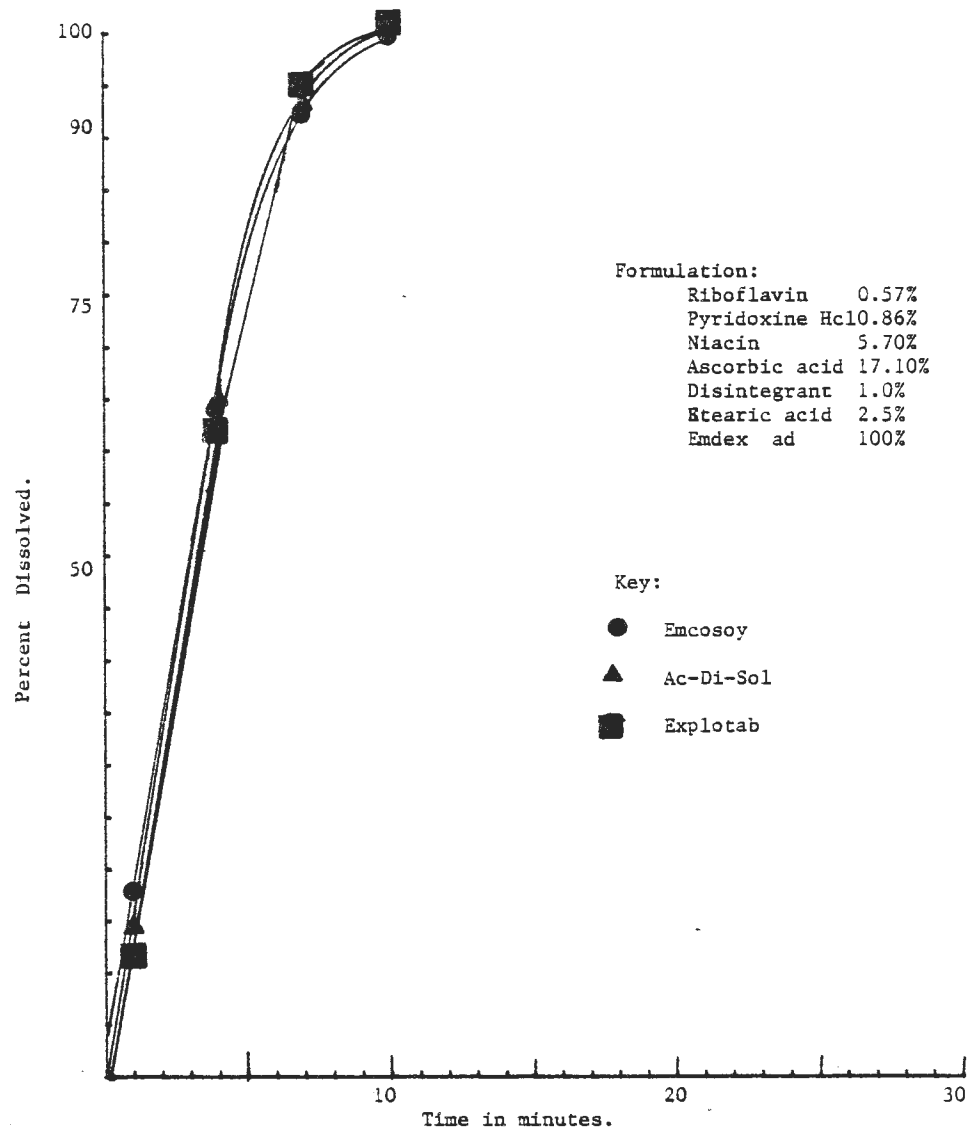


Fig. 34

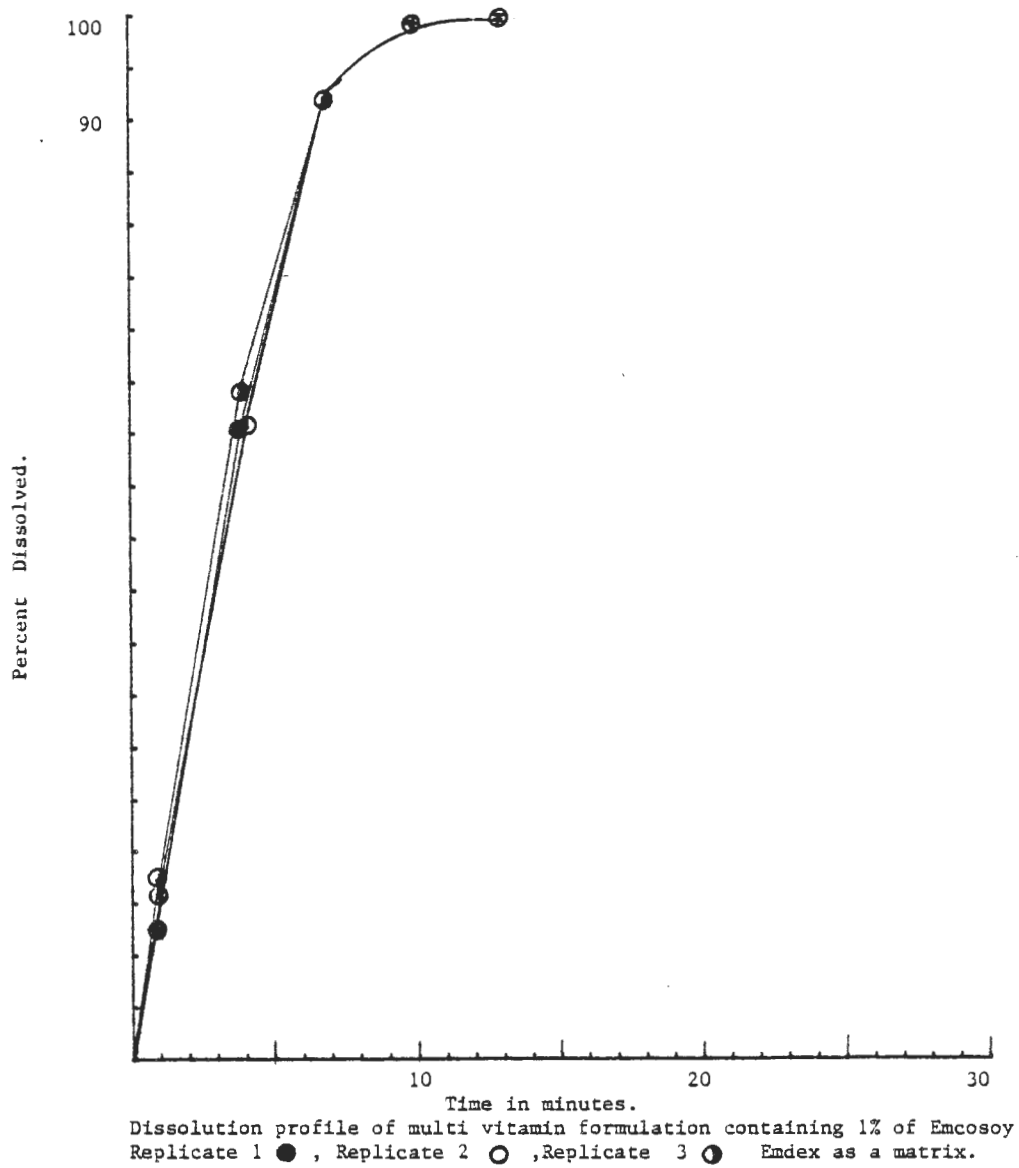
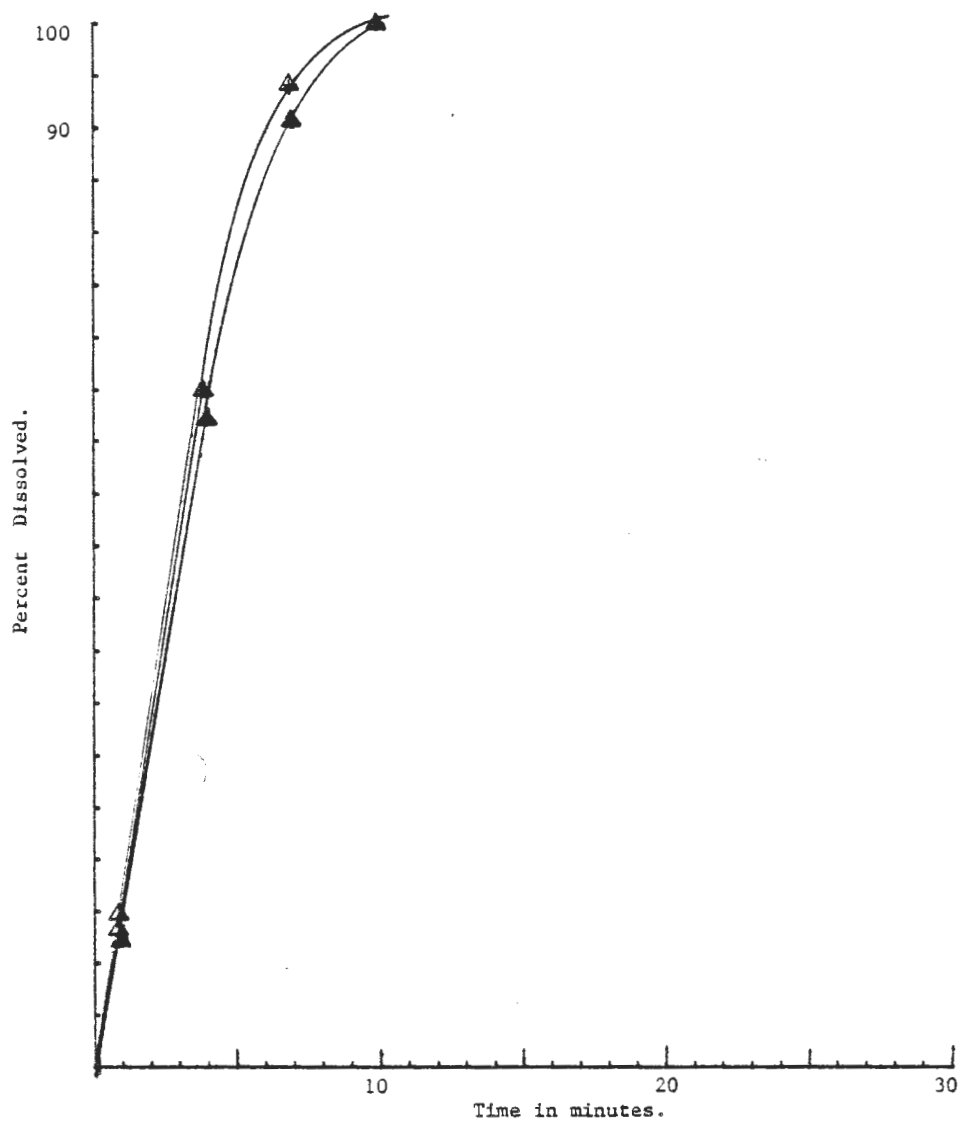


Fig. 35





Dissolution profile of multi vitamin formulation containing 1% of Ac-Di-Sol  
Replicate 1 ▲ , Replicate 2 ▲ , Replicate 3 ▲ Emdex as a matrix.

Fig. 36

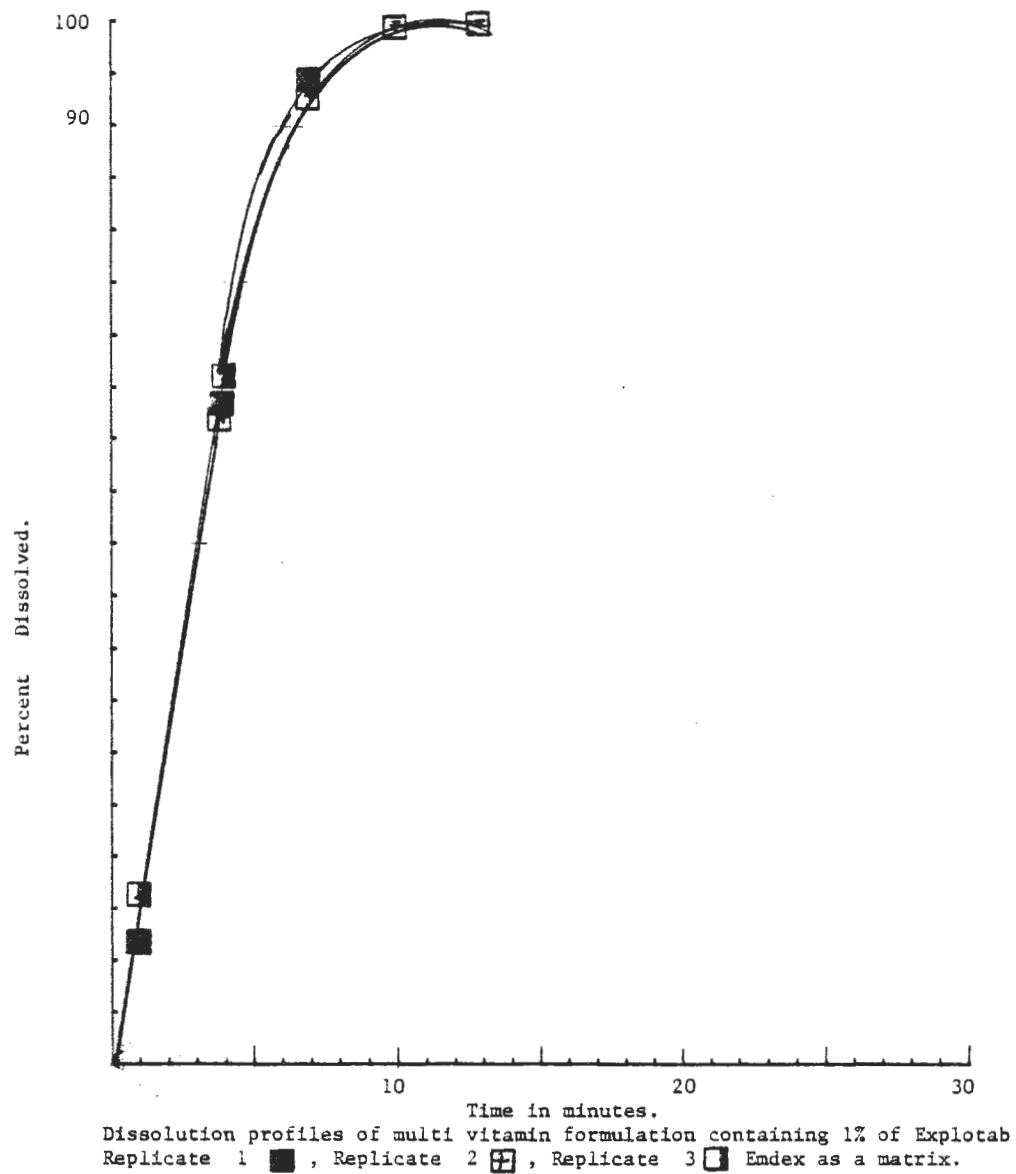


Fig. 37

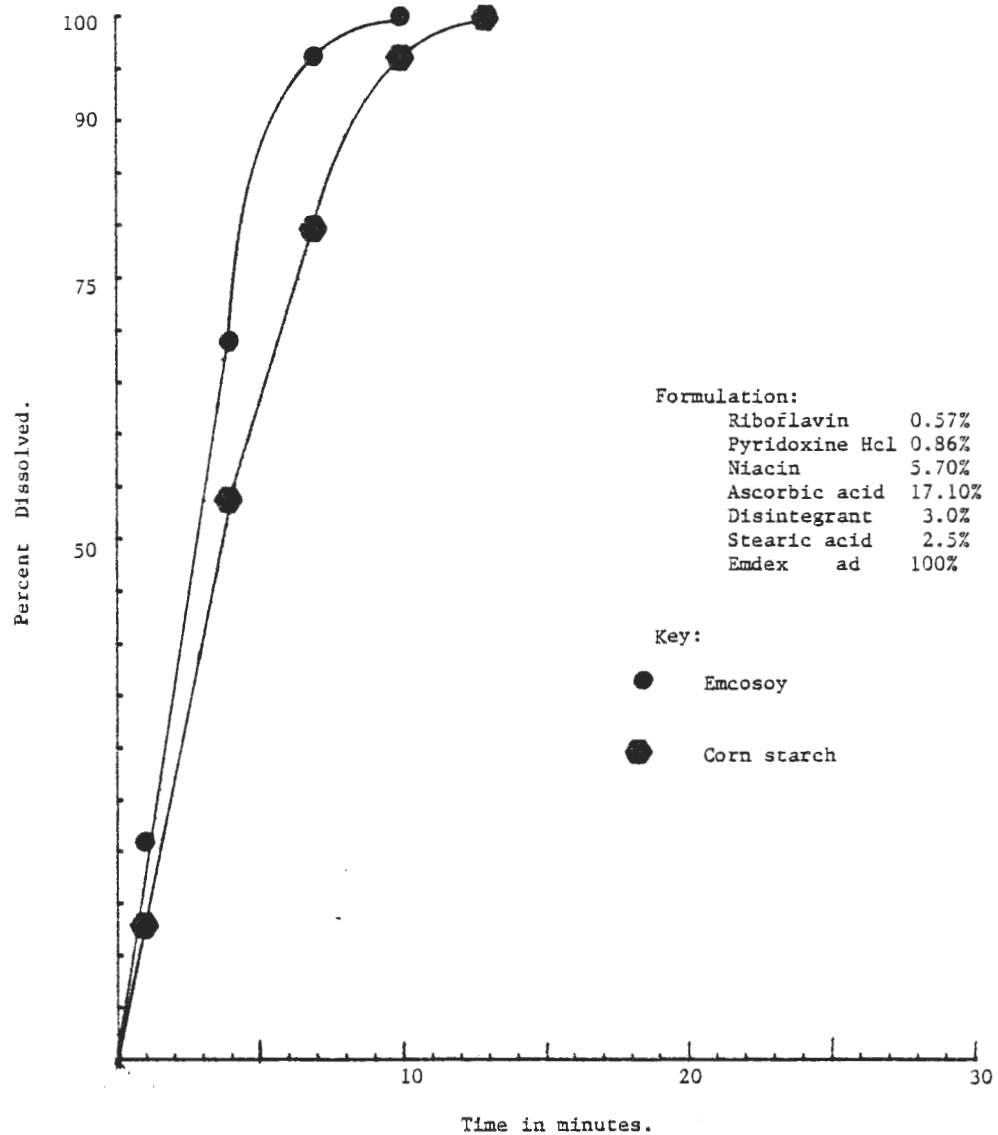


Fig. 38

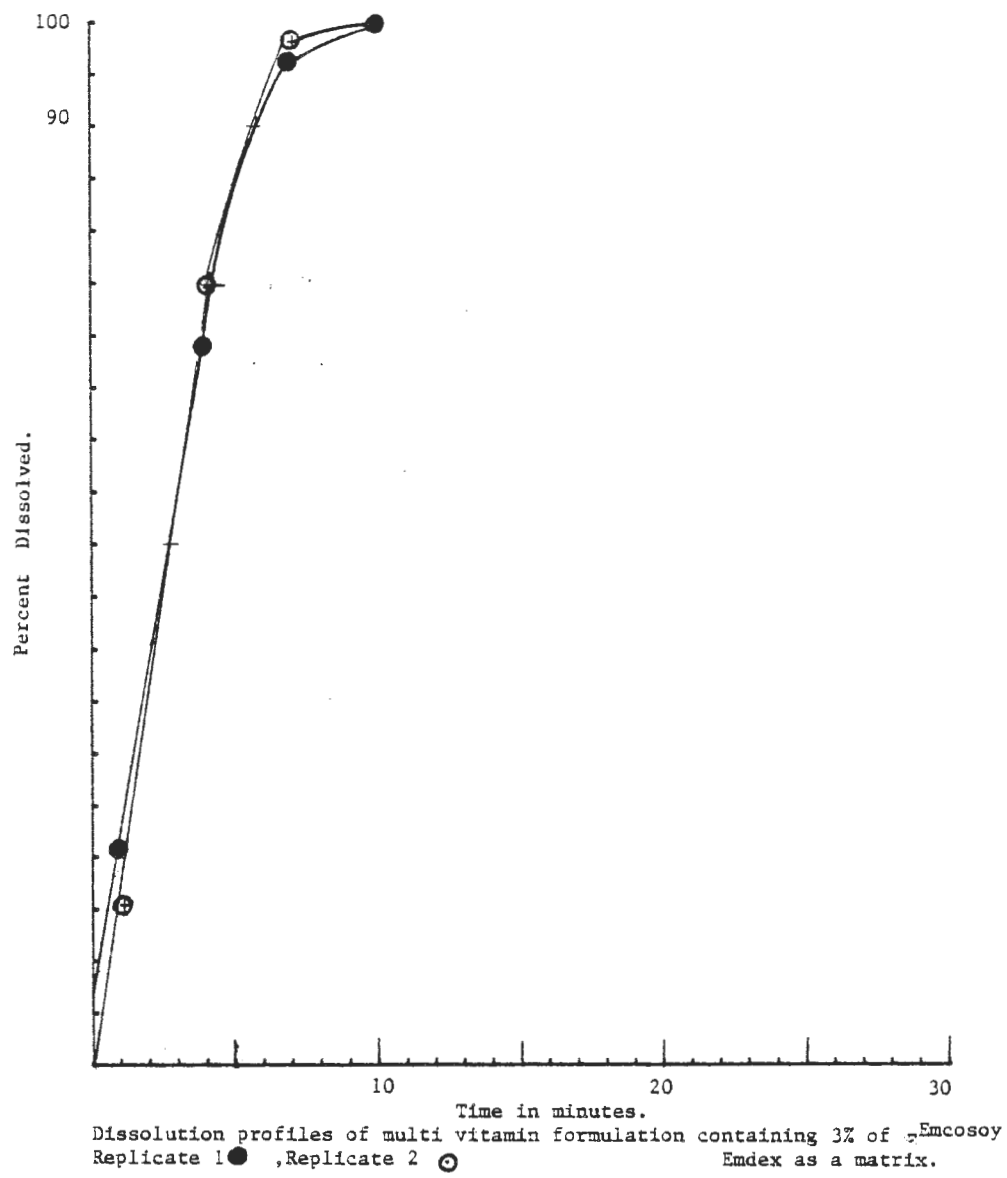
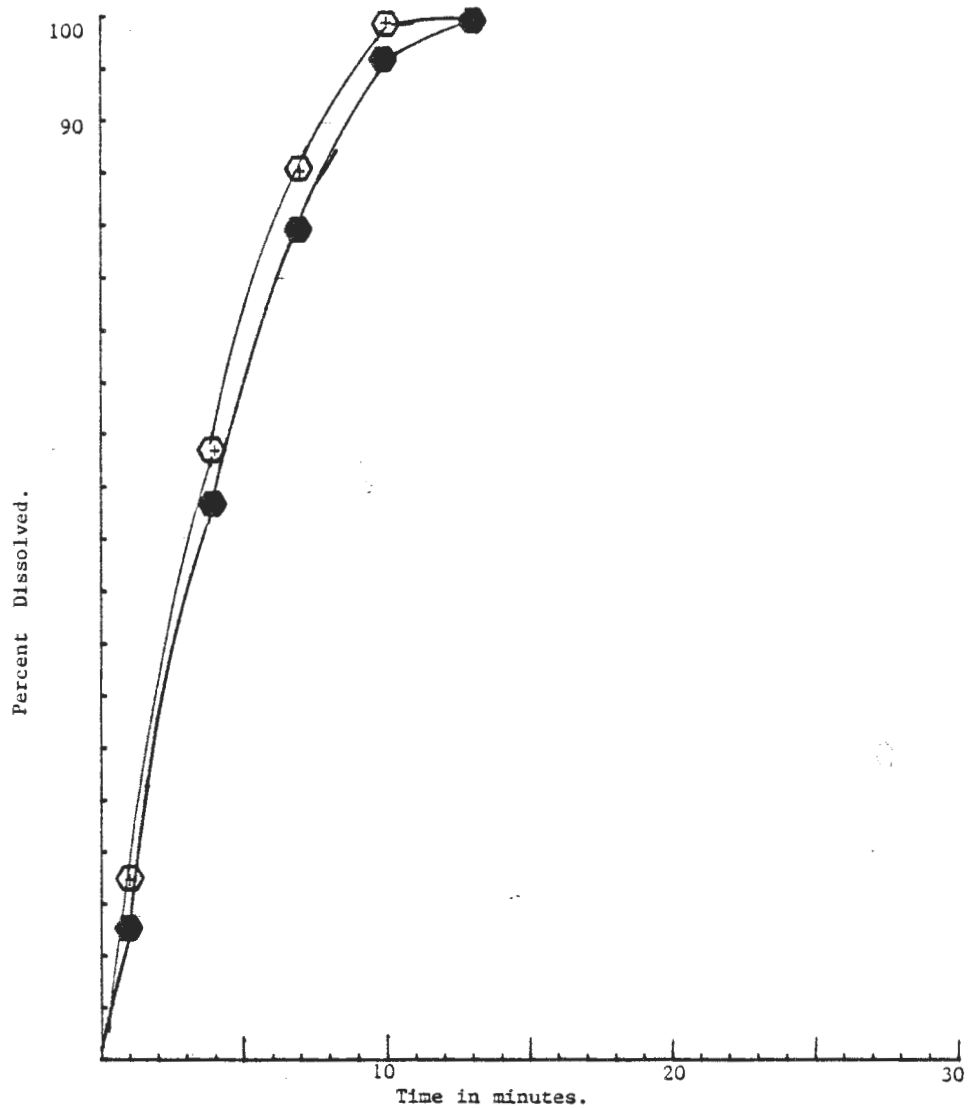


Fig. 39



Dissolution profiles of multi vitamin formulation containing 3% of Corn starch  
Replicate 1  $\circ$ , Replicate 2  $\bullet$  Emdex as a matrix.

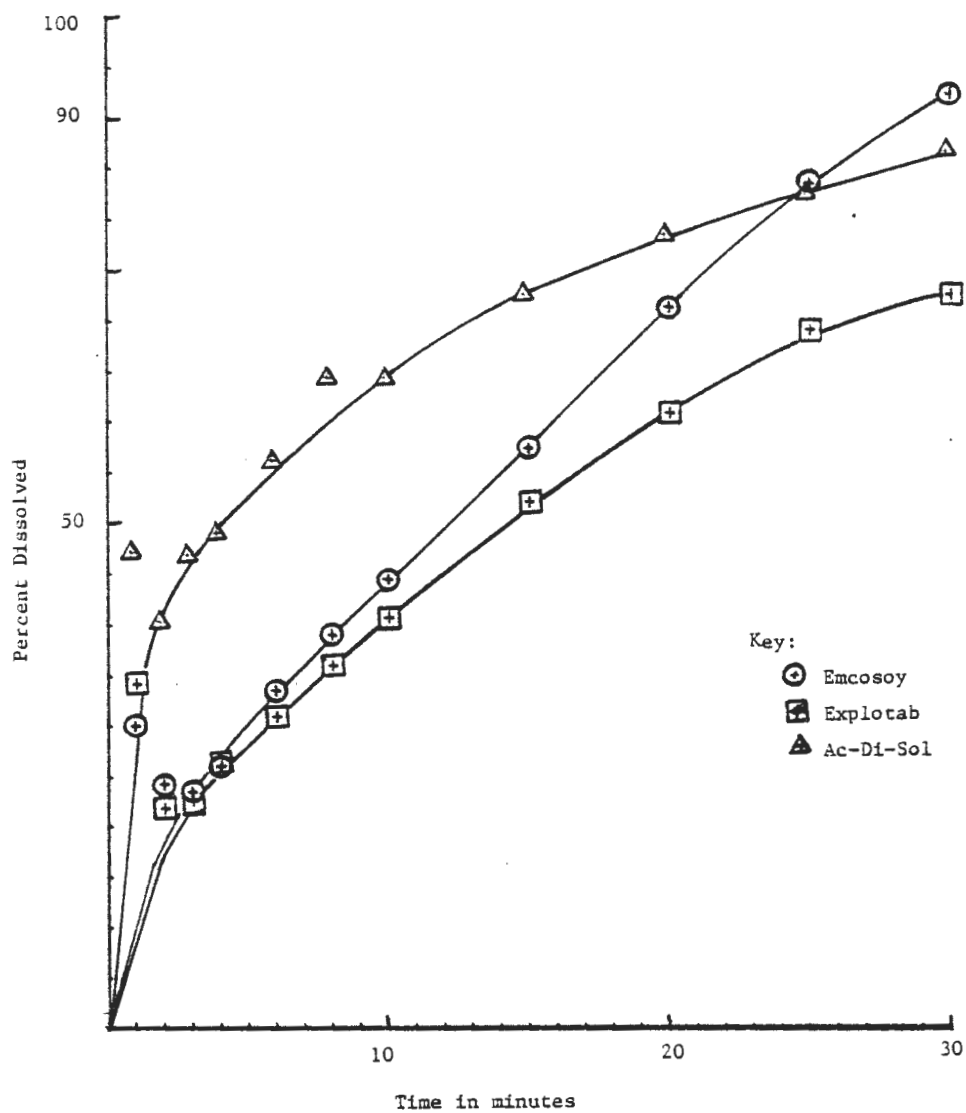
Fig. 40

TABLE XXXV

Formula II: Pyridoxine Hcl 5%  
 Disintegrant 0.5% or 1.0% or 2.0%  
 Mag. Stearate 0.5%  
 Emcompress ad 100%

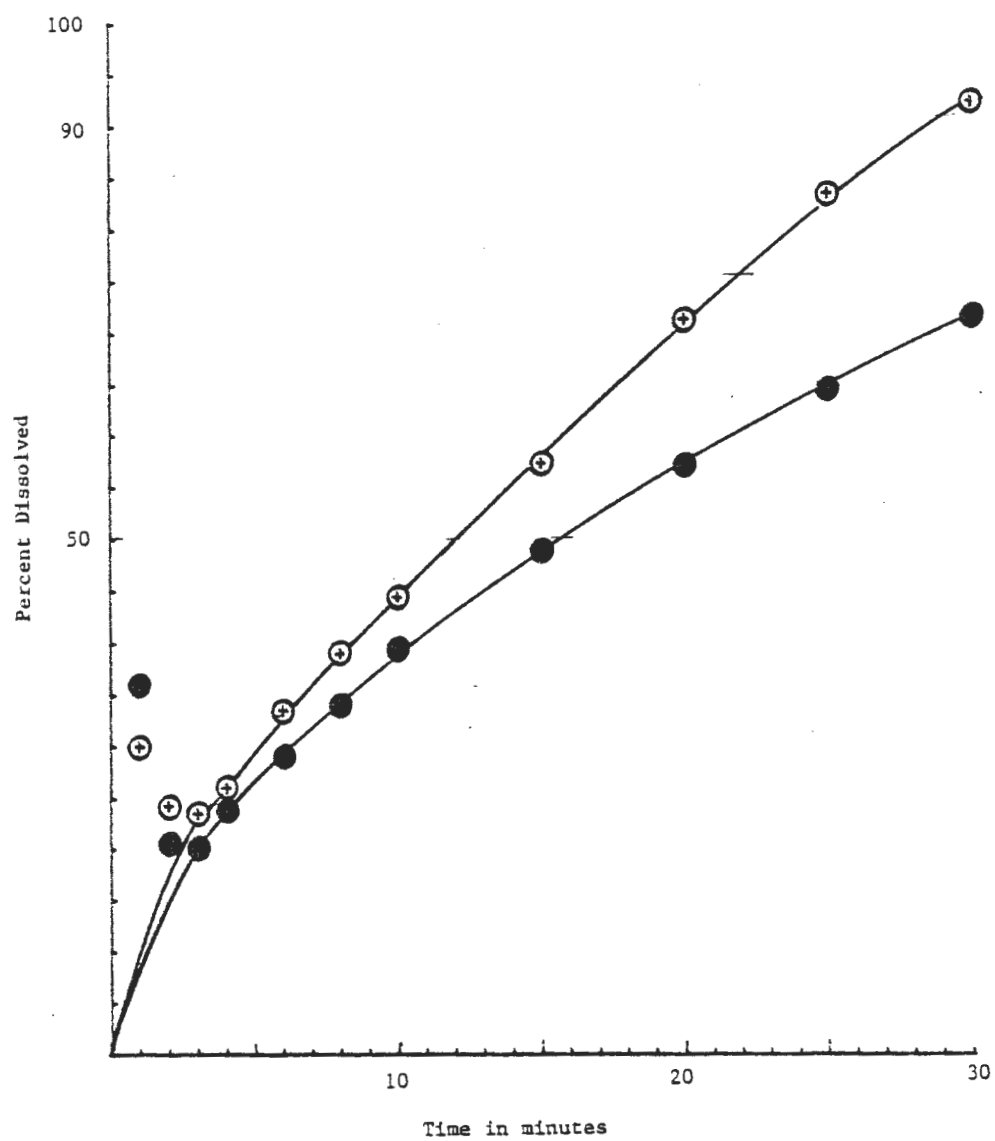
Formulation	$t_{50}$ (min)	$t_{75}$ (min)	$t_{90}$ (min)
<u>0.5% Disintegrant</u>			
Emcosoy	12.2	22.2	29.1
	16.0	> 30.0	> 30.0
Explotab	14.1	> 30.0	> 30.0
	19.9	> 30.0	> 30.0
Ac-Di-Sol	2.7	9.5	18.4
	3.8	15.8	> 30.0
<u>1.0 % Disintegrant</u>			
Emcosoy	1.8	3.8	6.5
	4.8	10.5	18.2
Explotab	3.7	14.0	approx 30.0
	6.2	15.0	approx 30.0
Ac-Di-Sol	2.0	5.1	13.1
	2.9	6.8	15.2
<u>2.0% Disintegrant</u>			
Explotab	0.9	3.8	9.5
	1.6	4.3	9.5
Ac-Di-Sol	0.8	1.2	2.0
	0.8	1.2	- 3.0
Emcosoy	0.5	0.8	1.0
	0.5	1.0	1.5
Corn starch	16.0	27.0	> 30.0
	25.0	> 30.0	> 30.0

Formulation : Pyridoxine HCl 5.0%  
 Disintegrant 0.5%  
 Mag. Stearate 0.5%  
 Emcompress ad 100%



Dissolution profiles for pyridoxine HCl tablet formulations containing 0.5% of different disintegrants

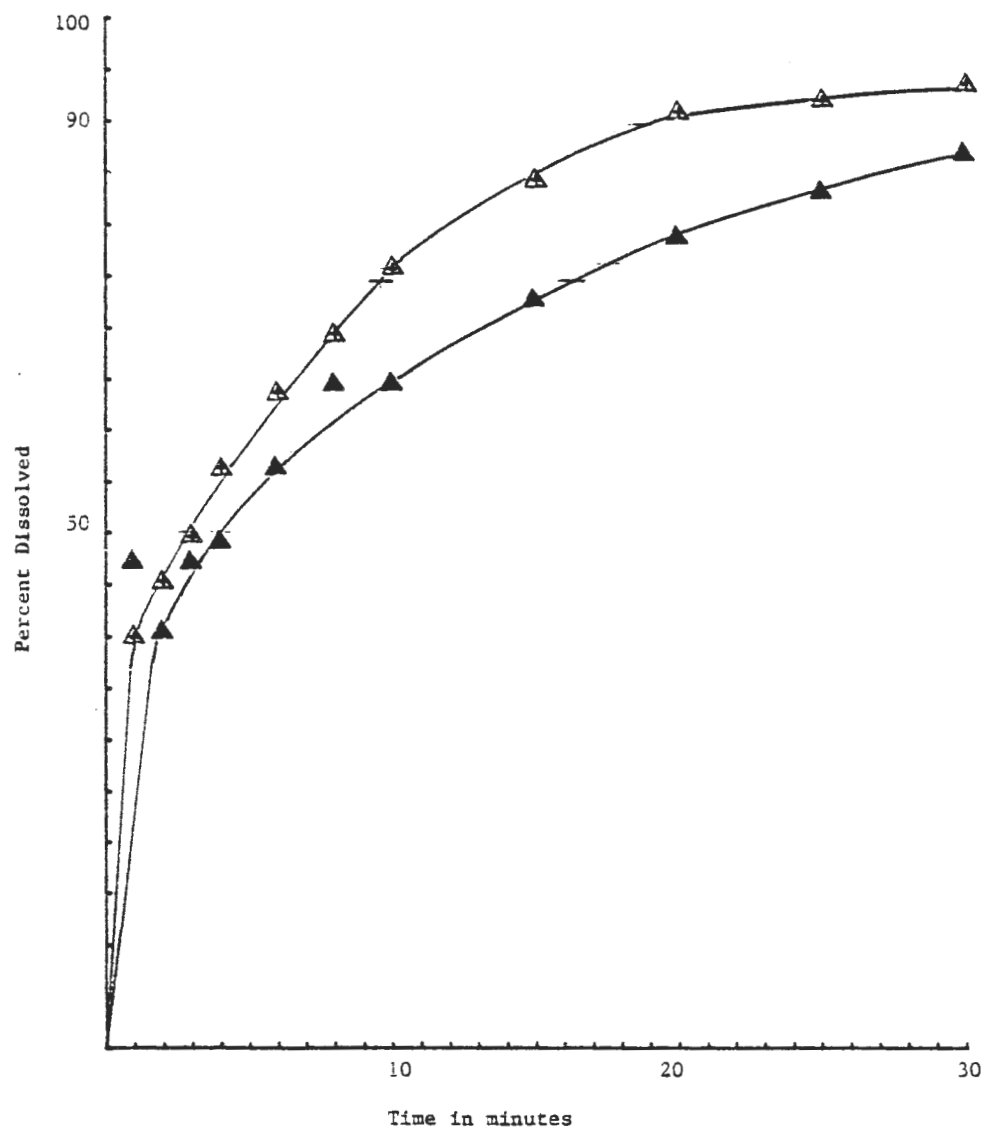
Fig. 41



Dissolution profiles for Pyridoxine HCl formulations containing 0.5% Emcosoy  
Replicate 1 (⊕), and Replicate 2 (●) Froccompress as matrix

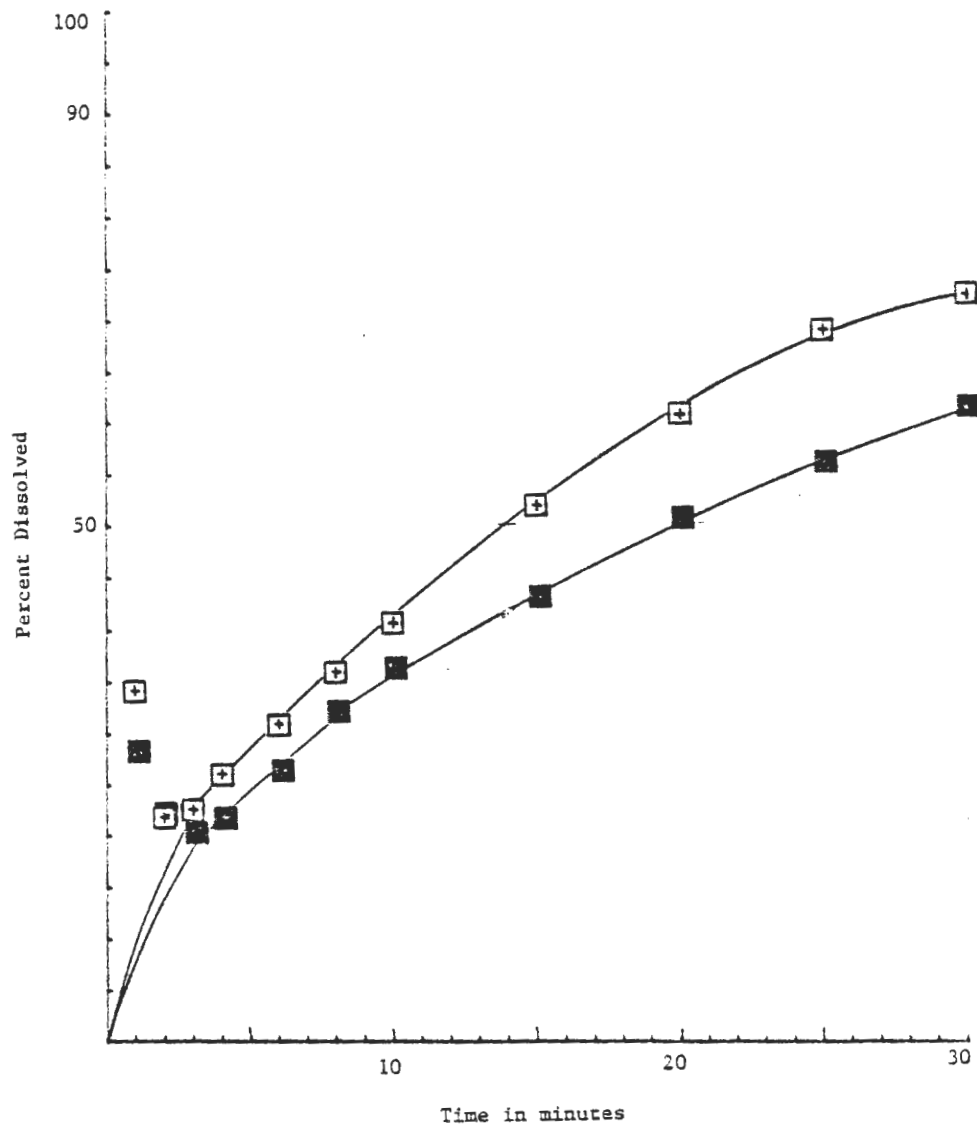
Fig. 42





Dissolution profiles for pyridoxine HCl tablets containing 0.5% Ac-Di-Sol  
Replicate 1  $\triangle$  , and Replicate 2  $\blacktriangle$  Emcompress as a matrix.

Fig. 43



Dissolution profiles for pyridoxine HCl tablets containing 0.5% Explotab  
Replicate 1  $\square$ , and Replicate 2  $\blacksquare$  Emcompress as matrix.

Fig. 44

Formulation: Pyridoxine HCl 5%  
 Disintegrant 1.0%  
 Mag. Stearate. 0.5%  
 Emccompress ad 100%  
 Dissolution profiles.

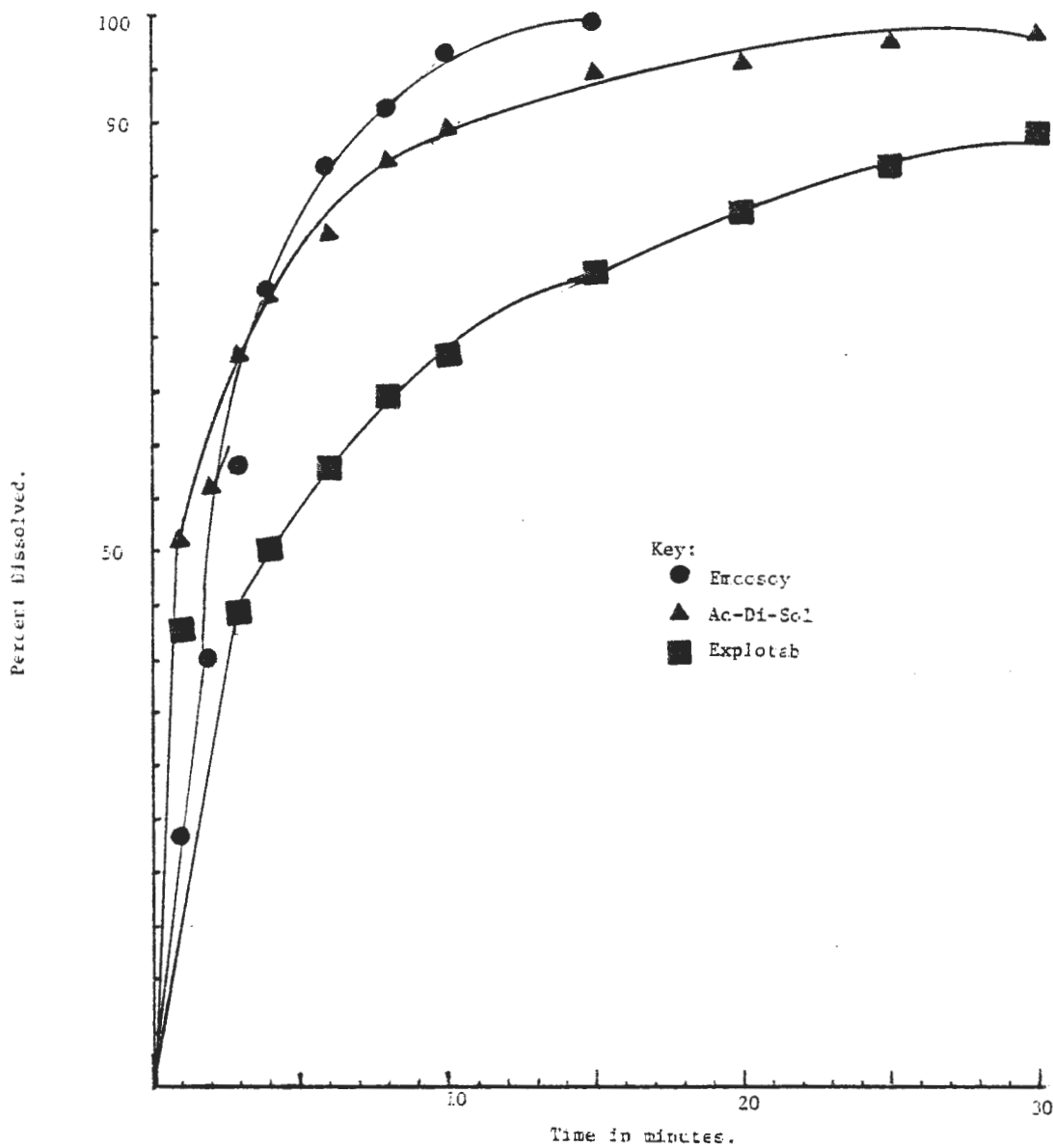
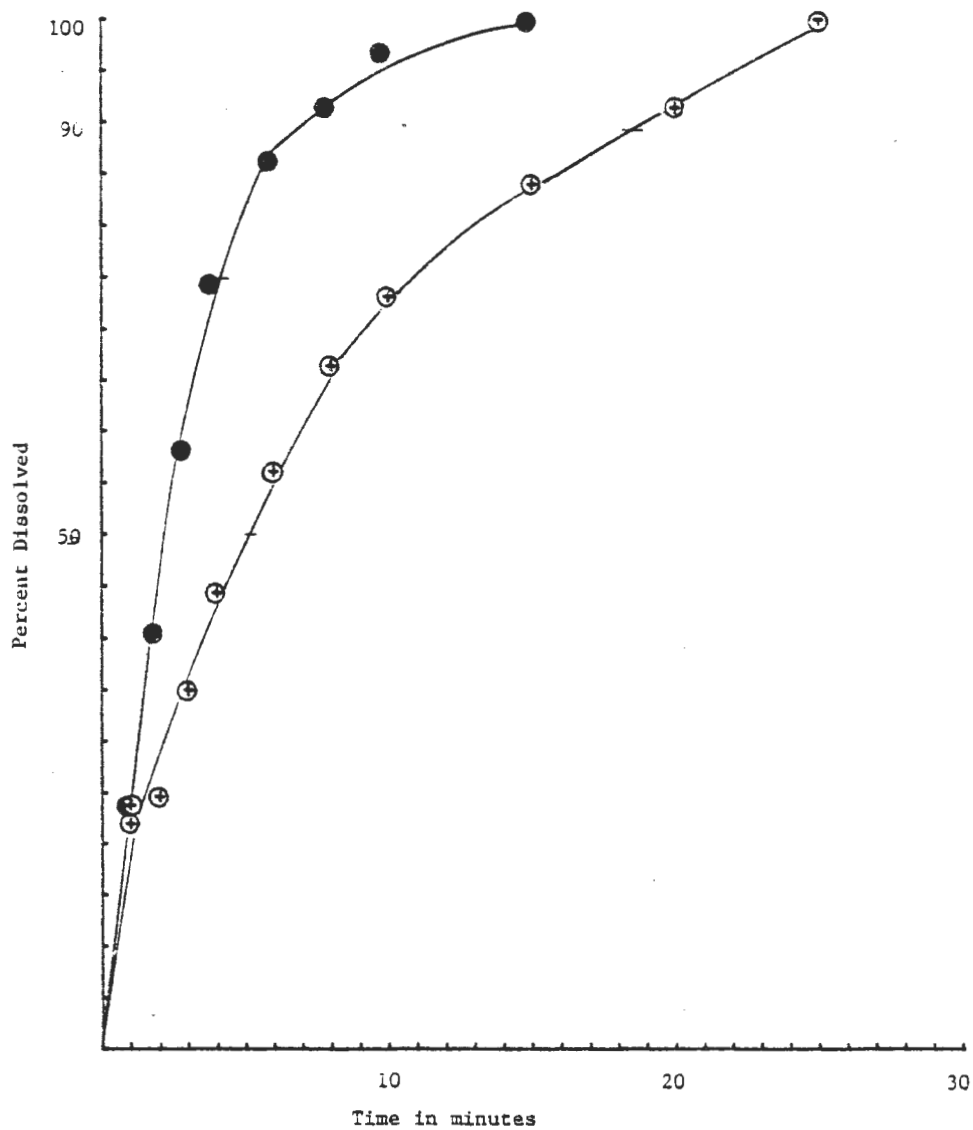
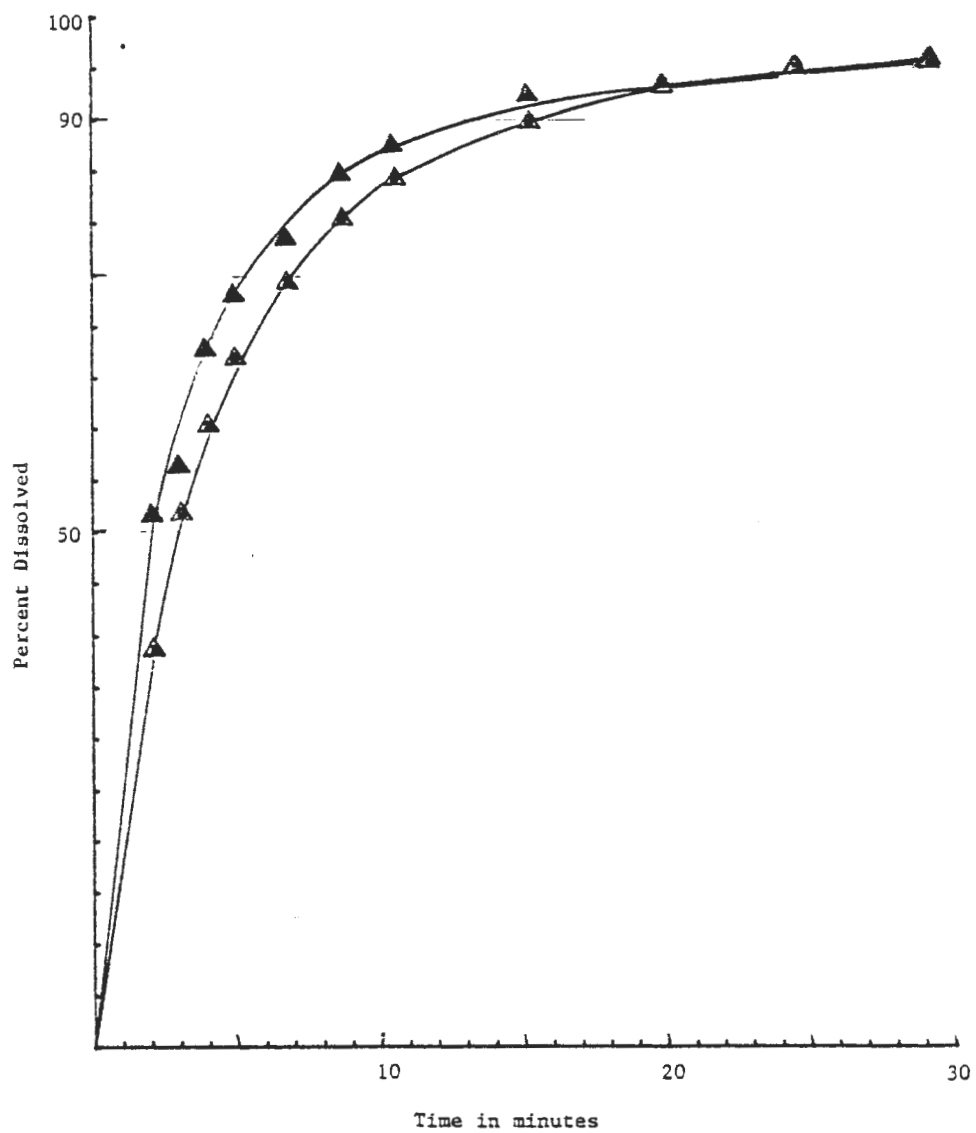


Fig. 45



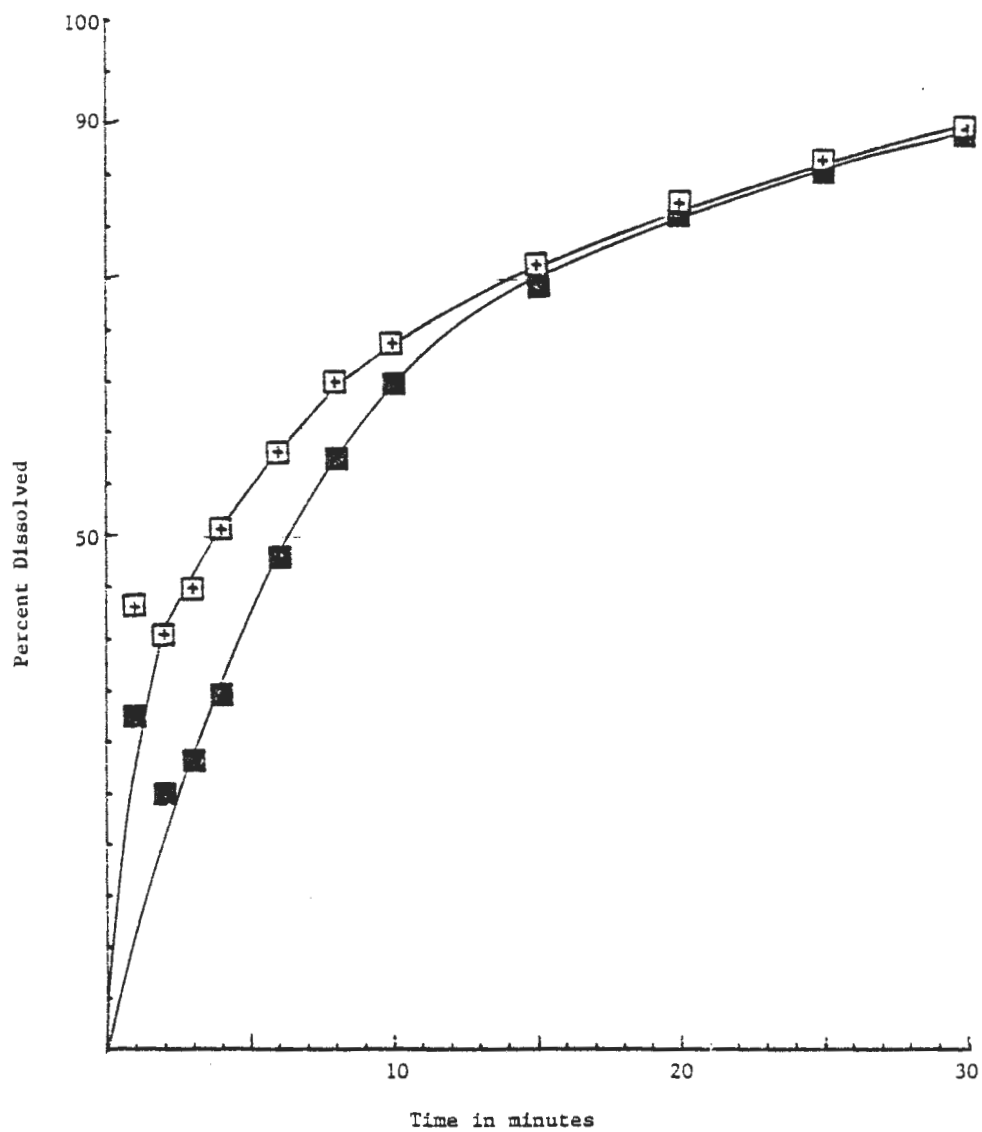
Dissolution profiles for pyridoxine HCl tablets containing 1% Emcosoy Replicate 1 (⊕), and Replicate 2 (●) Emcompress as a matrix.

Fig. 46



Dissolution profiles for pyridoxine HCl tablets containing 12 Ac-Di-Sol  
Replicate 1  $\Delta$  , and Replicate 2  $\blacktriangle$  Emcompress as a matrix.

Fig. 47



Dissolution profiles for pyridoxine HCl tablets containing 1% Explotab  
Replicate 1  $\square$  , and Replicate 2  $\blacksquare$  Emocompress as a matrix.

Fig. 48

Dissolution profiles.

Formulation: Pyridoxine Hcl 5%  
Disintegrant 2.0%  
Mag. Stearate. 0.5%  
Emcompress ad 100%

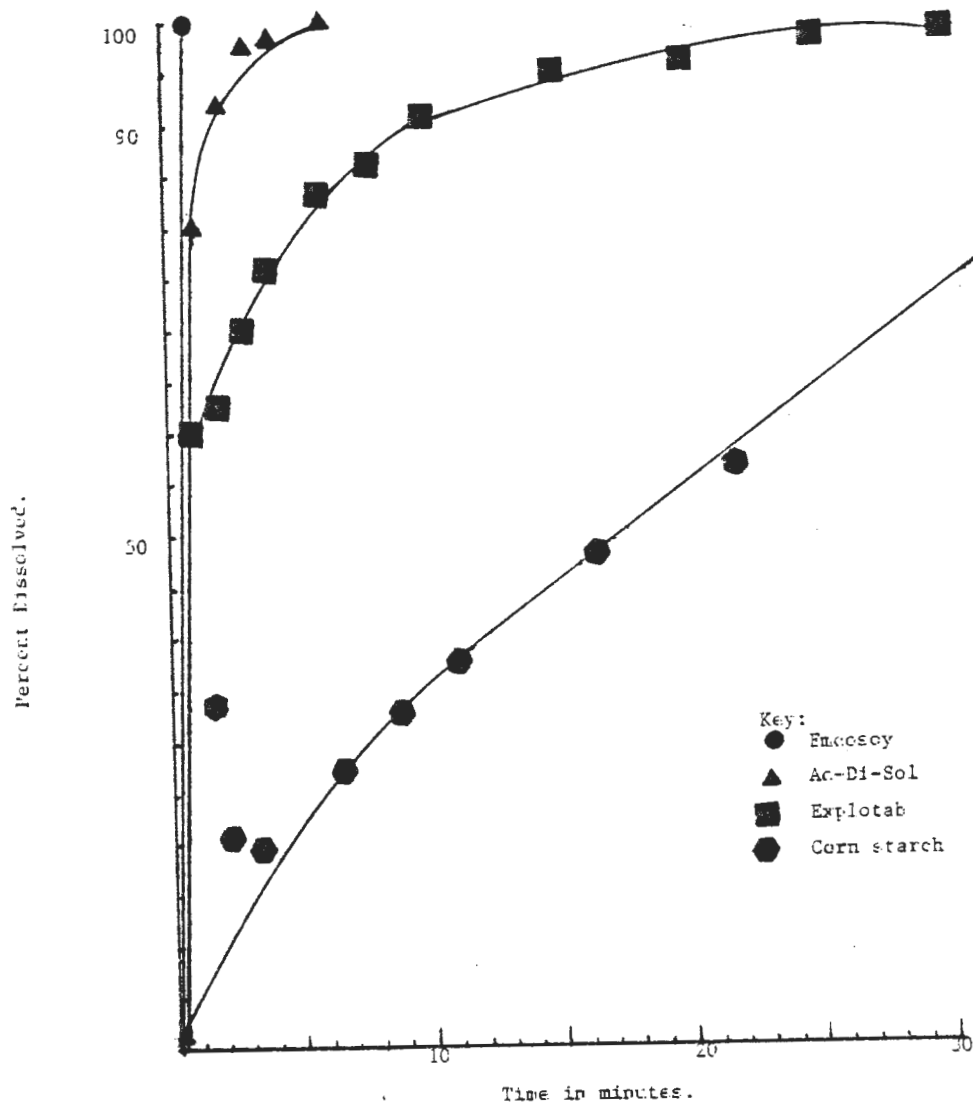
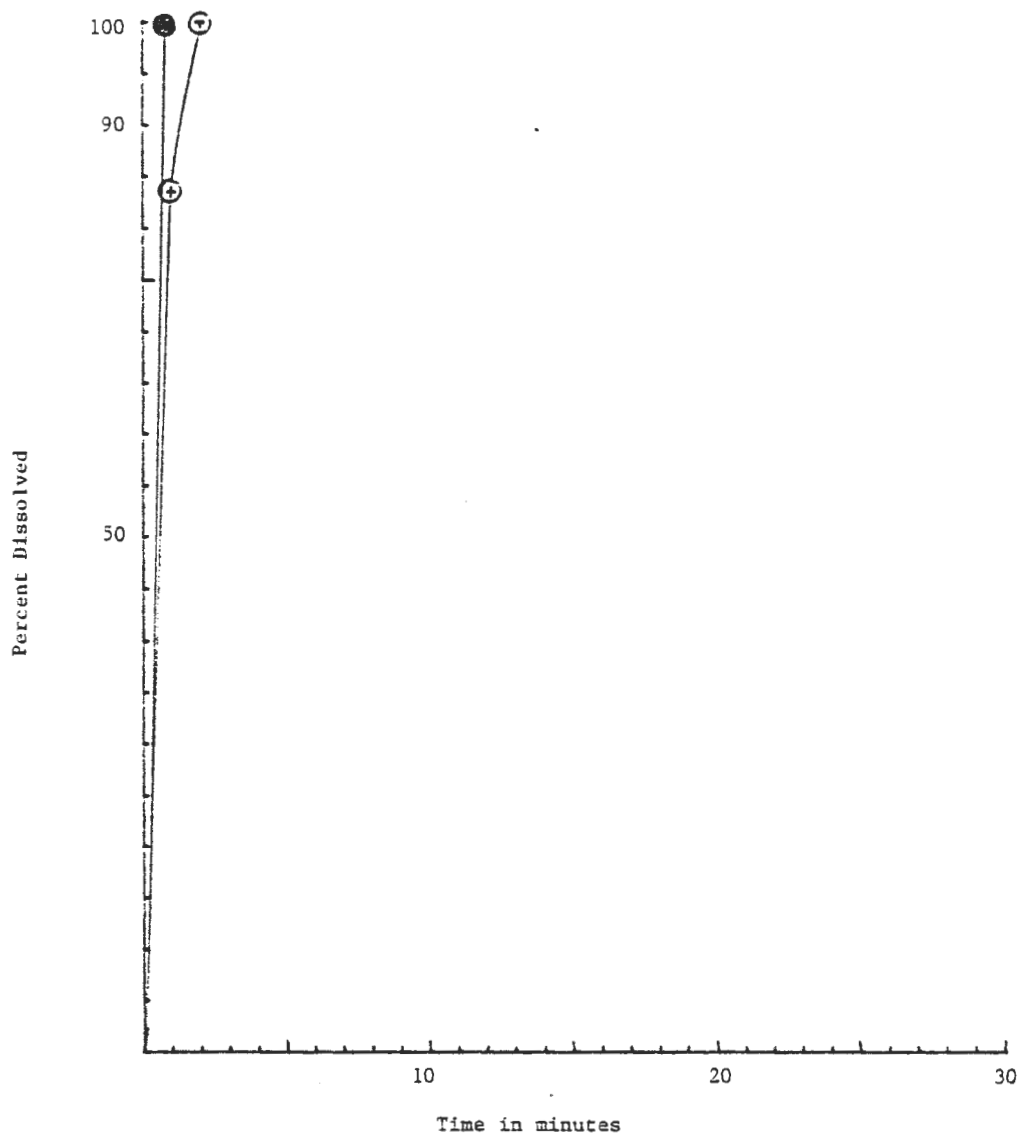


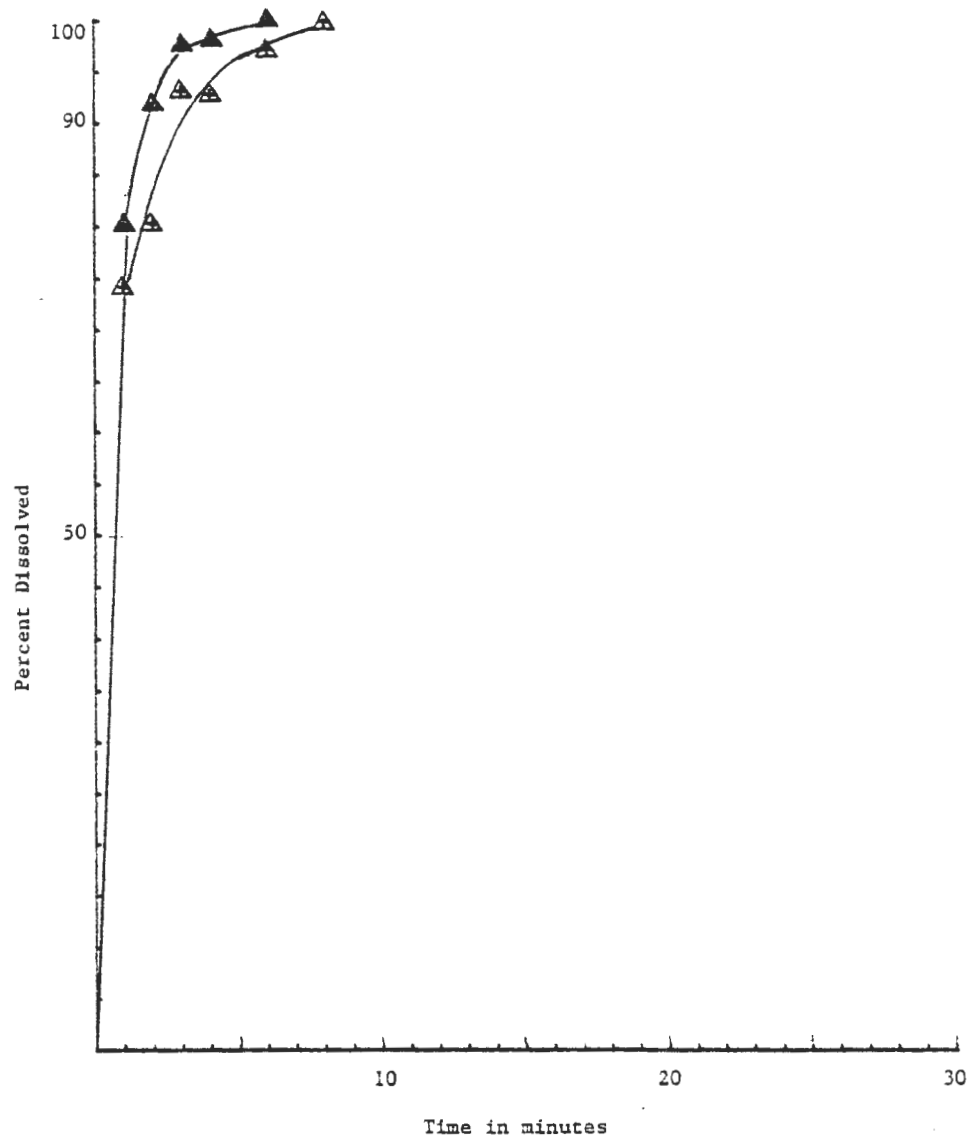
Fig. 49



Dissolution profiles for pyridoxine HCl tablets containing 2% Emcosoy  
Replicate 1 ⊕, and Replicate 2 ● Emocupress as a matrix.

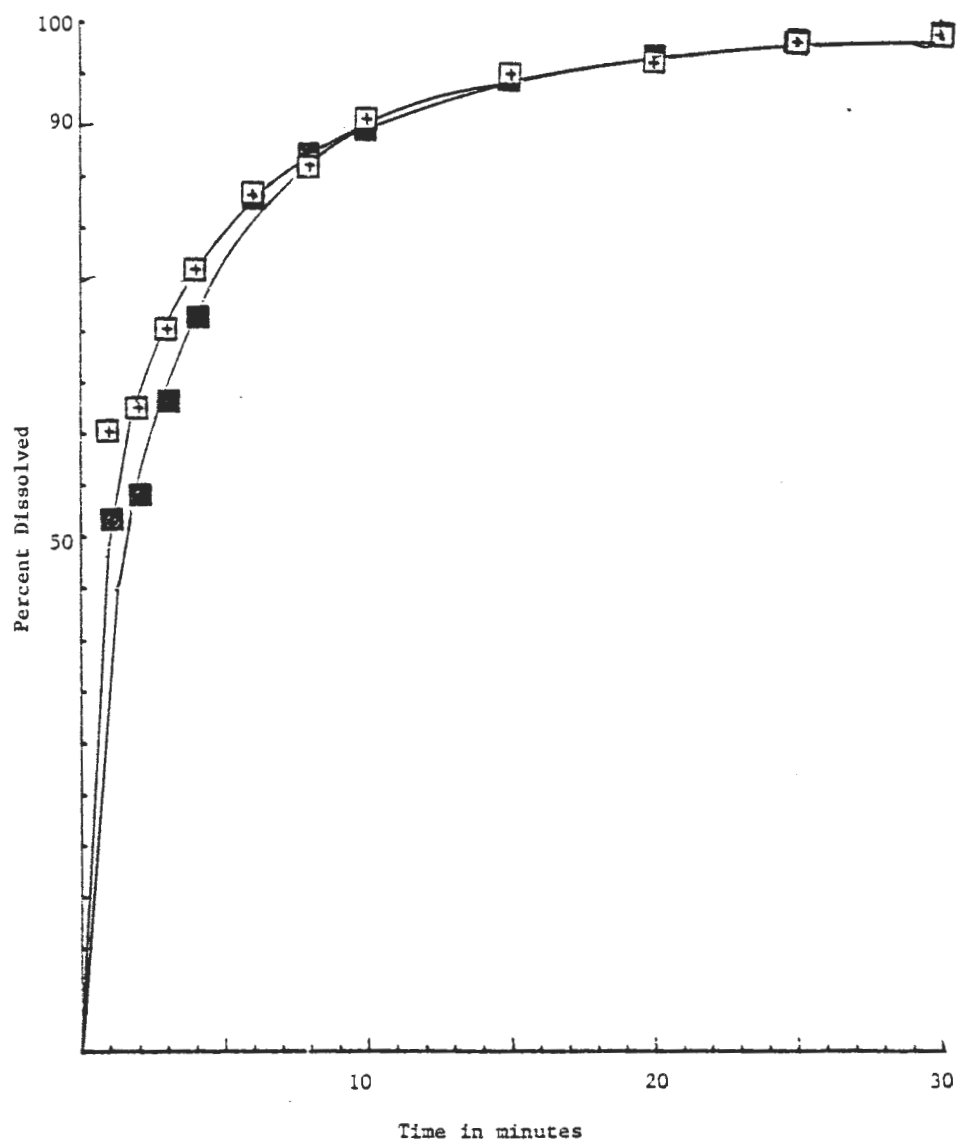
Fig. 50





Dissolution profiles for pyridoxine HCl tablets containing 2% Ac-Di-Sol  
Replicate 1  $\triangle$  , and Replicate 2  $\blacktriangle$  Emcompress as a matrix.

Fig. 51



Dissolution profiles for pyridoxine HCl tablets containing 2% Explotab  
Replicate 1 (□), and Replicate 2 (■) Emcompress as a matrix.

Fig. 52

TABLE XXXVI

Formulation: Pyridoxine Hcl 5%  
 Disintegrant 0.5% or 1.0% or 3.0%  
 Mag. Stearate 0.5%  
 Emdex ad 100%

DISSOLUTION DATA:  
 -----

Formulation	$t_{50}(\text{min})$	$t_{75}(\text{min})$	$t_{90}(\text{min})$
<u>0.5% Disintegrant</u>			
Emcosoy	4.35	9.60	15.10
	4.42	9.75	17.45
Ac-Di-Sol	3.40	9.20	14.70
	4.05	9.70	16.30
Explotab	4.20	11.60	21.20
	5.30	15.60	26.80
<u>1.0% Disintegrant</u>			
Emcosoy	5.10	10.40	15.70
	5.15	12.30	19.15
Ac-Di-Sol	4.60	10.50	16.90
	6.70	16.85	30.00
Explotab	2.35	8.10	14.30
	4.60	13.15	16.60
<u>3.0% Disintegrant</u>			
Emcosoy	3.20	7.40	12.65
Corn starch	6.50	17.85	27.25

## Dissolution profiles.

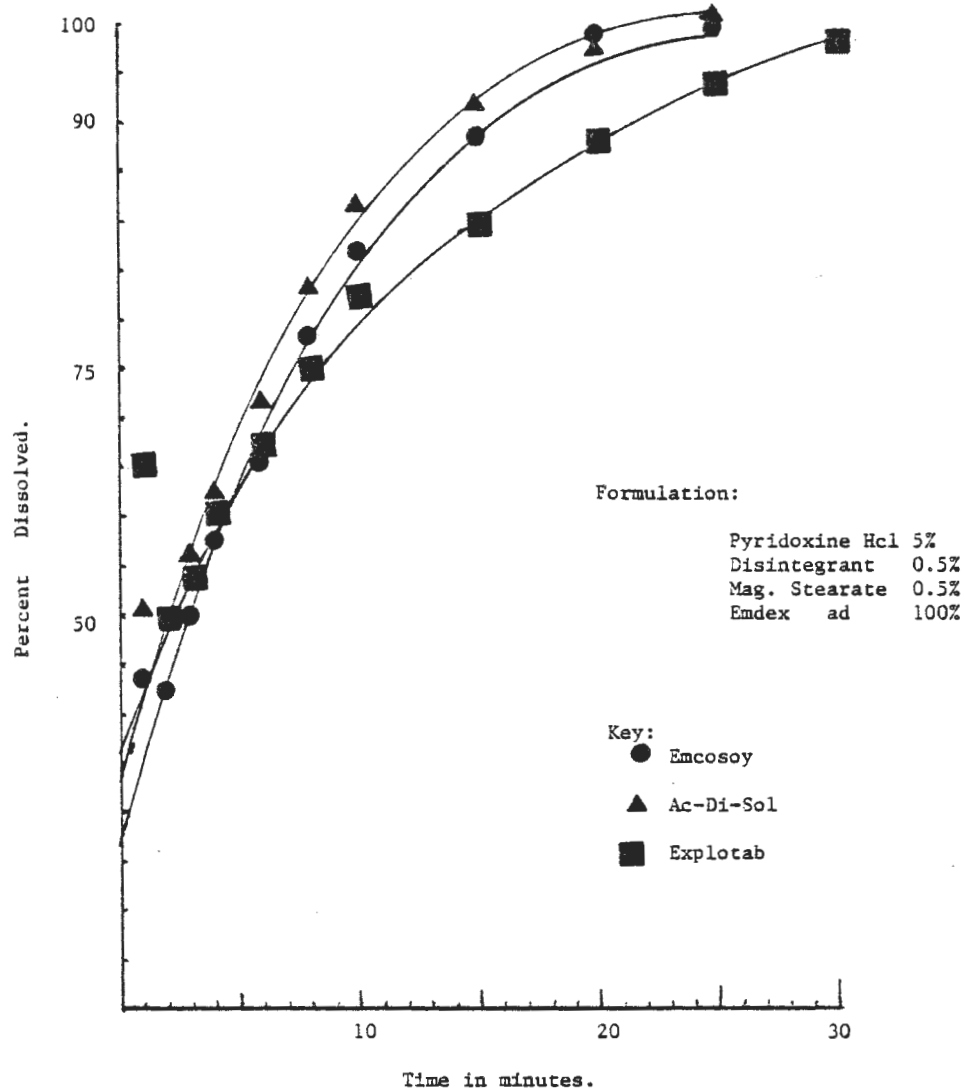
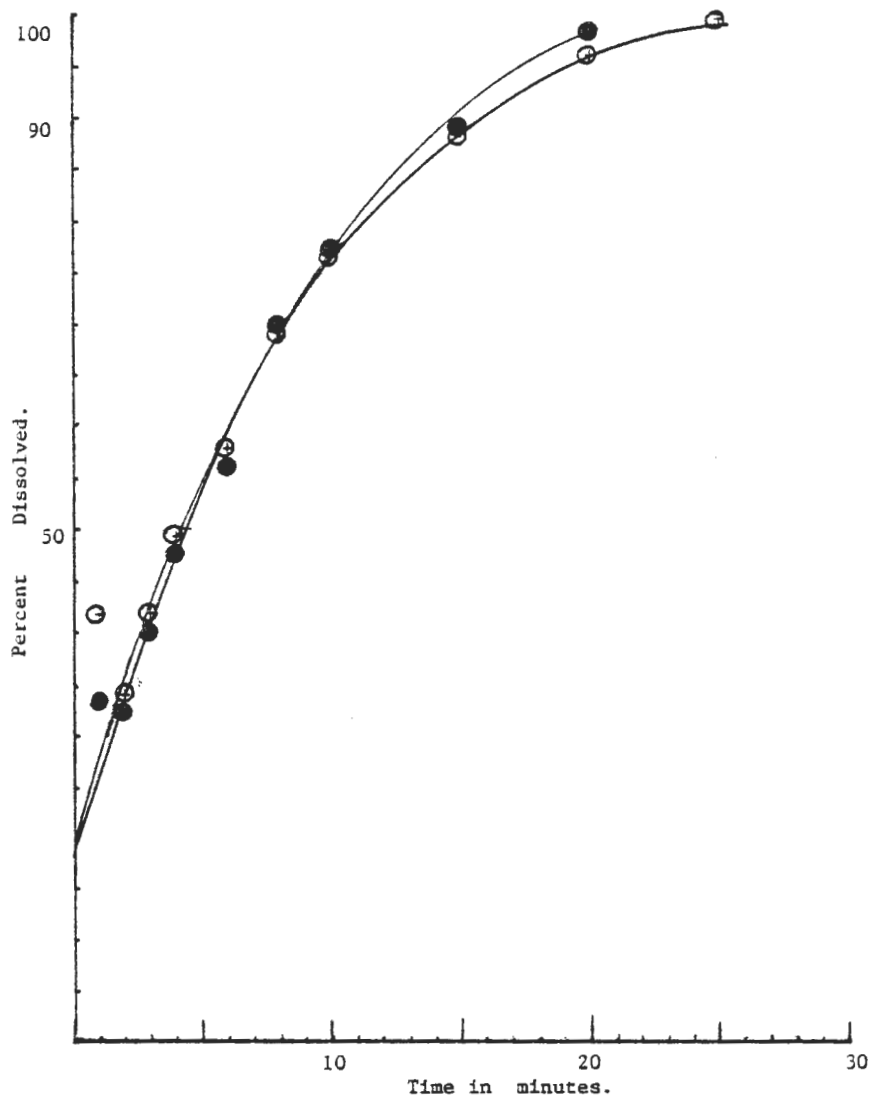


Fig. 53



Dissolution profile of pyridoxine formulation containing 0.5% of Emcosoy  
Replicate 1 ● , Replicate 2 ○ Emdex as a matrix.

Fig. 54

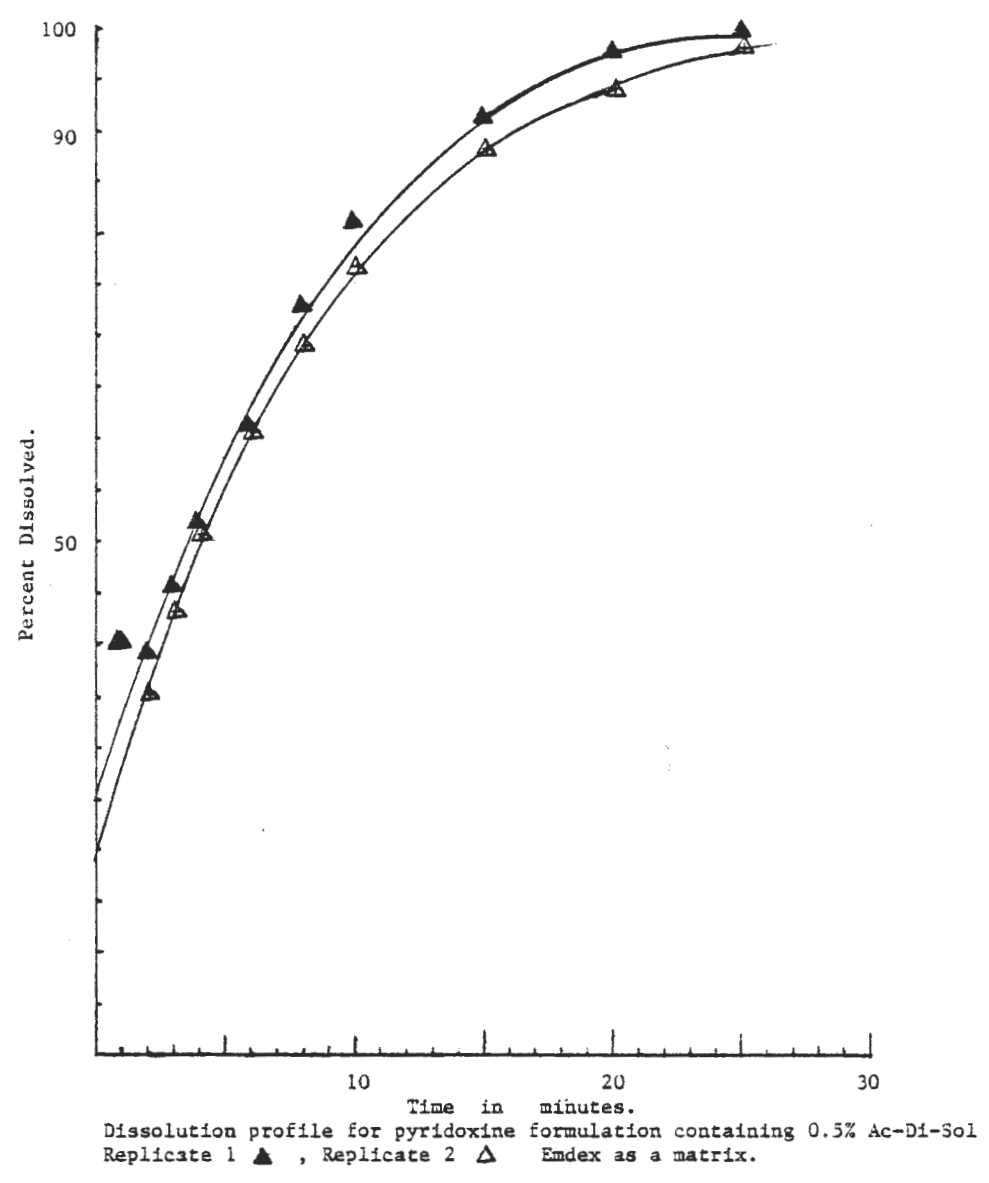
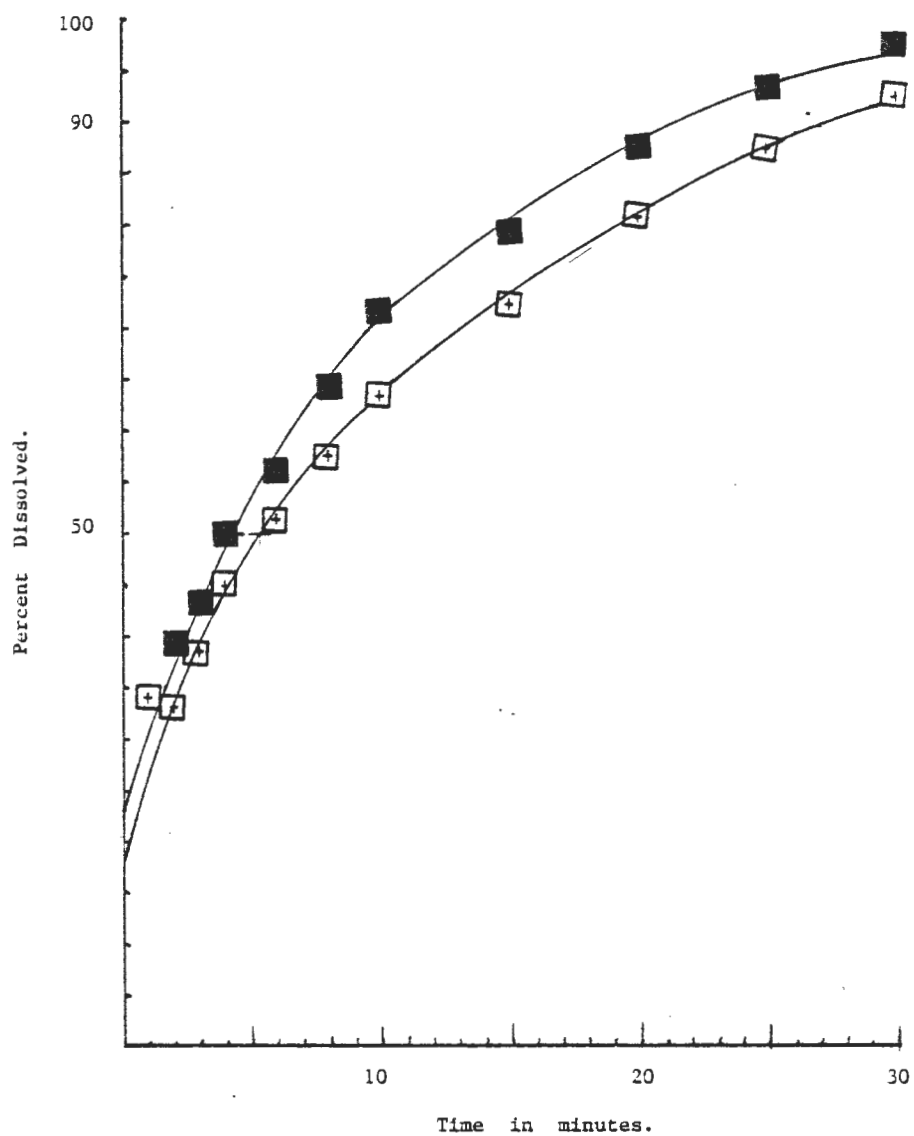


Fig. 55



Dissolution profile of pyridoxine formulation containing 0.5% of Explotab  
Replicate 1 ■ , Replicate 2 □+ Endex as a matrix.

Fig. 56

## Dissolution Profiles.

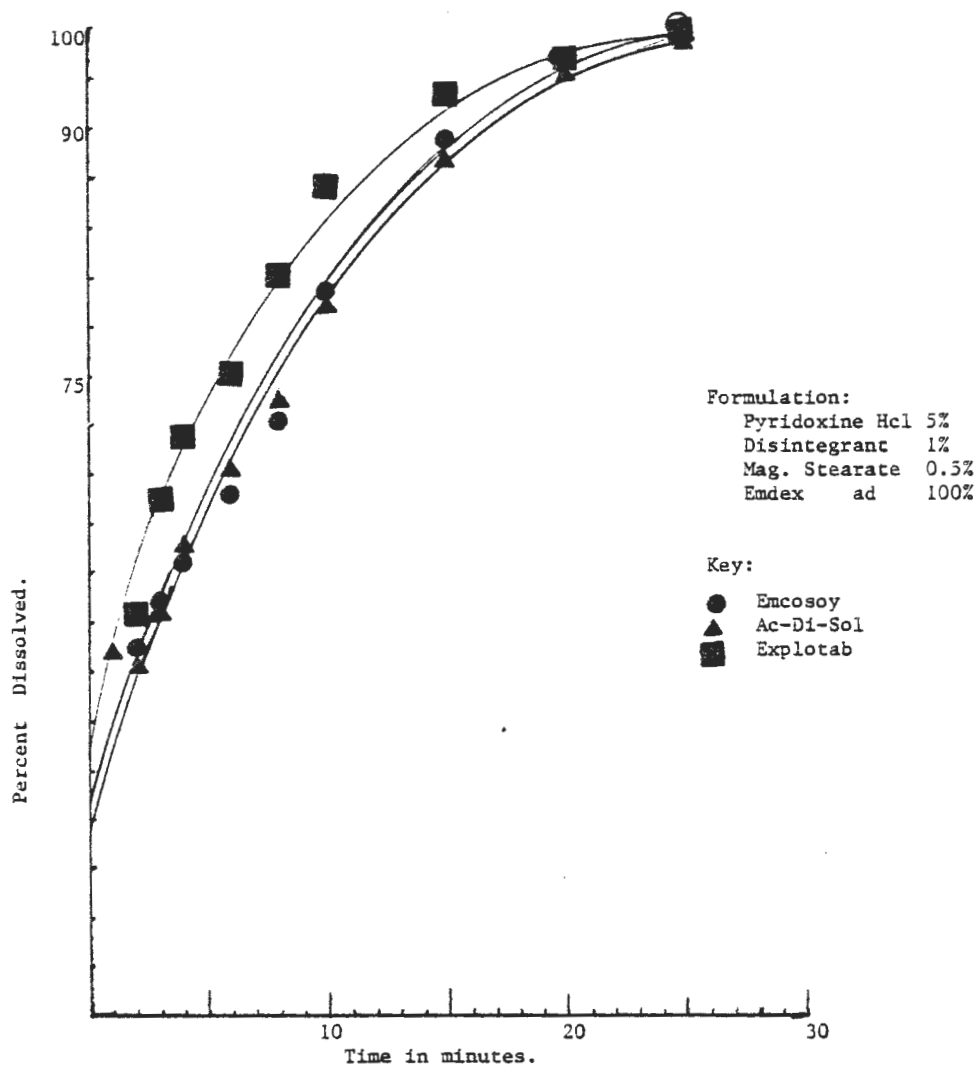


Fig. 57



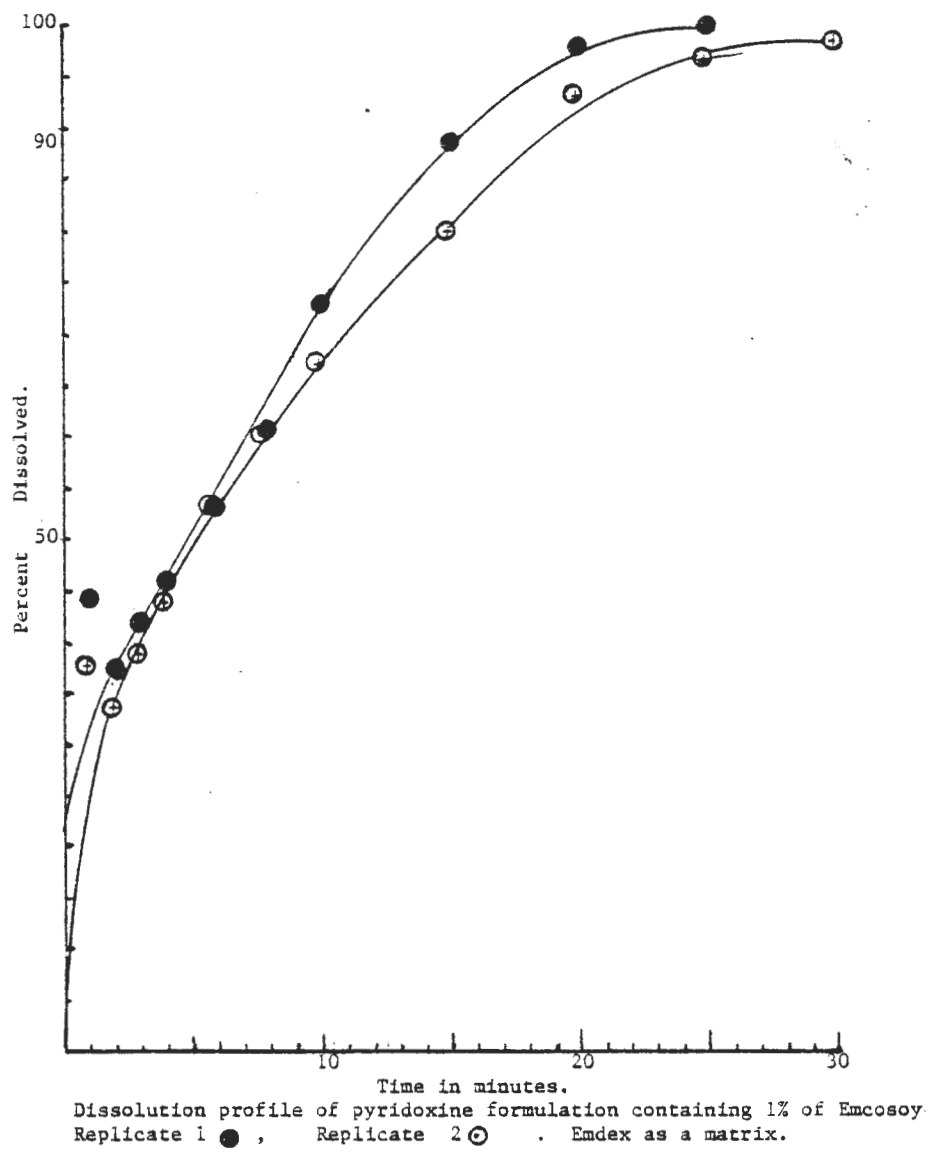
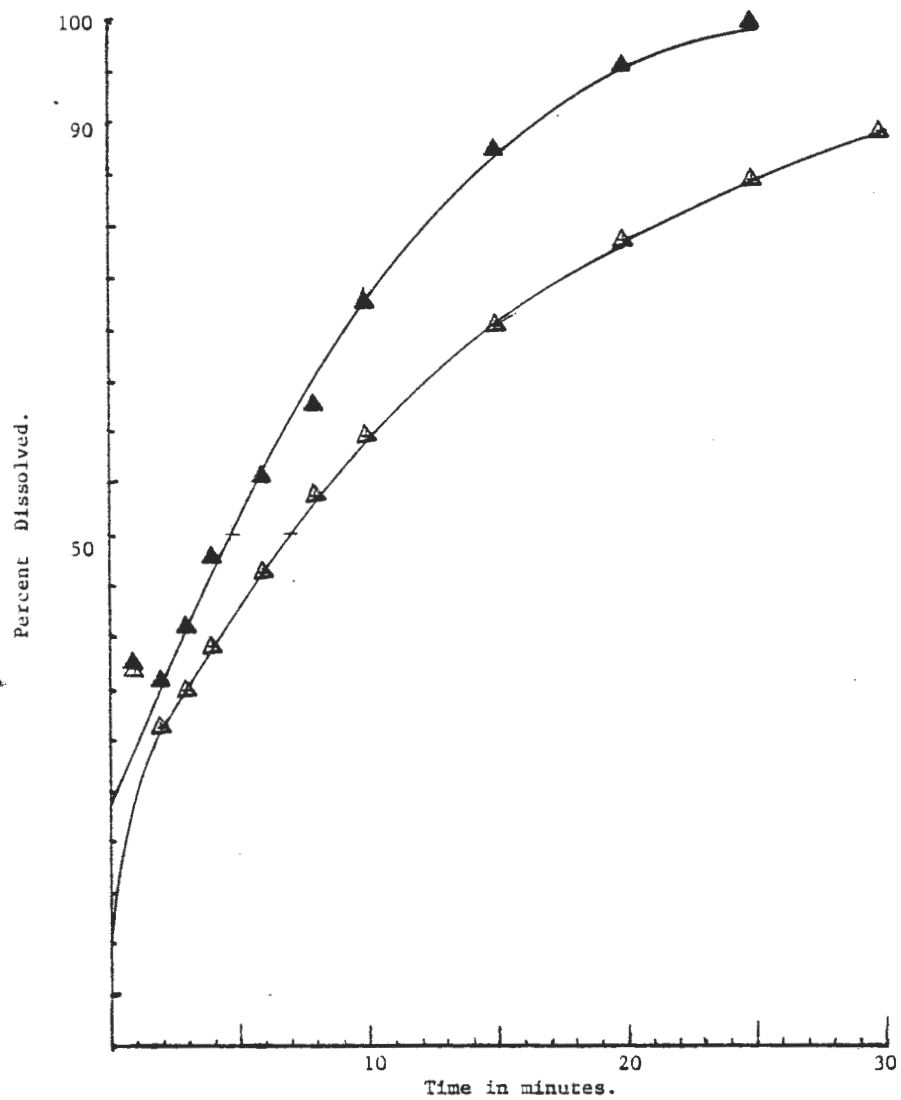
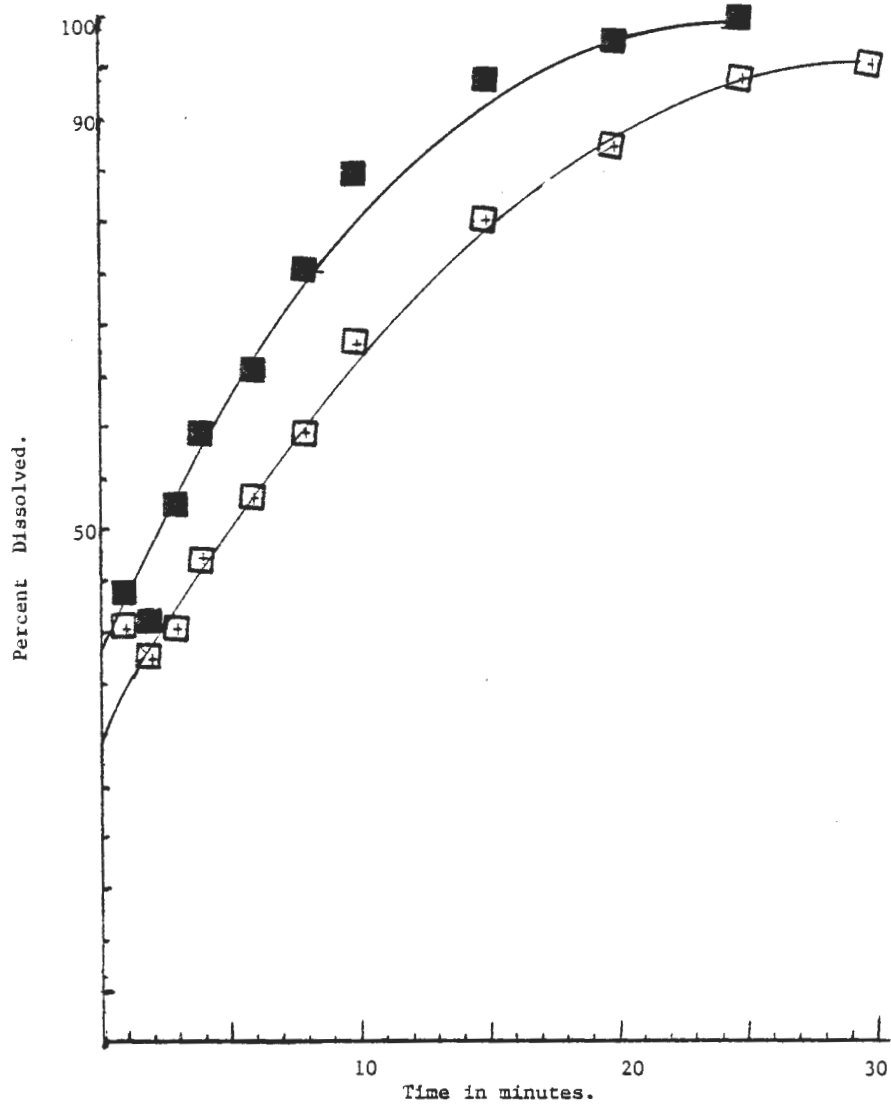


Fig. 58



Dissolution profile of pyridoxine formulation containing 1% of Ac-Di-Sol Replicate 1 ▲ , Replicate 2 △ Emdex as a matrix.

Fig. 59



Dissolution profile for pyridoxine formulation containing 1% of Explotab  
Replicate 1 ■ , Replicate 2 □ Emdex as a matrix.

Fig. 60

## Dissolution profiles.

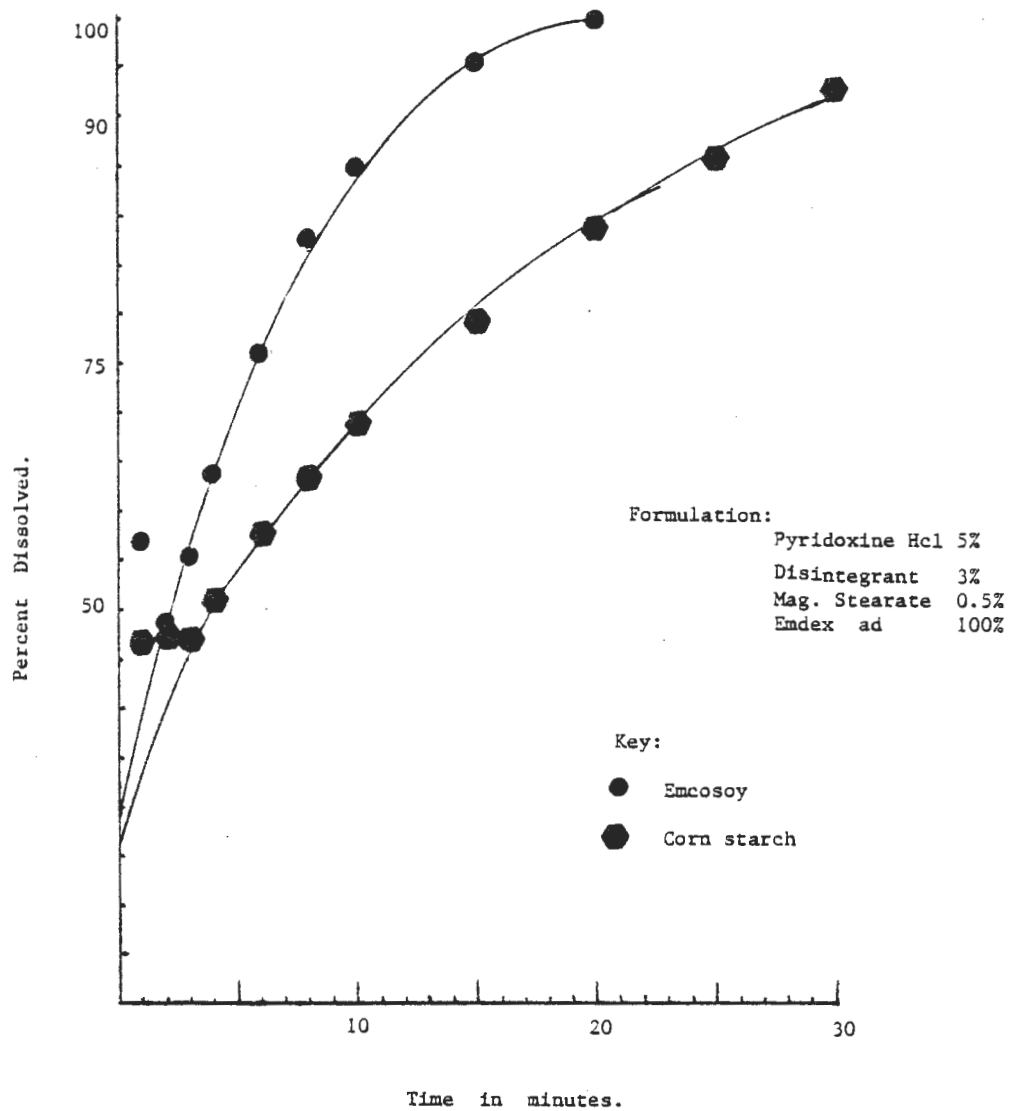
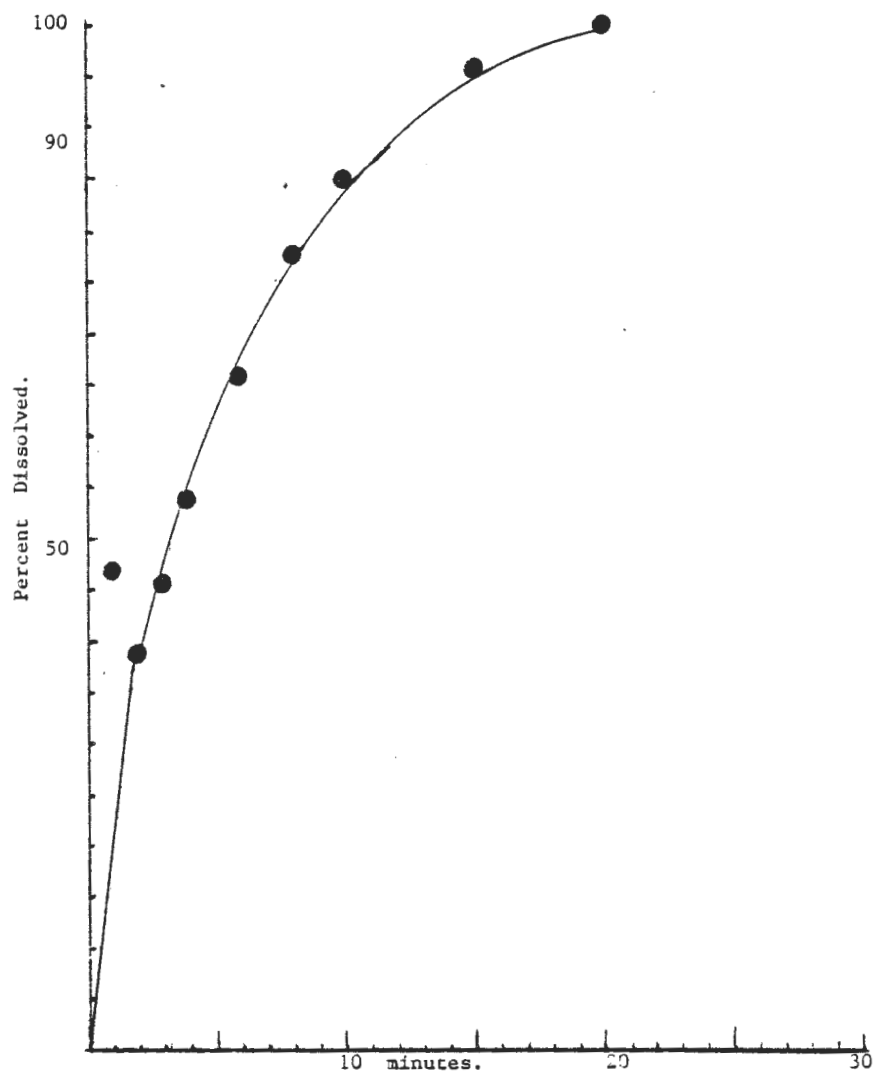
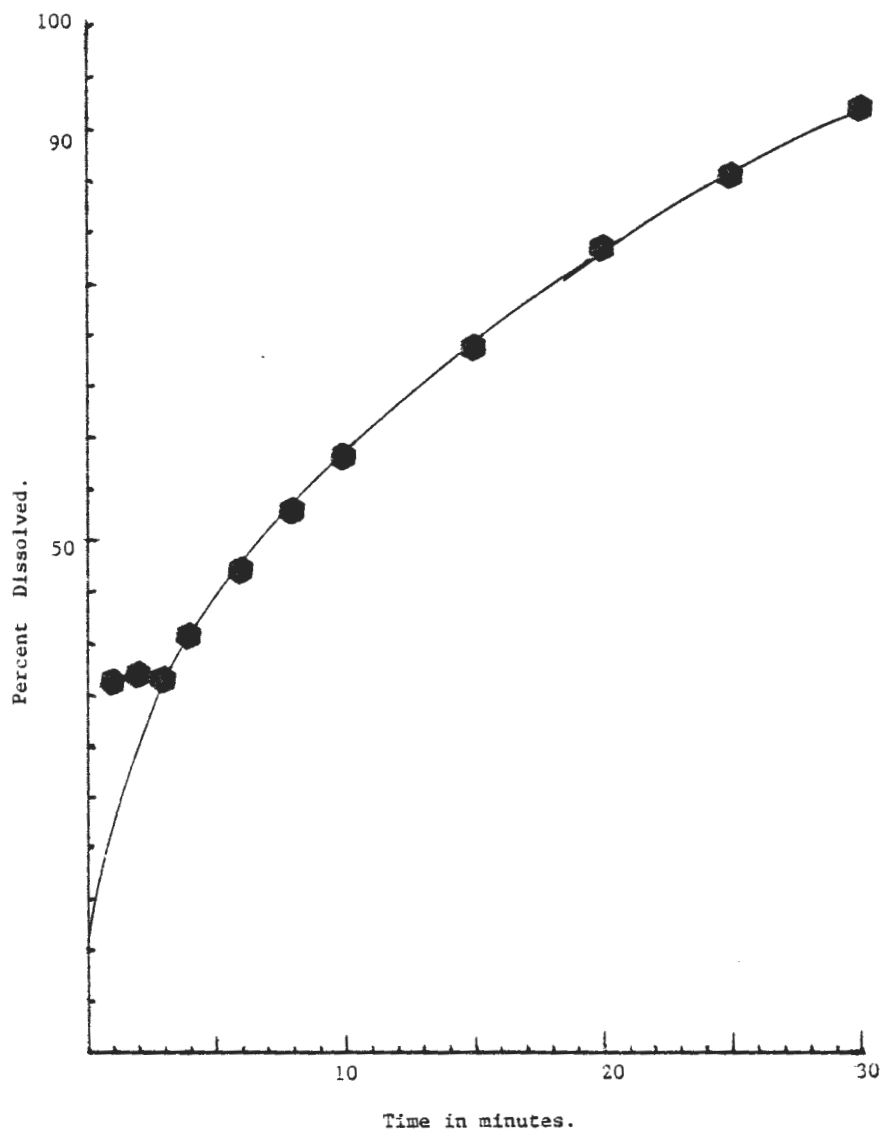


Fig. 61



Dissolution profile for pyridoxine formulation containing 3% of Emcosoy Emdex as a matrix.

Fig. 62



Dissolution profile for Pyridoxine formulation containing 3% of Corn starch.  
Emdex as a matrix.

Fig. 63

TABLE XXXVII

Formulation: Ascorbic acid 25%  
 ----- Disintegrant 0.5%, 1%, 2% or 3%  
 stearic acid 2.5%  
 Emcompress ad 100%

DISSOLUTION DATA :  
 -----

Formulation	t <sub>50</sub> (min)	t <sub>75</sub> (min)	t <sub>90</sub> (min)
<u>0.5% Disintegrant</u>			
Emcosoy	2.20	3.20	4.05
	2:50	3.70	5.60
Ac-Di-Sol	1.20	1.60	2.40
	1.35	1.95	2.55
<u>1.0% Disintegrant</u>			
Emcosoy	0.60	0.84	1.80
	1.05	1.50	1.80
Ac-Di-Sol	0.85	1.30	2.20
	0.85	1.30	2.20
Explotab	2.25	4.80	5.75
	2.45	5.10	6.15
<u>2.0% Disintegrant *</u>			
Emcosoy	1.30	2.00	3.20
	1.20	1.40	2.00
Ac-Di-Sol	1.50	2.55	4.00
	1.80	2.40	2.80
Explotab	4.35	6.60	7.85
	6.00	8.30	9.60
Corn Starch	23.10	> 30.00	> 30.00
	30.00	> 30.00	> 30.00
<u>3.0% Disintegrant</u>			
Emcosoy	0.45	0.90	1.05
	0.80	1.40	2.00
Corn starch	17.60	> 30.00	> 30.00
	18.40	> 30.00	> 30.00

\* Granulated Ascorbic acid(90%) (Roche) was used at 2.0% disintegrant level, rest formulations contained Ascorbic acid U.S.P.

## Dissolution profiles

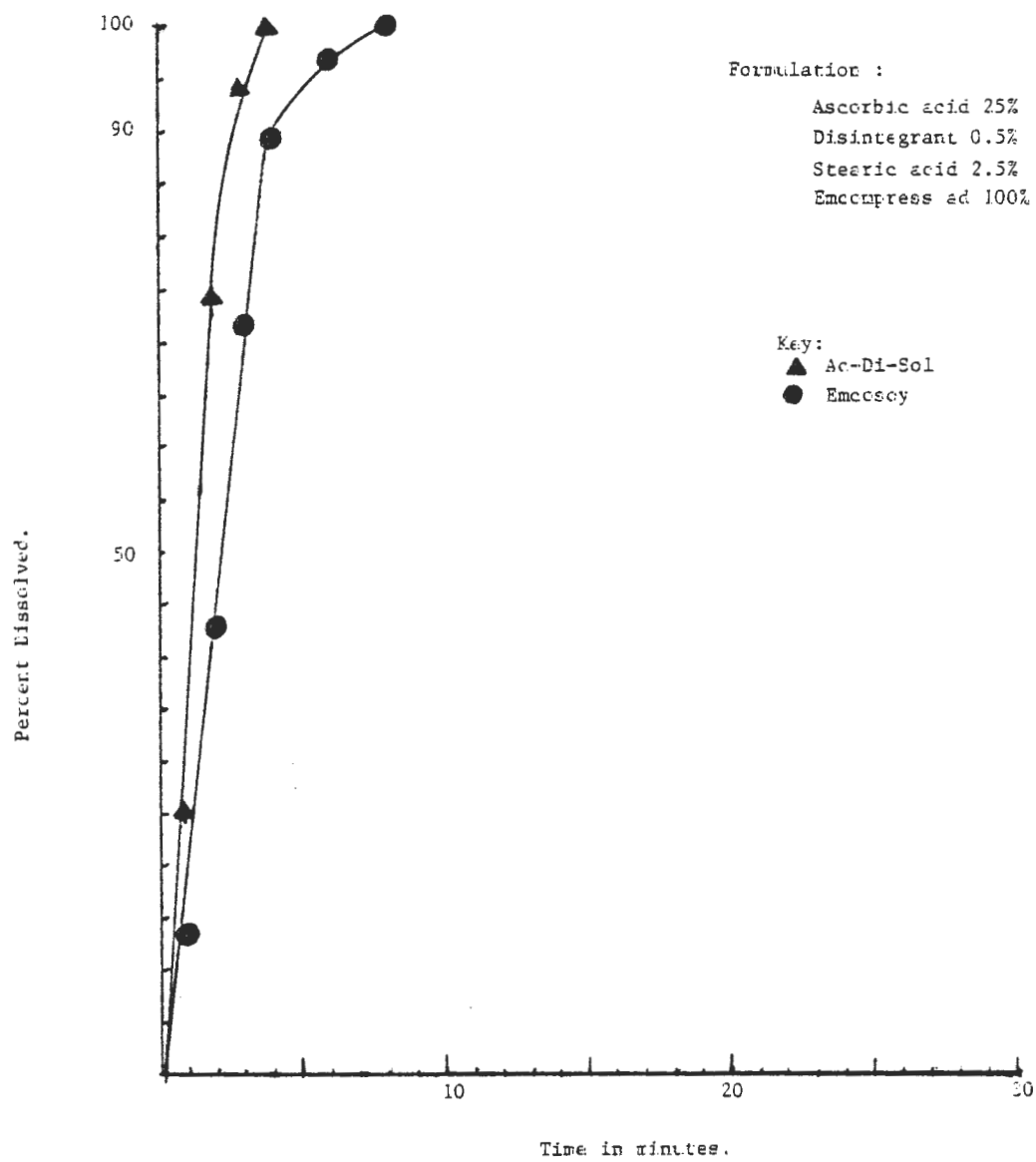
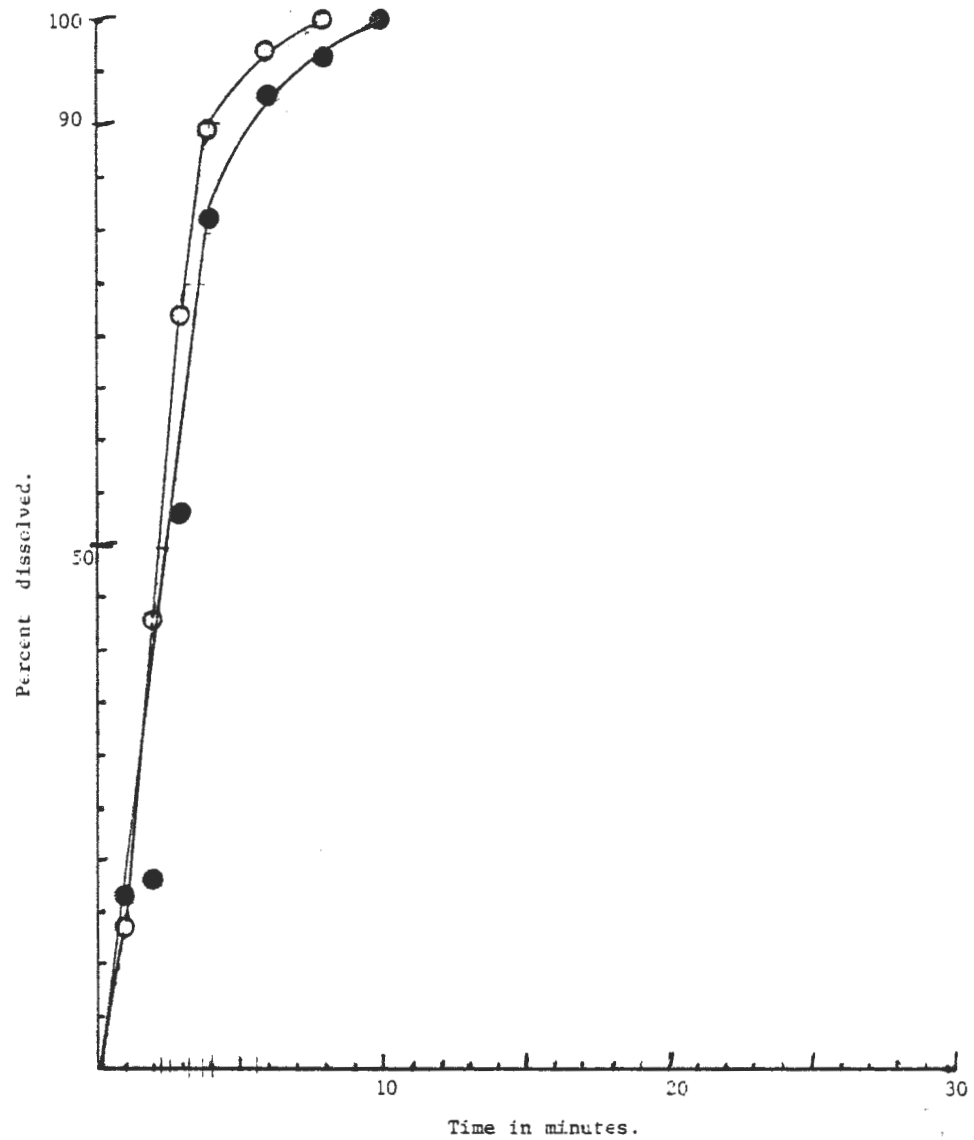


Fig. 64





Dissolution profiles for Ascorbic acid formulations containing 0.5% Emcoscy  
Replicate 1 (○), Replicate 2 (●)

Fig. 65

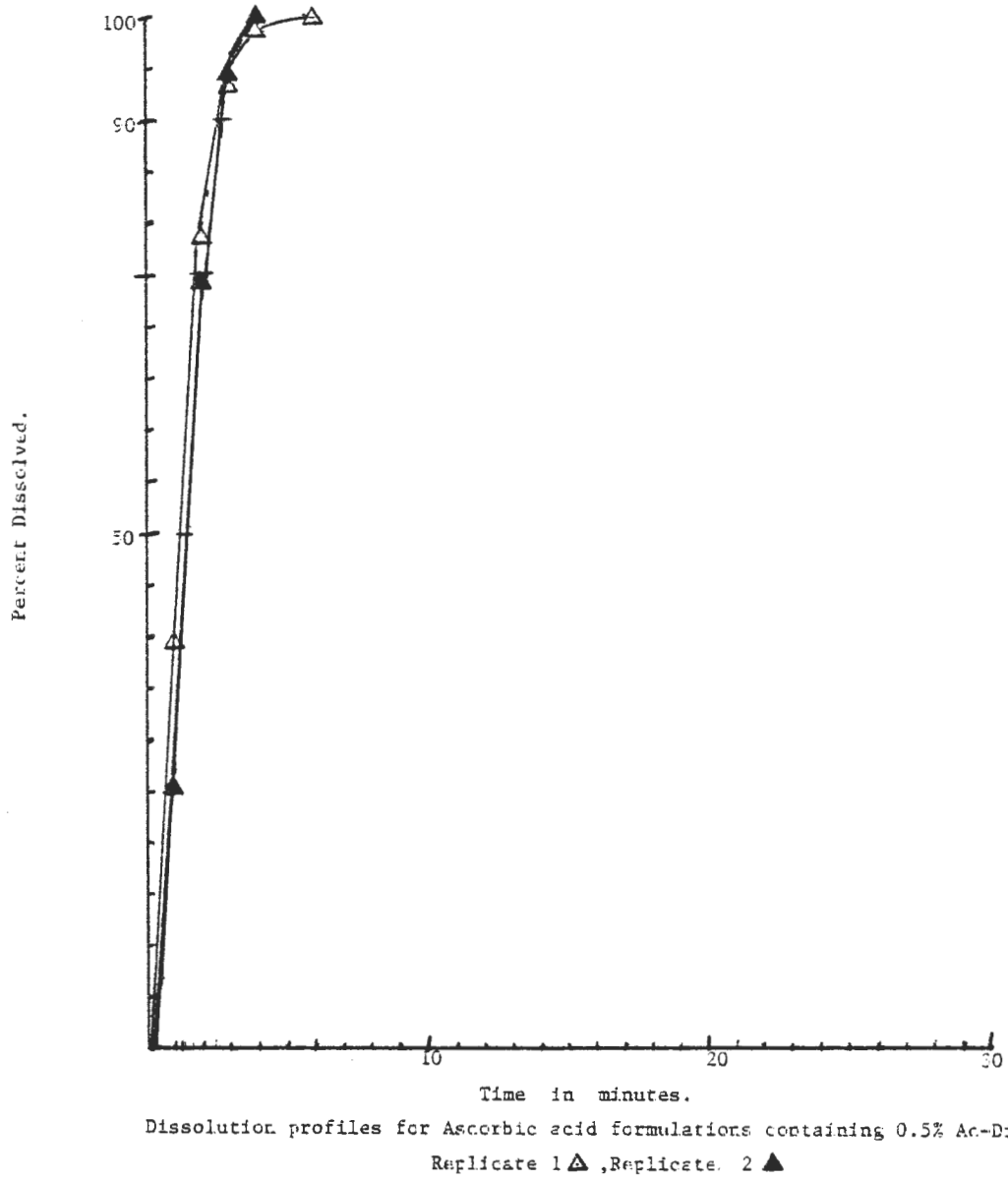


Fig. 66

Dissolution profiles.

Formulation:

Ascorbic acid 25%  
Disintegrant 1.0%  
Stearic acid 2.5%  
Emcompress ad 100%

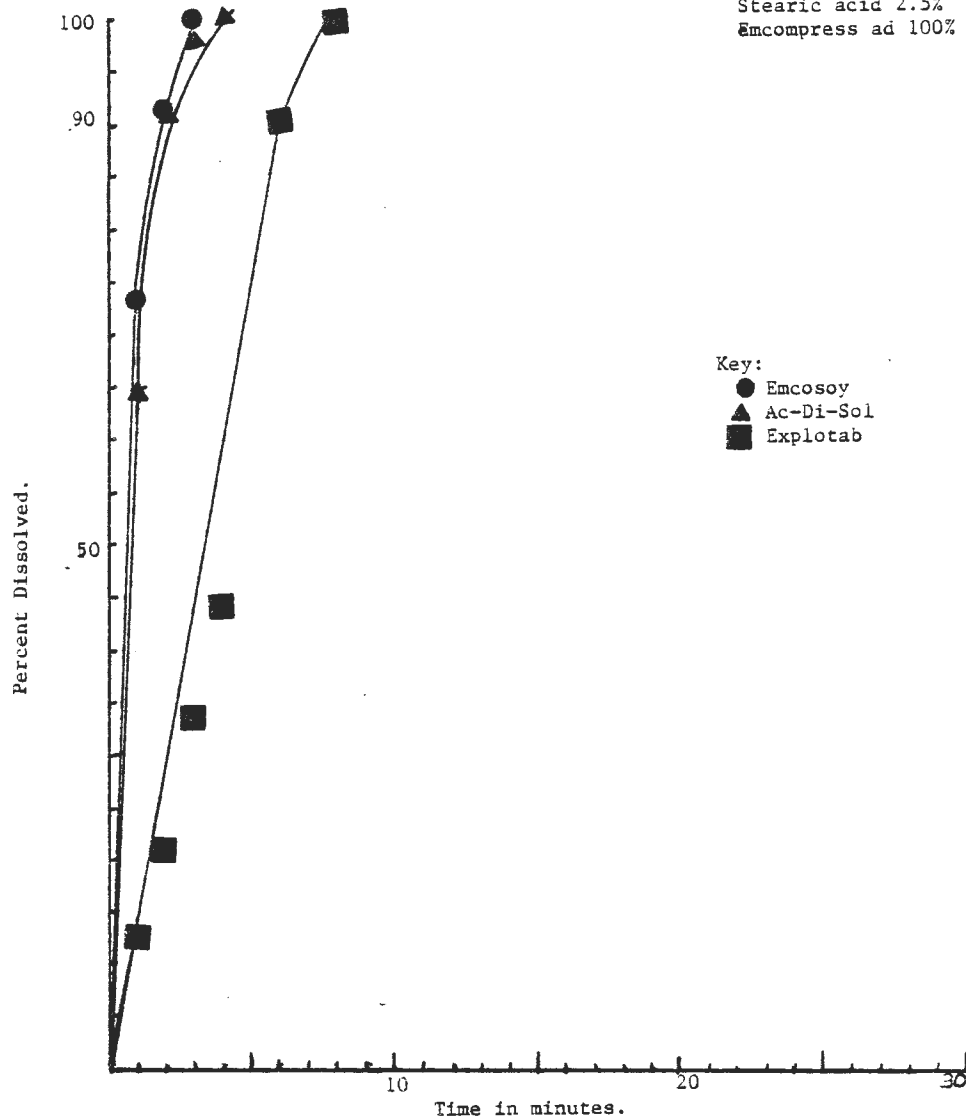
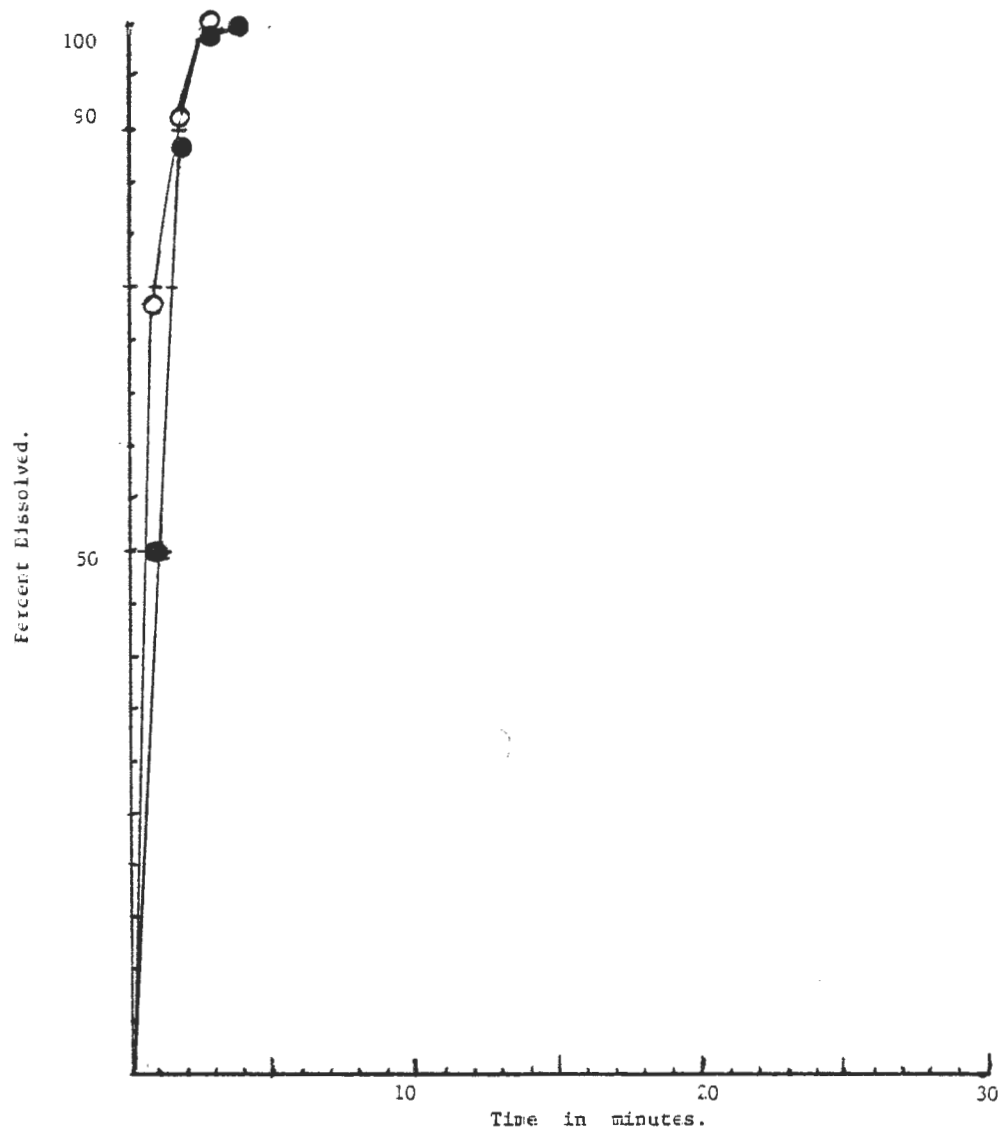
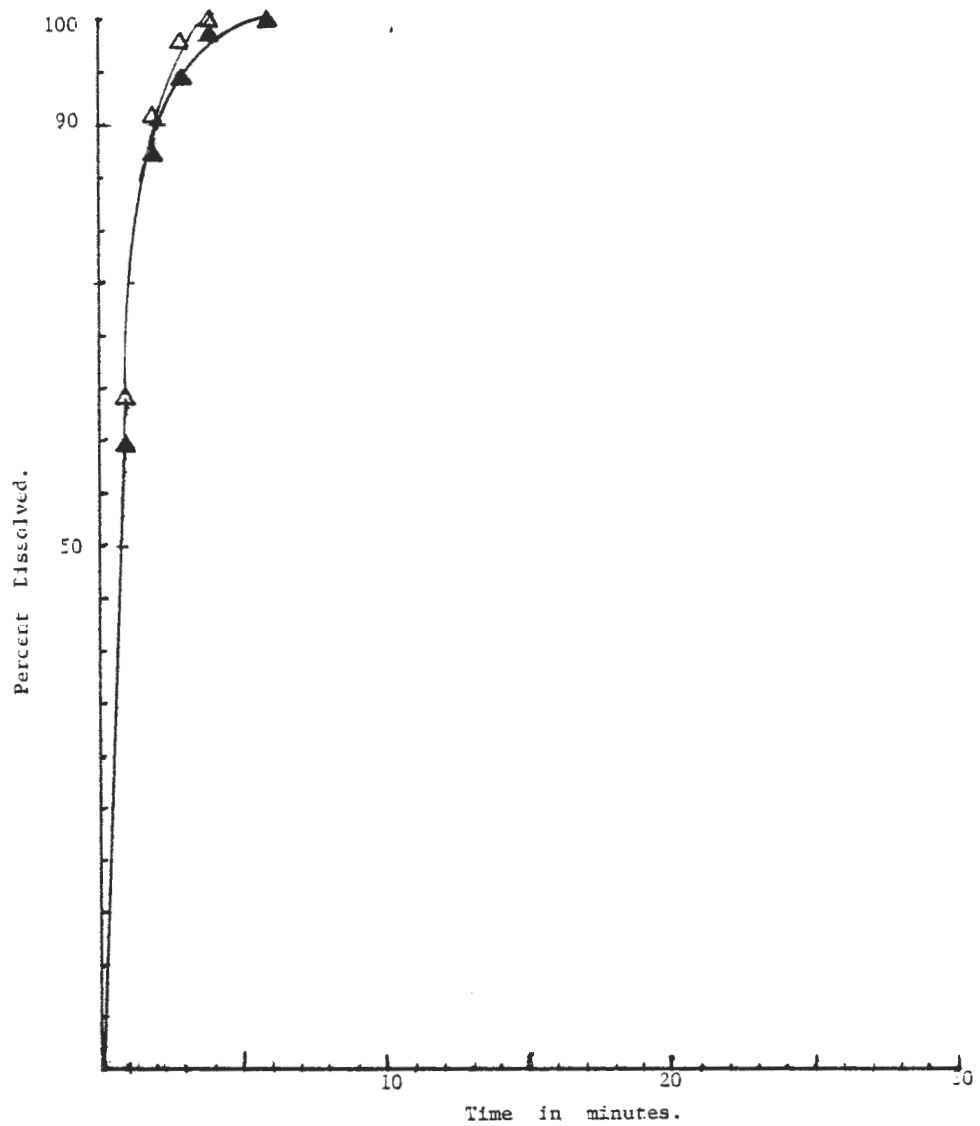


Fig. 67



Dissolution profiles for Ascorbic acid formulations containing 1% Emcesoy  
Replicate 1 (○) , Replicate 2 (●)

Fig. 68



Dissolution profiles containing 1% Ac-Di-Sol in Ascorbic acid formulations

Replicate 1 ▲ , Replicate 2 ▲

Fig. 69

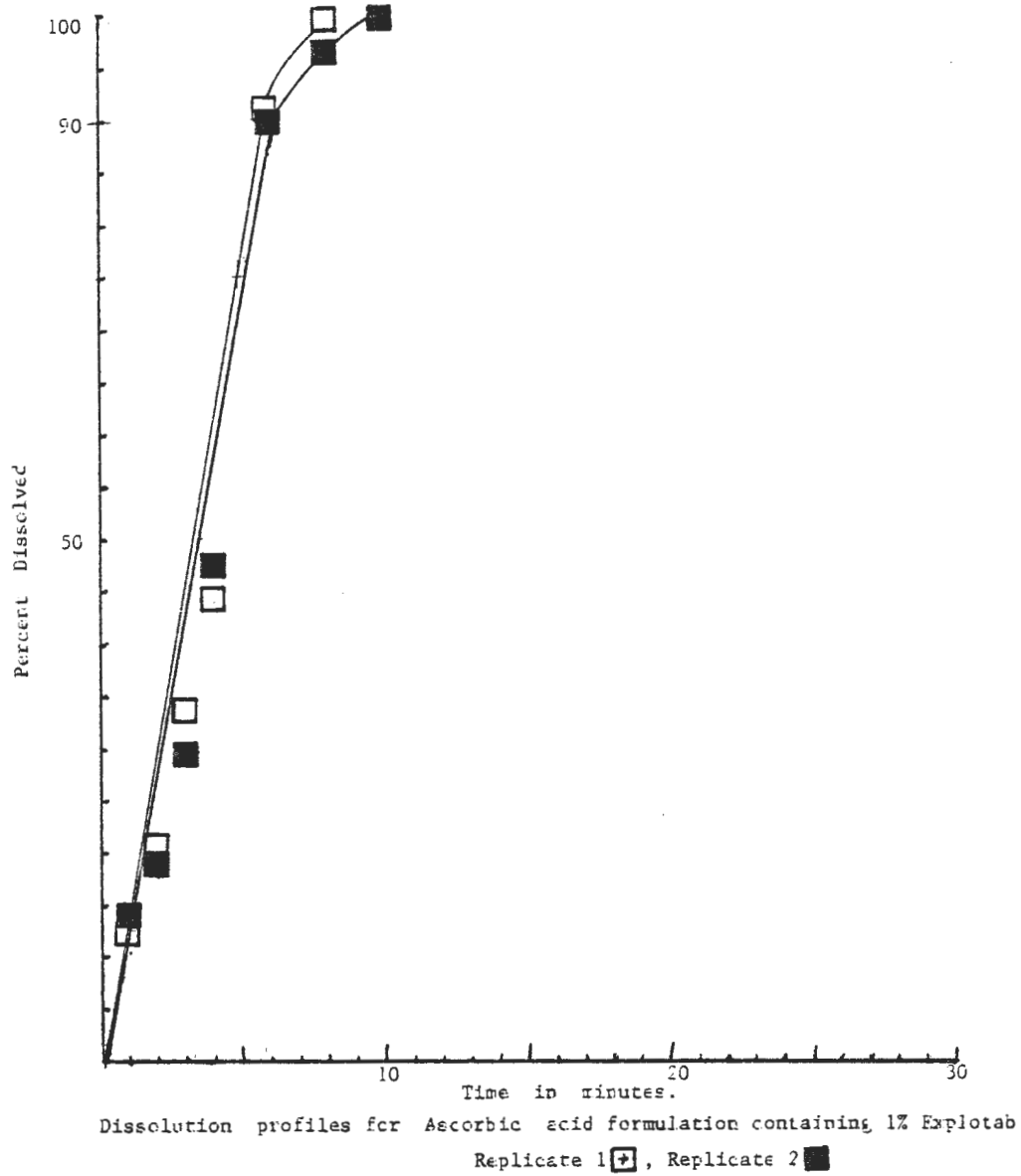


Fig. 70

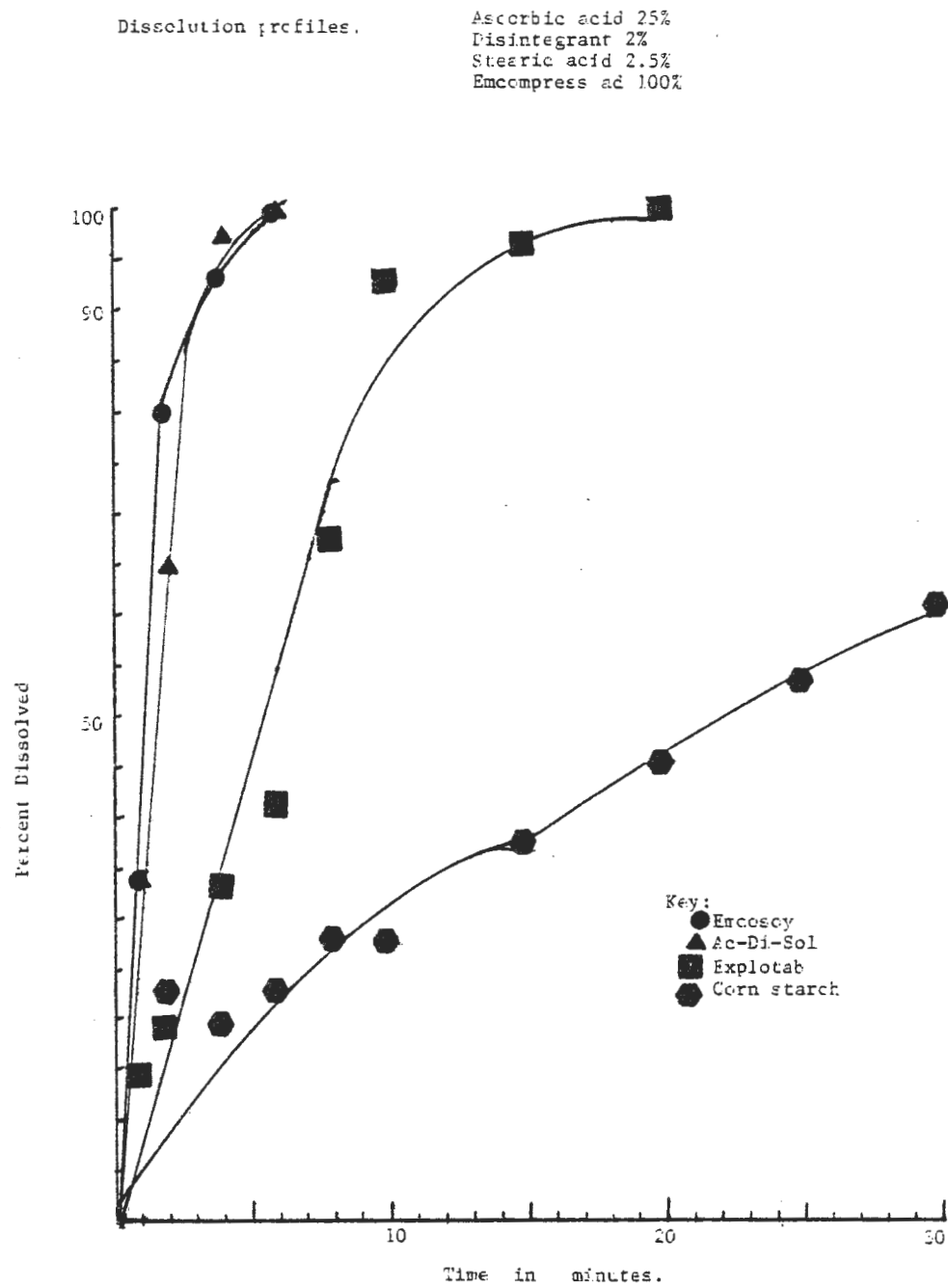
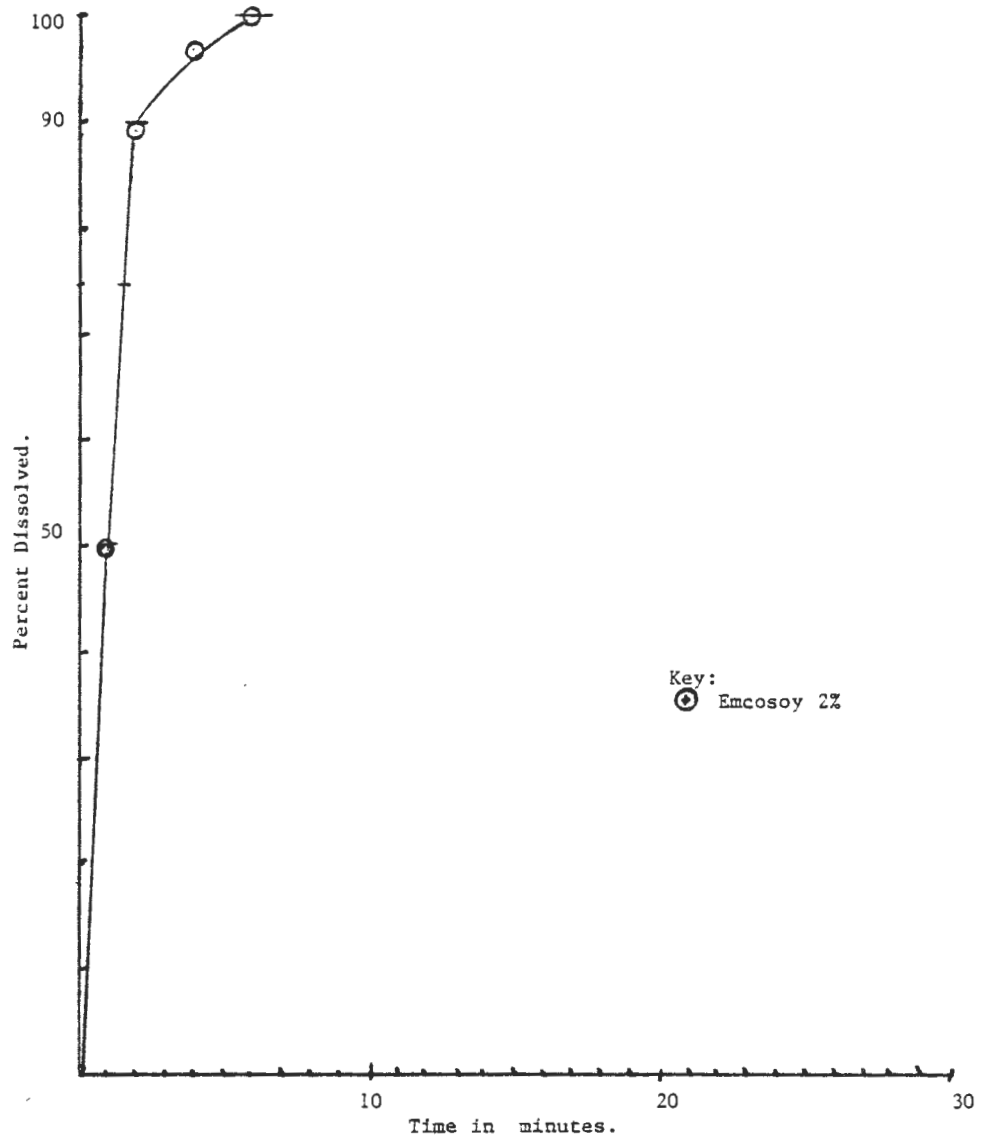


Fig. 71

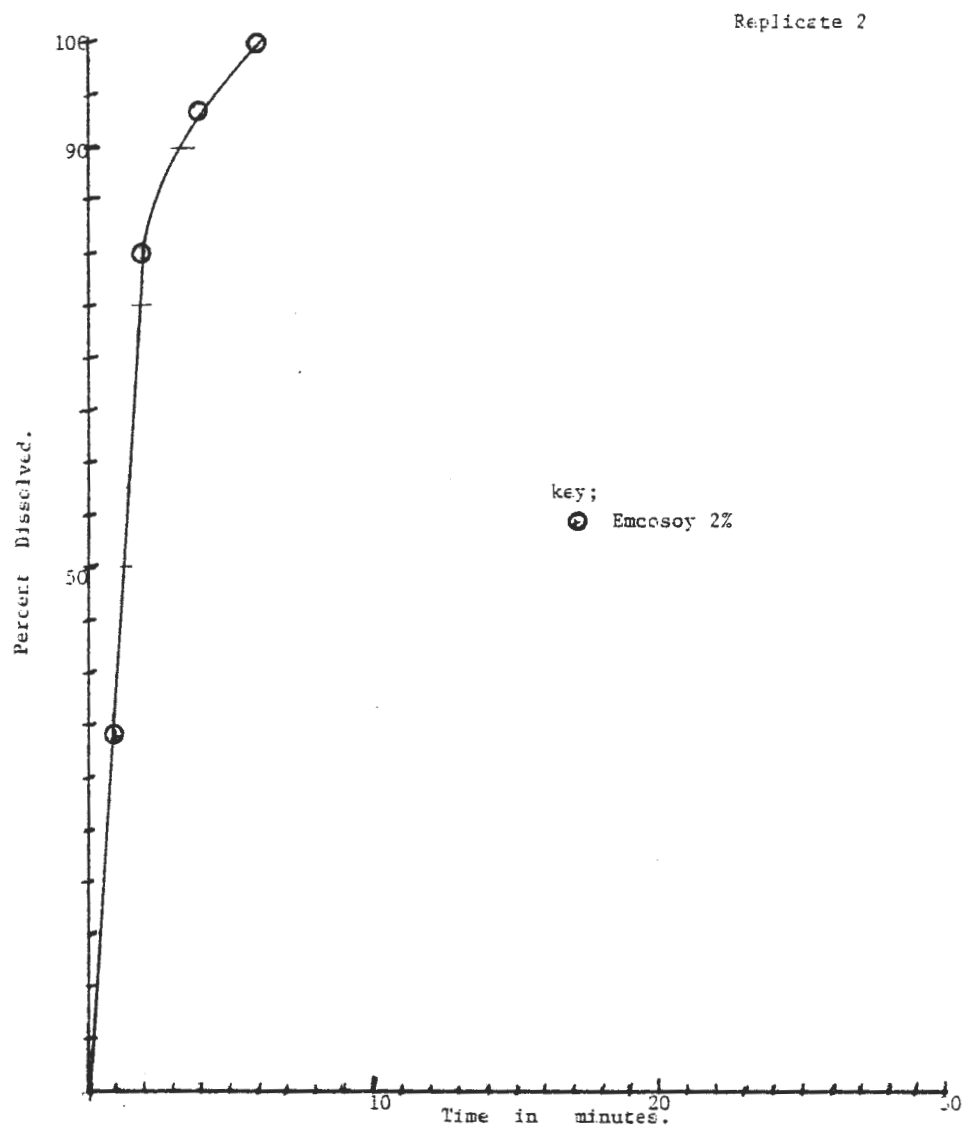
Replicate 1



Dissolution profile for Ascorbic acid formulation containing 2% of different disintegrants.

Fig. 72

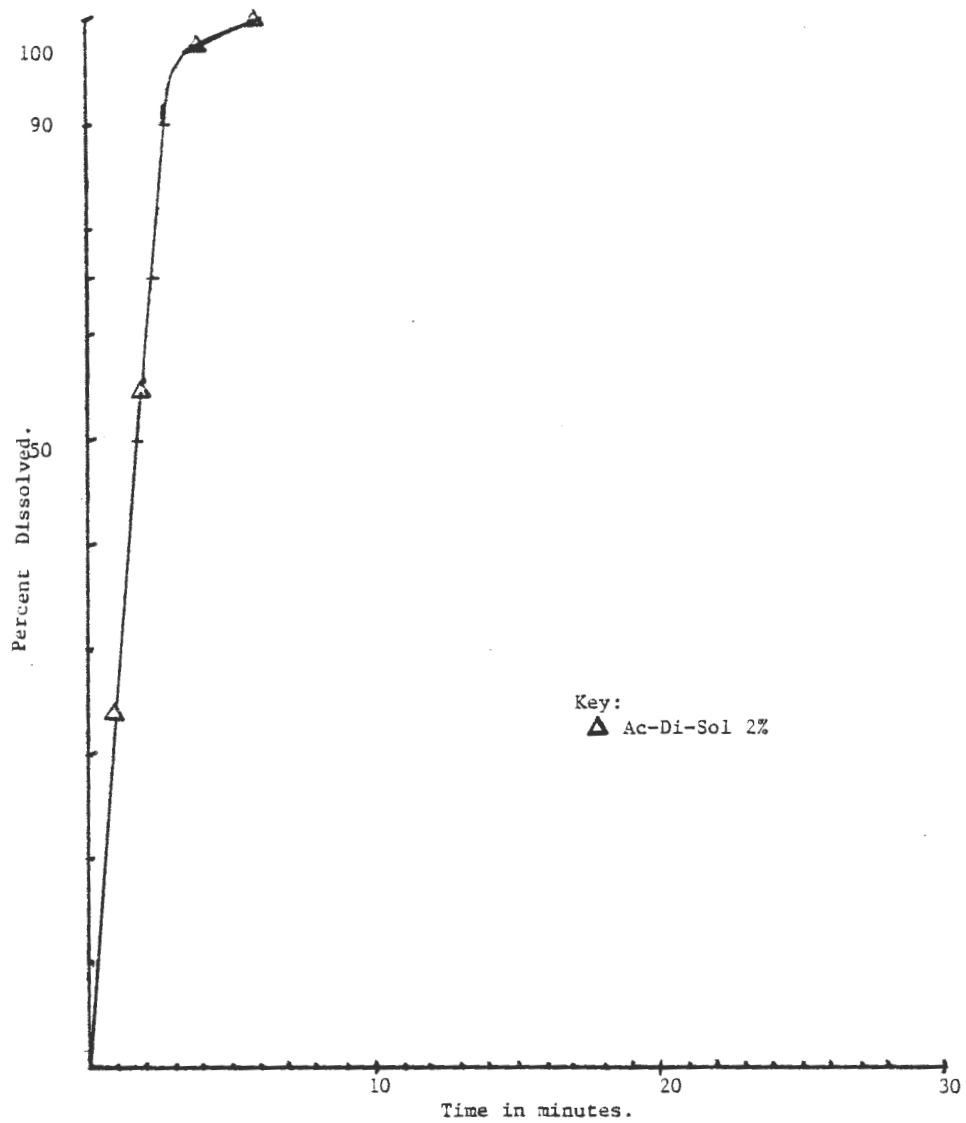




Dissolution profile for Ascorbic acid formulation containing 2% of different disintegrants.

Fig. 73

Replicate 1



DISSOLUTION PROFILE FOR ASCORBIC ACID FORMULATION CONTAINING  
2% OF DIFFERENT DISINTEGRANTS.

Fig. 74

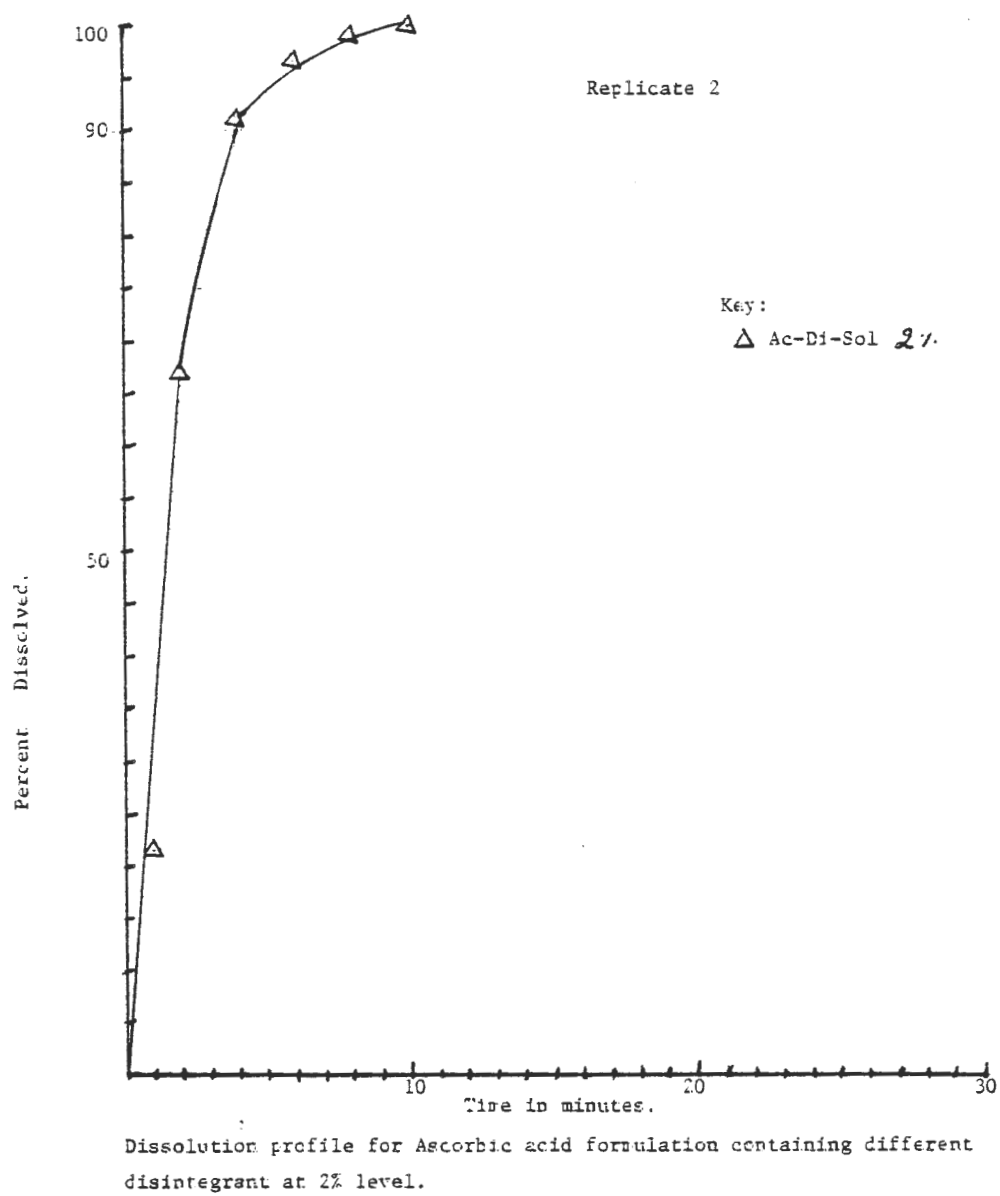
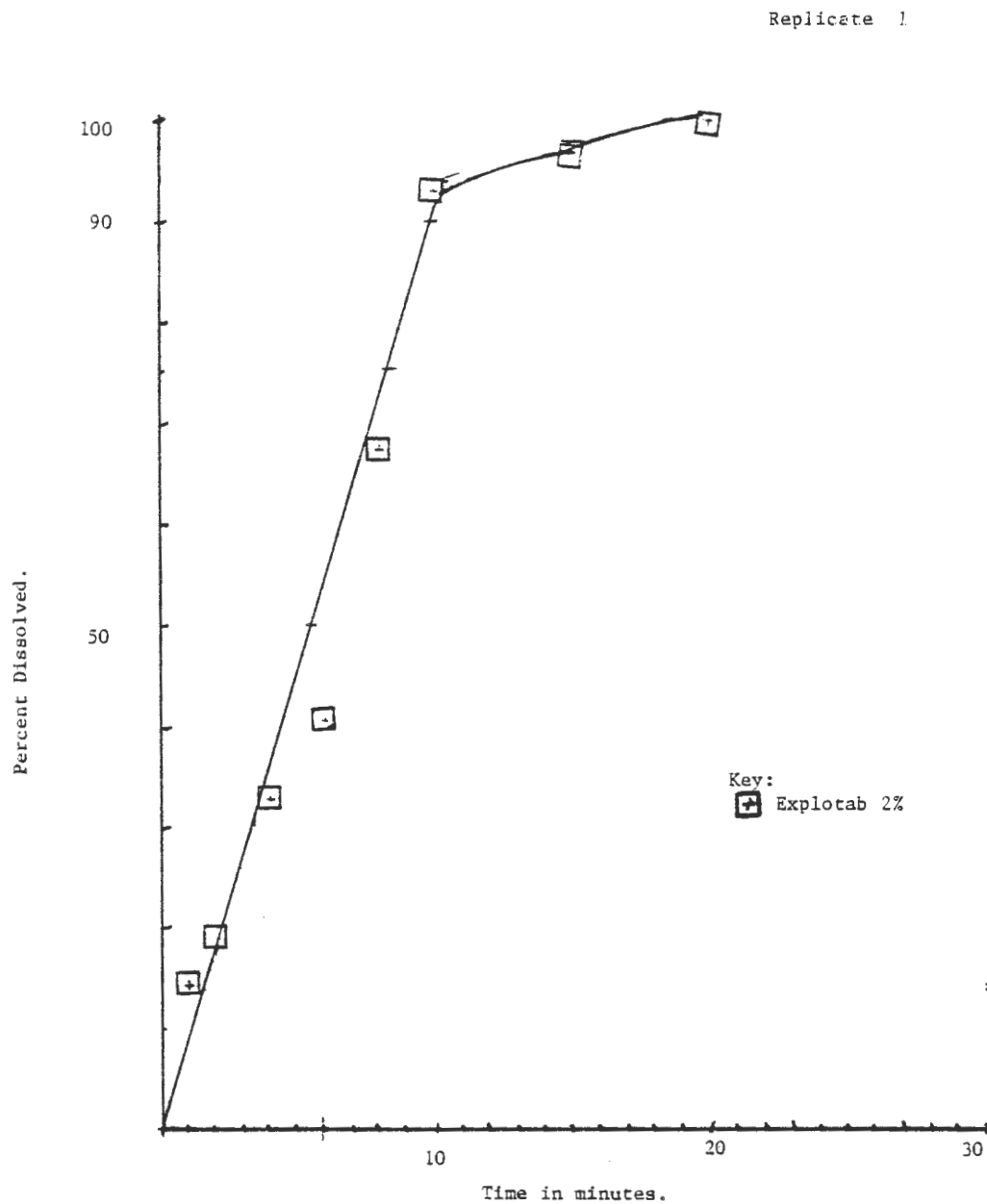


Fig. 75



DISSOLUTION PROFILE FOR ASCORBIC ACID FORMULATION CONTAINING  
2% OF DIFFERENT DISINTEGRANTS.

Fig. 76

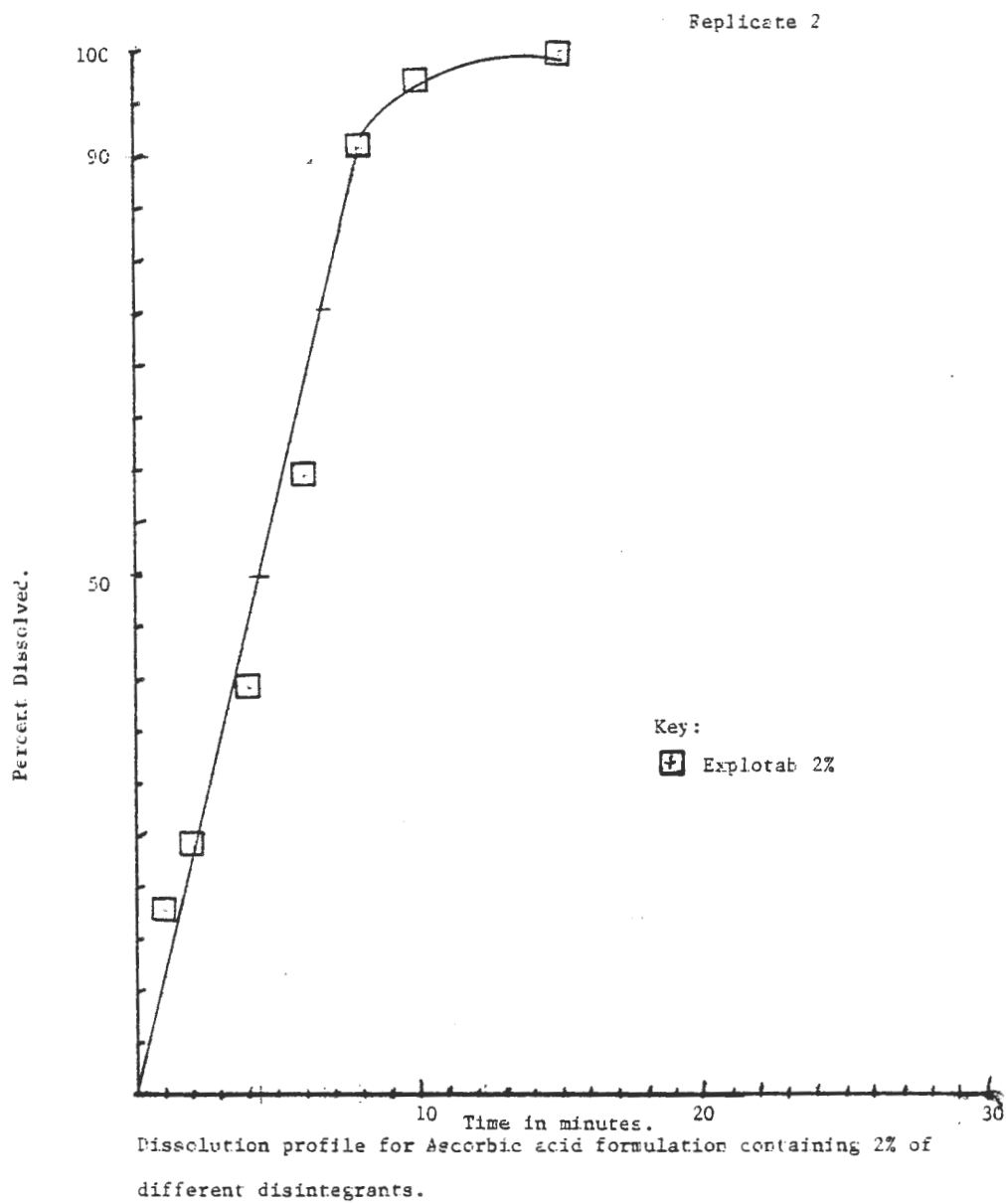
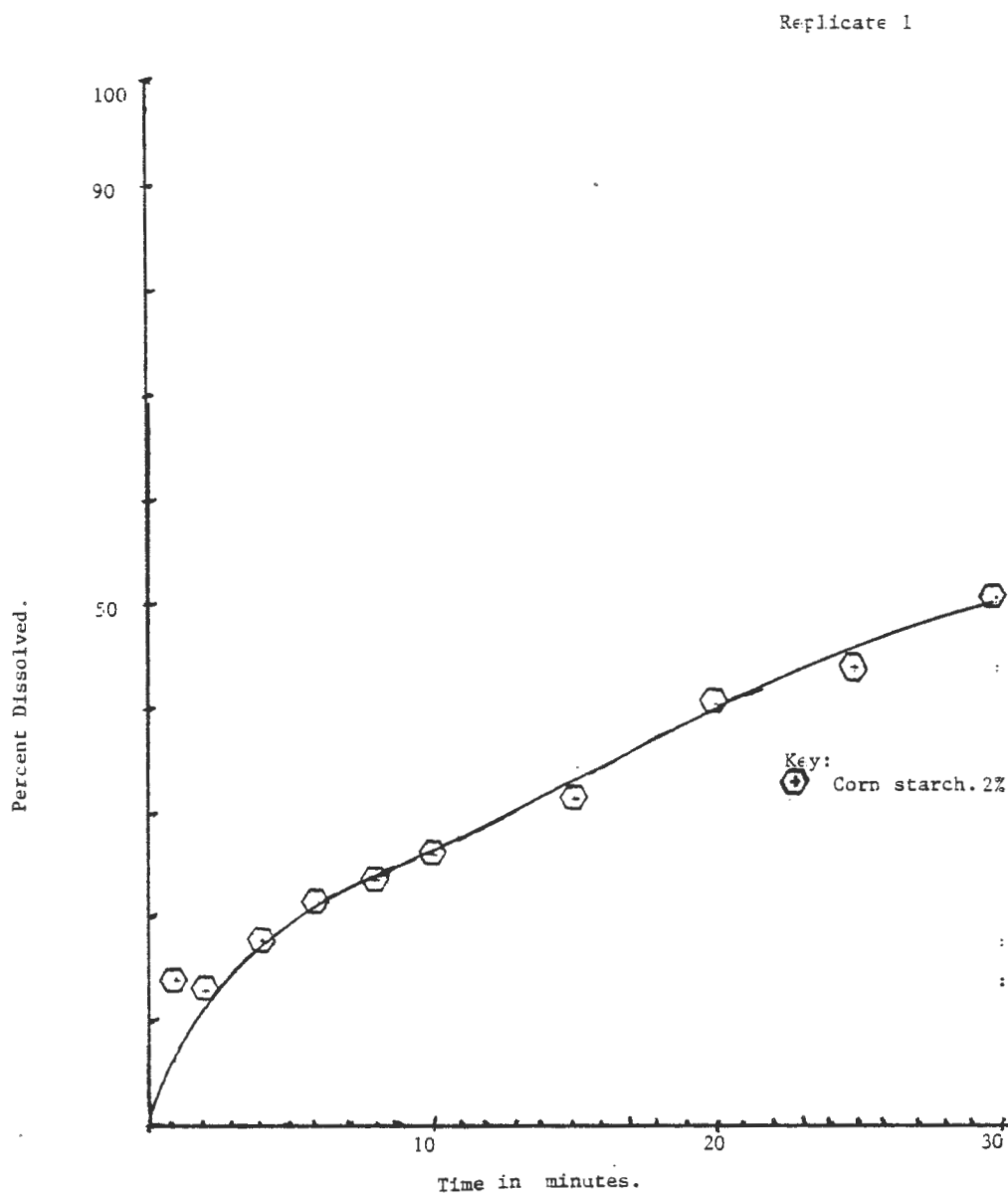
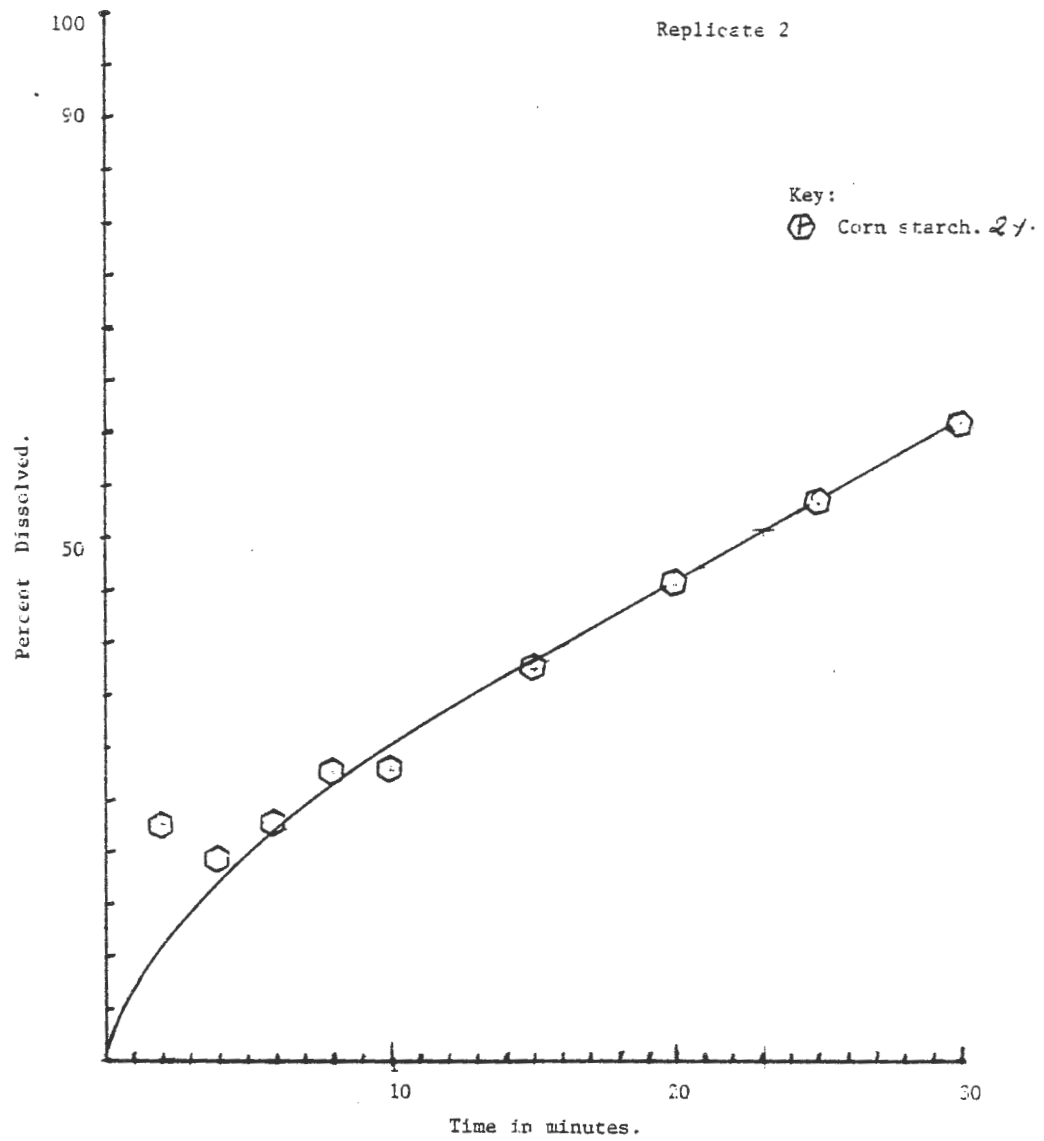


Fig. 77



DISSOLUTION PROFILE FOR ASCORBIC ACID FORMULATION CONTAINING  
2% OF DIFFERENT DISINTEGRANTS.

Fig. 78



Dissolution profile for Ascorbic acid formulation containing 2% of different disintegrant.

Fig. 79

## Dissolution Profiles.

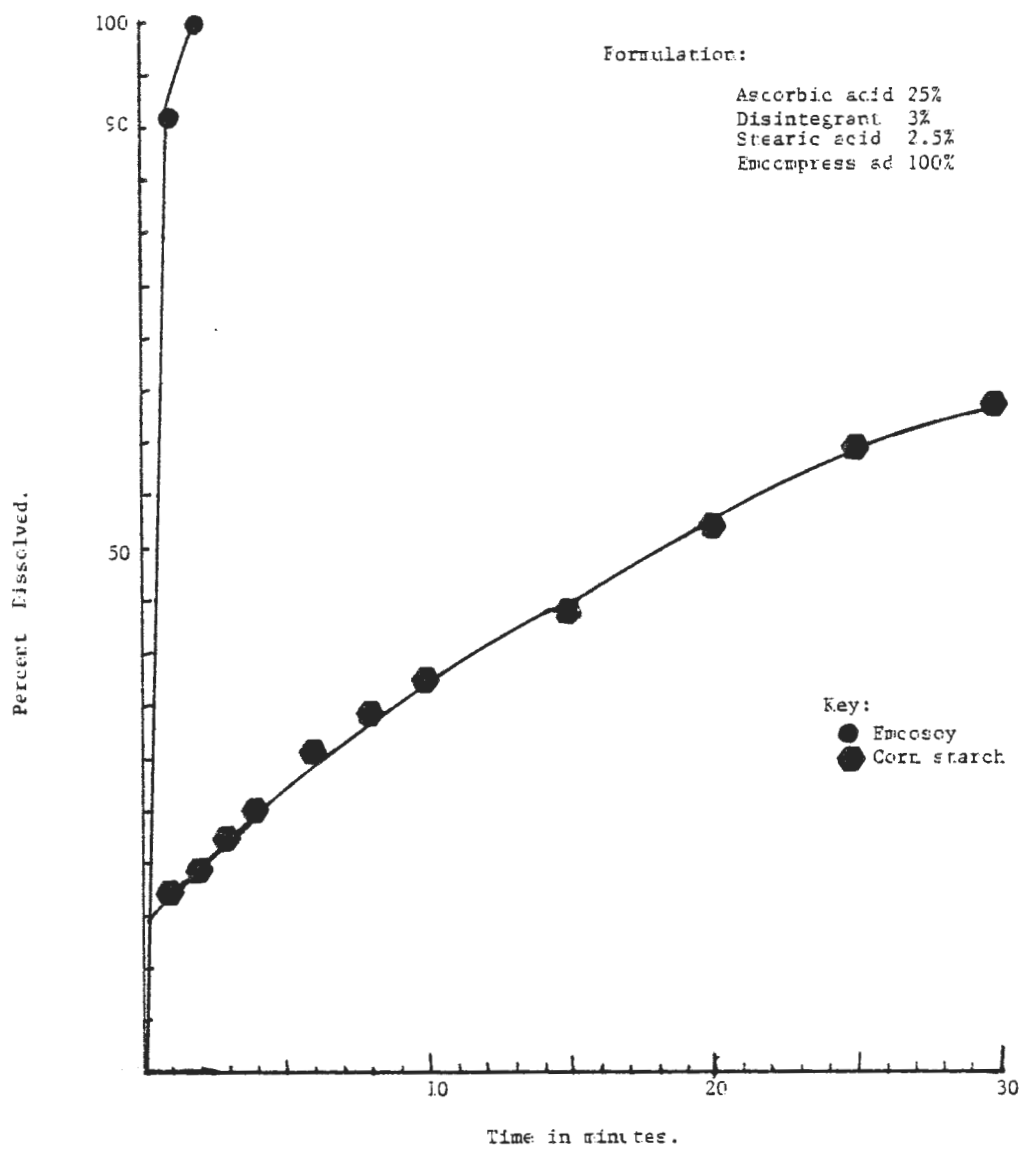
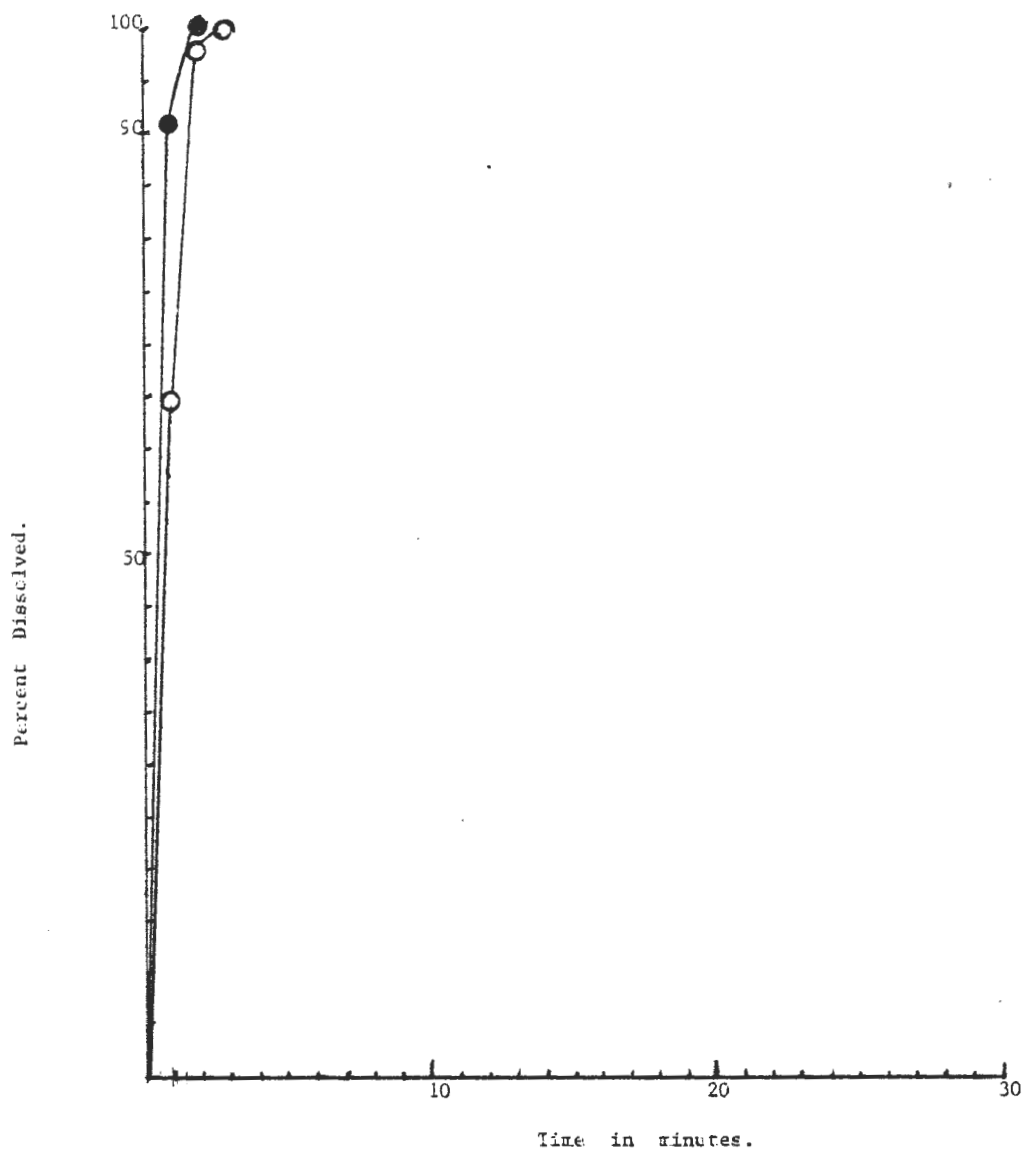


Fig. 80





Dissolution profile for Ascorbic acid formulation containing 3% Emcosoy  
Replicate 1 (○) , Replicate 2 (●)

Fig. 81

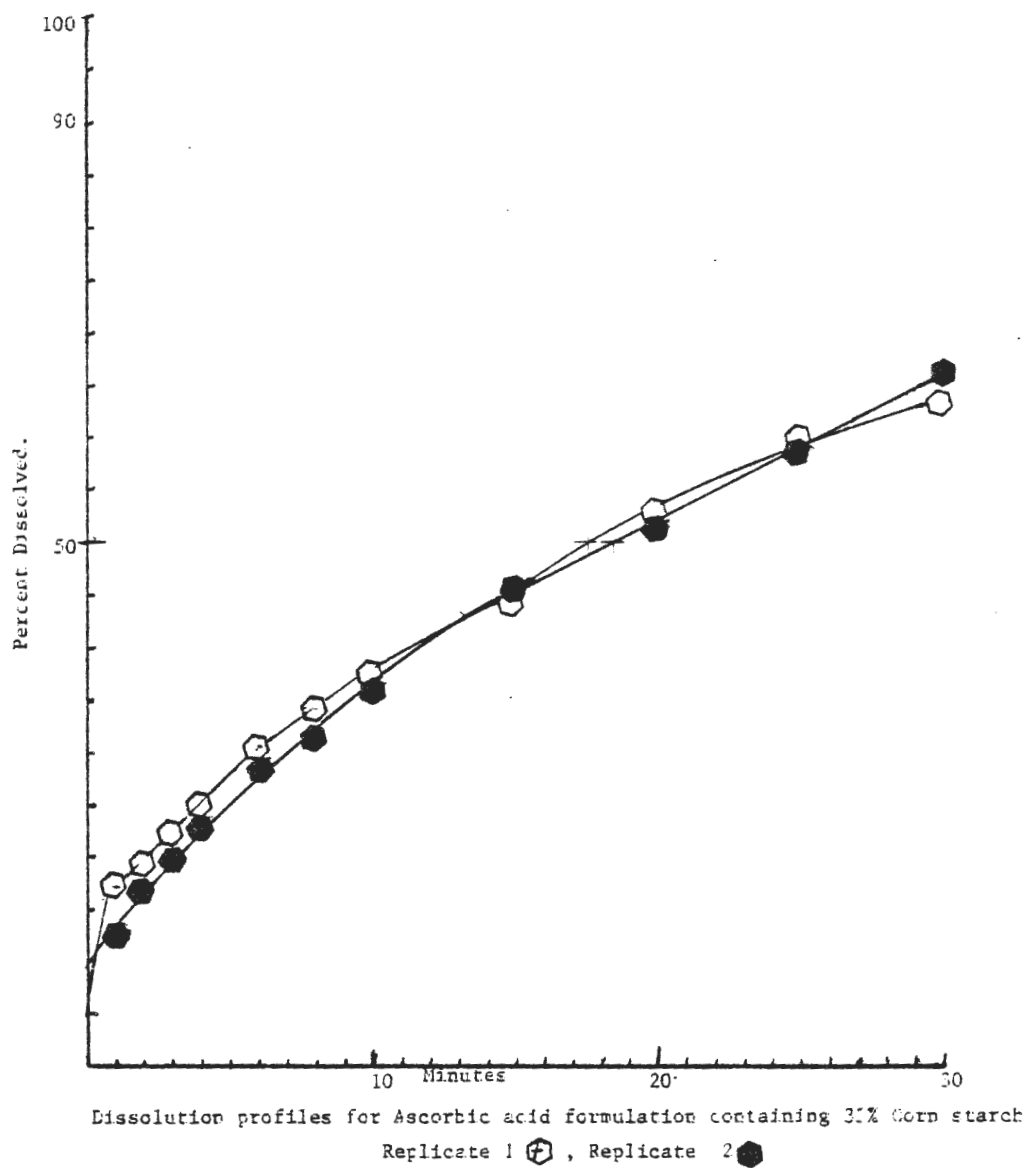


Fig. 82

## IV. CONCLUSION

Emcosoy was tested for three major mechanisms of disintegration: (i) swelling, (ii) porosity and capillary action (wicking), and (iii) deformation. Swelling and wicking were adequate in the comparison of other satisfactory disintegrants, but Emcosoy had a poor plastic deformation. The viscosity of the gel produced by Emcosoy on wetting is less than the gel produced by CLD II, but higher than that by Ac-Di-Sol. All these factors were considered separately; but in actual disintegration of a tablet, inter-relationships always take place.

Proper storage precautions are advised for this disintegrant because it has a tendency to promote microbial growth in the presence of high moisture at room temperature as observed during the wetting and drying test.

In this project, a total of 45 tablet formulations were studied, and about 100 dissolution profiles were obtained on various vitamin formulations using either Emcompress or Emdex as the tablet matrix. The results obtained clearly indicate that:

1. Emcosoy is superior to corn starch as a tablet disintegrant at two or three percent levels;
  2. Emcosoy competes favorably with other disintegrants, such as Explotab and Ac-Di-Sol, at half, one, and two percent levels;
  3. Emcosoy demonstrates superior dissolution profiles when evaluated against other disintegrants in vitamin formulations;
- and

4. Water-insoluble matrix is more helpful in the evaluation of different disintegrants.

Emcosoy is demonstrably a powerful tablet disintegrant of natural source which deserves careful consideration by all formulators.

## V. SUGGESTIONS FOR FUTURE WORK

Life is the sum of unfulfilled dreams. So is research -- there is always the nagging doubt that one could have done better.

The author would have liked to change the particle size of Emcosoy to determine its effect on disintegration and other properties of the tablet.

Emcosoy has one great advantage over other popular disintegrants in that it is of natural origin. If we want to retain this advantage, there is very little we can do to change its property. It would be of great advantage to reduce the proteinaceous fraction of Emcosoy composition (at present, about 17.5%) by some changes in processing procedure. The author anticipates that such changes would improve the plastic deformation of the Emcosoy particles; and consequently, the disintegration property. Low protein fraction would also help in protecting disintegrant at higher moisture level from lumping, as well as from microbial degradation.

REFERENCES

1. Bioavailability Considerations. Committee on Biologic Performance of Drug Products: Guidelines for Biopharmaceutical Studies in Man, Washington, D.C.: Am. Pharm. Assoc., Acad. Pharm. Sci., 1972. p 2.
2. D.S. Greene and C.T. Rhodes: An Introduction to Biopharmaceutics and Pharmacokinetics. A book for PHC 338 students at the University of Rhode Island. Fourth Edition, 1980. p XVII/1.
3. A.J. Glazko, A.W. Kinkel, W.C. Alegnani, and E.L. Holmes: An Evaluation of the Absorption Characteristics of Different Chloramphenicol Preparations in Normal Human Subjects. Clin. Pharmacol. Ther. 9, 472-483 (1968).
4. G. Levy, K. Arnold, and N. Gerber: Absorption and Dissolution Studies on Sodium Diphenylhydantoin in Capsules. Can. J. Pharm. Sci. 5, 89-92 (1970).
5. J.G. Wagner: Biopharmaceutics and Relevant Pharmacokinetics. Drug Intelligence Publication, First Edition (1971). p 66.
6. R. Shangraw, A. Mitrevej, and M. Shah: Pharm. Tech., Vol. 4, No. 10, 1980, p 49.
7. D.S. Greene and C.T. Rhodes: Introduction to Biopharmaceutics and Pharmacokinetics, Vol. 1, p v/1.
8. J.G. Wagner: Biopharmaceutics and Relevant Pharmacokinetics. Drug Intelligence Publication, First Edition (1971). p 66.
9. E. Nelson: J. Pharm. Sci., 46:607 (1957).
10. G.S. Banker, G.E. Peck, and G. Baley: Tablet Formulation and Design. In: Pharmaceutical Dosage Forms: Tablet. Vol. 1 (H.A. Lieberman and L. Lachman, Eds.) Marcel Dekker, Inc. 1980, p 70.
11. G. Levy, J.M. Antkowiak, J.A. Procknar, and D.D. White: J. Pharm. Sci., 52:1047 (1963).
12. L. Lachman. J. Pharm. Sci. S4:1519 (1965).
13. P. Rieckmann. Pharm. Z. 34:1207 (1971).
14. F.A. Campagna, G. Crulton, R.A. Mirigian, and E. Nelson: J. Pharm. Sci. 52:605 (1963).
15. The National Formulary, 13th Edition, Mack Publishing Co., Easton, PA. p 878.

16. J.M. Richards: U.S. Pat. 216, 107 (1879).
17. Sheth, Bandelin, and Shangraw: In: Pharmaceutical Dosage Forms: Vol. 1, p 135.
18. G.S. Banker, G.E. Peck, and G. Baley: Ibid, p 86.
19. W. Lowenthal: J. Pharm. Sci. Vol. 61, No. 11, 1695-1711.
20. R.F. Shangraw, J.W. Wallace, and F.M. Bowers: Pharm. Tech., Vol. 5, No. 10, pp 44-60.
21. J. Bennett: Pharm. Acta Helv. 12, 1337 (1937).
22. L. Chwialkowska and L. Krowczynski: Acta. Pol. Pharm. 25, 583 (1958).
23. I. Krowczynski, K. Kolarski, and K. Swoboda: Ibid, 25, p 443 (1968).
24. H. Berry and C. Ridout: J. Pharm. Pharmacol: 2, 619 (1950).
25. F. Jaminet: J. Pharm. Belg. 19, 144 (1964).
26. H.V. Czetsch-Lindenwald, F. Elkhawas, and R. Tawaski: J. Soc. Cosmet. Chem. 16, 251 (1965).
27. E.E. Borzunov and T.Y. Nesmiyan: Khim-Form. Zh., 2, 44 (1968); through Int. Pharm. Abstr., 8, 1283 (1971).
28. E.E. Borzunov and S.M. Shevchenko: Formatsiya (Moscow) 13, 20 (1969); through Chem. Abstr. 71, (1969).
29. K.S. Manudhane, A.M. Contractor, H.Y. Kim, and R.F. Shangraw: J. Pharm. Sci. 58:616 (1969).
30. N.R. Patel and R.E. Hopponen: J. Pharm. Sci. Vol. 55, 1966, pp 1065-1068.
31. V.E. Nurenberg: Pharm. Ind., Vol. 34, 1972, pp 193-206.
32. F. Modezejewski and L. Wochna: Acta. Pol. Pharm. Vol. 28, 1965, pp 396-402.
33. C.D. Fox, M.D. Richman, G.E. Reier, and R.F. Shangraw: Drug Cosmet. Ind. 92, 161 (1963).
34. T. Higuchi, A.N. Rao, L.W. Burre, and J.V. Swintosky: J. Amer. Pharm. Ass. Sci. Ed. 42, 194 (1953).

35. D. Ganderton and A.B. Selkiek: J. Pharm. Pharmacol. 22, 345 (1970).
36. A.B. Selkirk and D. Ganderton: Ibid. 22, 795 (1970).
37. Ibid. 22, 865 (1970).
38. B. Reich and F. Gstiener: Arch. Pharm. 301, 830 (1968).
39. P. Fuchs, Ibid., 303, 471 (1970).
40. H. Nogami, H. Fukuzawa, and Y. Nakai: Chem. Pharm. Bull., 11, 1389 (1963).
41. H. Nogami, J. Hasegawa, and M. Miyamoto: Ibid., 15, 279 (1967).
42. T. Bano, T. Szarvas, and L. Aradi: Pharm. Zentrath. 100, 221 (1961).
43. U. Bogs and H. Moldenhauer: Pharmazie, 19, 708 (1964).
44. R. Yamamoto and T. Tahahashi: Ann. Rep. Shiongi Res. Lab. 3, 310 (1953).
45. H. Matsumaru: Yakugaku Zasshi, 78, 1201 (1958).
46. K. Minzel and W. Kagi: Sci. Pharm. 24, 237 (1956).
47. T.A. Fakouki and L. Krowczynski: Acta. Pol. Pharm. 27, 593 (1970).
48. R.A. Burrus and W. Lowenthal: J. Pharm. Sci. 60, 1325 (1971).
49. K.C. Commons, A. Bregen and G.C. Walker: J. Pharm. Sci. 58, 1253 (1968).
50. H. Nogami, T. Nagai, E. Fukuoka, and T. Sonobe: Chem. Pharm. Bull. 17, 1450 (1969).
51. P. Singh, S.J. Desai, A.P. Simonelli, and W.I. Higuchi: J. Pharm. Sci., 57, 217 (1968).
52. H. Burlinson and C. Pickering: J. Pharm. Pharmacol. 2, 630 (1950).
53. L.C. Curlin: J. Pharm. Assoc. Sci. Ed. Vol. 40, 1955, p 16.
54. A.M. Schwartz: Ind. Eng. Chem. 61, 10 (1969).
55. H. Nogami, T. Nagai, and H. Uchida: Chem. Pharm. Bull. 14, 132 (1966).



56. J.H. Wood: Pharm. Acta. Helv. 42, 129 (1967).
57. H. Hess: Pharm. Tech. Vol. 2, No. 9, 1978, pp 36-37.
58. C. Fuhrer: Informationsdienst APV, Vol. 10, 1964, p 58, From: Pharm. Acta. Helv., Vol. 48, 1973, pp 589-609.
59. C.T. Rhodes: In: Proceedings of 1981 Pharm. Tech. Conf., in New York.
60. "Emcosoy," Edward Mendell Technical Bulletin, Edward Mendell, Carmel, NY.
61. E.M. Rudnic, S. Weich, P. Bernardo, and C.T. Rhodes: Drug Development and Industrial Pharmacy, 8 (1), 87, 1981.
62. P. Fuchs: Arch. Pharm., 1970, p 303. From: Pharm. Acta. Helv. Vol. 48, 1973, pp 589-609.
63. H. Hess: In: Pharm. Tech., Vol. 2, No. 9, 1978, pp 36-37.
64. C. Fuhrer: Informationsdienst APV, Vol. 10, 1964, p 58. From: Pharm. Acta. Helv., Vol. 48, 1979, pp 589-609.
65. C.M. Lee and R.T. Toledo: Journal of Food Science, Vol. 41, 1976, pp 391-397.
66. K.N. Wai, H.G. Dekay, and G.S. Banker: J. Pharm. Sci., Vol. 11, 1076-1080, 1962.
67. R.B. Wortz: J. Pharm. Sci., Vol. 9, 1169-1173, 1967.
68. "The National Formulary," XIV Ed. Mack Publishing Co., Easton, PA, 1975.
70. W. Lowenthal: Pharm. Acta. Helv., 48, 1973, pp 589-609.
71. H. Nogami, J. Hasegawa, and M. Miyamoto: Chem. Pharm. Bull., 15(3), 279-289 (1967).
72. E.M. Rudnic, J.M. Lausier, R.N. Chilamkurti, and C.T. Rhodes: In: Drug Development and Industrial Pharmacy, 6, 291 (1980).

## VII BIBLIOGRAPHY

- Banker G.S., Peck G.E., and Baley G: Tablet Formulation and Design. In: Pharmaceutical Dosage Forms: Tablet. Vol. 1. (H.A. Lieberman and L. Lachman, Eds.), Marcel Dekker, Inc., 1980, p. 70.
- Banker G.S., Peck G.E., and Baley G: Pharmaceutical Dosage Forms: Vol. 1. p. 86.
- Bano T, Szarvas F, and Aradi L: Pharm. Zentrath, 100, 221.
- Bennet J: Pharm. Acta. Helv. 12, 1337 (1937).
- Berry H, and C. Ridout: J. Pharm. Pharmacol. 2, 619 (1950).
- Bogs V, and Moldenhauer H: Pharmazie, 19, 708 (1964).
- Burlinson H, and Pickering C: J. Pharm. Pharmacol. 2, 630 (1950).
- Burrus R.A. and Lowenthal W: J. Pharm. Sci. 60, 1325 (1971).
- Borzunov E.E., and Shevchenko S.M.: Formatsiya (Moscow) 18, 20 (1969); through Chem. Abstr. 71 (1969).
- Borzunov E.E., and Nesmiyan T.Y.: Khim-Form. Zh. 2, 44 (1968); through Int. Pharm. Abstr. 8, 1283 (1971).
- Campagna F.A., Crulton G, Nirigian R.A., and Nelson E: J. Pharm. Sci. 52:605 (1963).
- Chwialkowska L, and Krowczynski L: Acta. Pol. Pharm. 25, 583 (1958).
- Commons K.C., Bregen A, and Walker G.C.: J. Pharm. Sci. 58, 1253 (1968).
- Curlin L.C.: J. Pharm. Assoc. Sci. Ed., Vol. 40, 1955, p 16.
- Czetsch-Lindenwald H.V., Elkhawas F, and Tawaski R: J. Soc. Cosmet. Chem. 16, 251 (1965).
- Fakouki T.A., and Krowczynski L: Acta. Pol. Pharm. 27, 593 (1970).
- Fox C.D., Richman M.D., Reier G.E., and Shangraw R.F.: Drug Cosmet. Ind. 92, 161 (1963).
- Fuchs P: Arch. Pharm. 1970, p. 303. From: Pharm. Acta. Helv. Vol. 48, 1973, pp 589-609.

Fuchs P: Arch. Pharm. 303, 471 (1970).

Fuhrer C: Informationsdiense APV, Vol. 10, 1964, p. 58. From: Pharm. Acta. Helv., Vol. 48, 1979, pp 589-609.

Fuhrer C: Informationsdiense APV, Vol. 10, 1964, p 58. From: Pharm. Acta. Helv., Vol. 48, 1979, pp 589-609.

Ganderton C, and Selkeik A.B.: J. Pharm. Pharmacol. 22, 345 (1970).

Glazko A.J., Kinkel A.W., Alegnani W.C., and Holmes E.L.: Clin. Pharmacol. Ther. 9, 472-483 (1968).

Greene D.S., and Rhodes C.T.: An Introduction to Biopharmaceutics and Pharmacokinetics. A book for PHC 338 students at the University of Rhode Island. Fourth Edition, 1980, p v/i.

Greene D.S., and Rhodes C.T.: Ibid. p XVII/1.

Hess H: Pharm. Tech., Vol. 2, No. 9, 1978, pp 36-37.

Higuchi T, Rao A.N., Burre L.W., and Swintosky J.V.: J. Amer. Pharm. Ass. Sci. Ed. 42, 194 (1953).

Jaminet F: J. Pharm. Belg. 19, 144 (1966).

Krowczynski I., Kolarski K., and Swoboda K.: Acta. Pol. Pharm., p 443 (1968).

Lachman L.: J. Pharm. Sci. 54:1519 (1965).

Lee C.M., and Toledo R.T.: Journal of Food Science, Vol. 41, 1976, pp 391-397.

Levy G., Arnold K., and Grebel N.: Can. J. Pharm. Sci., 5, 89-92 (1970).

Levy G., White D.D., Procknal J.A., and Antkowiak J.M.: J. Pharm. Sci. 52:1047 (1963).

Lowenthal W.: Pharm. Acta. Helv., 48, 1973, pp 589-609.

Lowenthal W.: J. Pharm. Sci., Vol. 61, No. 11, 1695-1711.

Manudhane K.S., Contractor A.M., Kim H.Y., and Shangraw R.F.: J. Pharm. Sci., 58:616 (1969).

- Mastumaru H.: Yakugaku Zasshi, 78, 1201 (1958).
- Minzel K., and Kagi W.: Sci. Pharm. 24, 237 (1956).
- Modezejewski F., and Wochna L.: Acta. Pol. Pharm., Vol. 23, 1965, pp 396-402.
- Nelson E.: J. Pharm. Sci., 46:607 (1957).
- Nogami H., Hasagawa J., and Miyamoto M.: Chem. Pharm. Bull., 15, 279 (1967).
- Nogami H., Fukuzawa H., and Nakai Y.: Ibid 11, 1389 (1963).
- Nogami H., Nagai T., and Uchida H.: Ibid, 14, 132 (1966).
- Nogami H., Hasegawa J., and Miyamoto M.: Ibid, 279-289 (1967).
- Nogami H., Nagai T., Fukuoka E., and Sonobe T.: Ibid, 17, 1450 (1969).
- Nurenberg V.E.: Pharm. Ind., Vol. 34, 1972, pp 193-206.
- Patel N.R., and Hopponsen R.E.: J. Pharm. Sci., Vol. 55, 1966, pp 1065-1068.
- Reich B., and Gstiener F.: Arch. Pharm. 301, 830 (1968).
- Reickmann P.: Pharm. Z., 34:1207 (1971).
- Rhodes C.T.: at Proceedings of 1981 Pharm. Tech. Conf. in New York, 1981.
- Richards J.M.: U.S. Pat., 216, 107 (1979).
- Rudnic E.M., Weich S., Bernardo P., and Rhodes C.T.: Drug Development and Industrial Pharmacy, 8CD, 87, 1981.
- Rudnic E.M., Lausier J.M., Chilamkurti R.N., and Rhodes C.T.: In: Drug Development and Industrial Pharmacy, 6, 291 (1980).
- Schwartz A.M.: Ind. Eng. Chem. 61, 10 (1969).
- Selkiek A.B., and Ganderton D.: J. Pharm. Pharmacol. 22, 795 (1970).

Sheth, Bandelin and Shangraw: In: Pharmaceutical Dosage Forms, Vol. 1, p 135.

Singh P., Desai S.J., Simonelli A.P., and Higuchi W.I.: J. Pharm. Sci., 57, 217 (1968).

Wagner J.G.: Biopharmaceutics and Relevant Pharmacokinetics. Drug Intelligence Publication, First Edition (1971), p 66.

Wai K.N., Dekay H.G., and Banker G.S.: J. Pharm. Sci., Vol. 11, 1076-1080, 1962.

Wood J.H.: Pharm. Acta. Helv., 42, 129 (1967).

Wortz R.B.: J. Pharm. Sci., Vol. 9, 1169-1173, 1967.

Yamamoto R., and Tahahashi T.: Ann. Rep. Shiongi Res. Lab., 3, 310 (1953).

## VIII APPENDIX

## 1. Computer Program For The Calculation Of Mean, Standard Deviation And Relative Standard Deviation

```

100 REM -----
110 REM I=COUNTER
120 REM N=NUMBER READ
130 REM S=SUM OF N
140 REM T=SUM OF N SQUARED
150 REM V=VARIANCE
160 REM D=STANDARD DEVIATION
170 REM R=RELATIVE STD, DEVIATION
180 REM S/(I-1)=MEAN
190 REM -----
200 LET I=0
210 LET S=0
220 LET T=0
230 LET V=0
240 LET D=0
250 LET R=0
260 FOR I=1 TO 100
270 INPUT N
280 IF N=999 THEN 320
290 LET S=S+N
300 LET T=T+N**2
310 NEXT I
320 LET V=(T-(S**2/(I-1)))/(I-2)
330 LET D=V**.5
340 LET R=(D/(S/(I-1)))*100
350 PRINT "MEAN=";S/(I-1)
360 PRINT "STANDARD DEVIATION=";D
370 PRINT "R.S.D.=";R
380 GO TO 200
390 END

```

## 2. Computer Program For The Calculation Of Percentage Of Drug Dissolved

```

100 DIMENSION TIME (30), ABS(30), CONC (30), AMT (30), TITLE (6)
110 WRITE (6,155)
120 155 FORMAT (//1X,'ENTER THE NAME OF FORMULATION')
130 READ (5,160) (TITLE (I), I=1,6)
140 160 FORMAT (6A4)
142 WRITE (6,112)
144 112 FORMAT(//1X, 'ENTER TOTAL VOLUME OF DISSOLUTION MEDIUM')
146 READ (5,*)TV
150 WRITE (6,105)
160 105 FORMAT (//1X,'ENTER SLOPE OF BEERS PLOT (ABS/MCG PER ML)')
170 READ (5,*) SLOPE
180 100 FORMAT (//1X,'SLOPE='F10.5)
182 WRITE (6,117)
184 117 FORMAT (//1X,'ENTER DILUTION FACTOR')
186 READ (5,*) D
190 WRITE (6,115)
200 115 FORMAT (//1X, 'ENTER NUMBER OF DATA POINTS')
210 READ (5,*)N

```

```
220 WRITE (6,135)
230 135 FORMAT (//1X,'ENTER TOTAL THEREOTICAL AMOUNT OF DRUG IN MG')
240 READ (5,*)TD
250 WRITE (6,125)
260 125 FORMAT (//1X,'ENTER TIME, ABSORBANCE')
270 READ(5,*) (TIME(I), ABS (I), I=1,N)
280 WRITE (6,100) SLOPE
290 DO 20 J=1,N
300 CONC(J)=0.0
310 20 CONTINUE
320 TLCONC=0.0
330 CORRX=0.0
340 DO 25 I=1,N
350 CONCX=0.0
360 CONCX=(ABS(I)/SLOPE)*D
370 TLCONC=TLCONC+CORRX
380 CORRX=(CONCX*5.0)/TV
390 CONC(I)=CONCX+TLCONC
400 AMT(I)=(CONC(I)*TV)/1000.00
410 25 CONTINUE
420 WRITE(6,165)(TITLE(I), I=1,6)
430 165 FORMAT(//1X,6A4)
440 WRITE(6,190)
450 190 FORMAT(//10X,'TIME (MIN)', 4X,'AMT(MG)',5X,'PERCENT', 5X,'PERCENT')
460 WRITE(6,300)
470 300 FORMAT(//33X,'(ACTUAL AMT)',2X,'(CLAIMED AMT)')
480 DO 30 I=1,N
490 PERCNT=(AMT(I)/AMT(N))*100
500 CPRCNT=(ANT(I)/TD)*100.00
510 WRITE(6,250)(TIME(I),AMT(I),PERCNT,CPRCNT)
520 250 FORMAT(//1X,4(3X,F10.2))
530 30 CONTINUE
540 PD=(AMT(N)/TD)*100.00
550 WRITE(6,145)PD
560 145 FORMAT(//1X,'PERCENT OF DRUG IN TABLET=',F10.2)
570 STOP
580 END
```

Spectrophotometer Reading For  
Pyridoxine Hydrochloride (USP)

Concentration( $\mu\text{g}/\text{ml}$ )	Absorbance
14.50	0.623
7.25	0.304
3.63	0.205
1.81	0.153

Spectrophotometer Reading For  
Ascorbic Acid (USP)

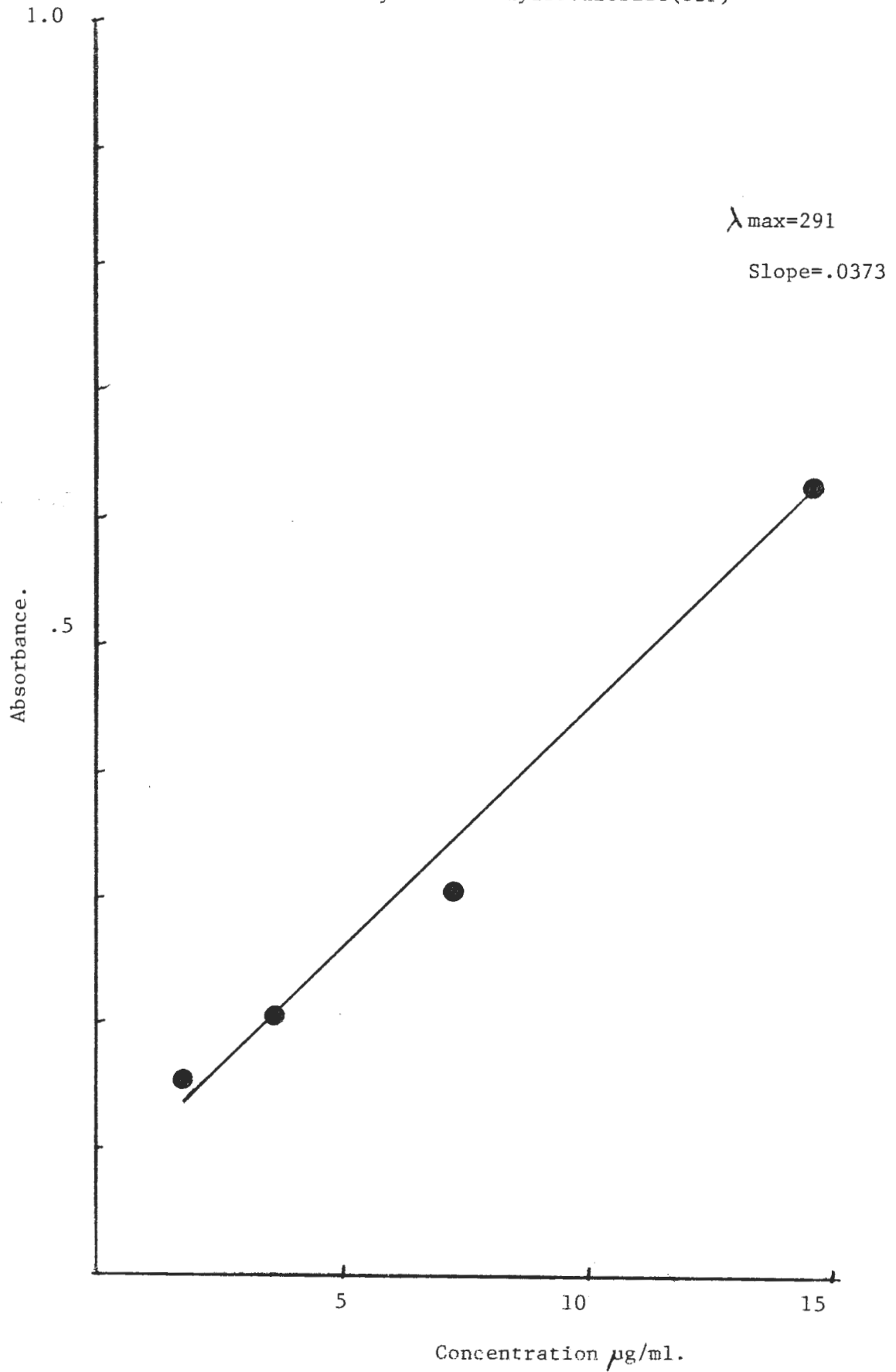
Concentration( $\mu\text{g}/\text{ml}$ )	Absorbance
14.10	0.833
9.04	0.567
7.05	0.386
5.64	0.303

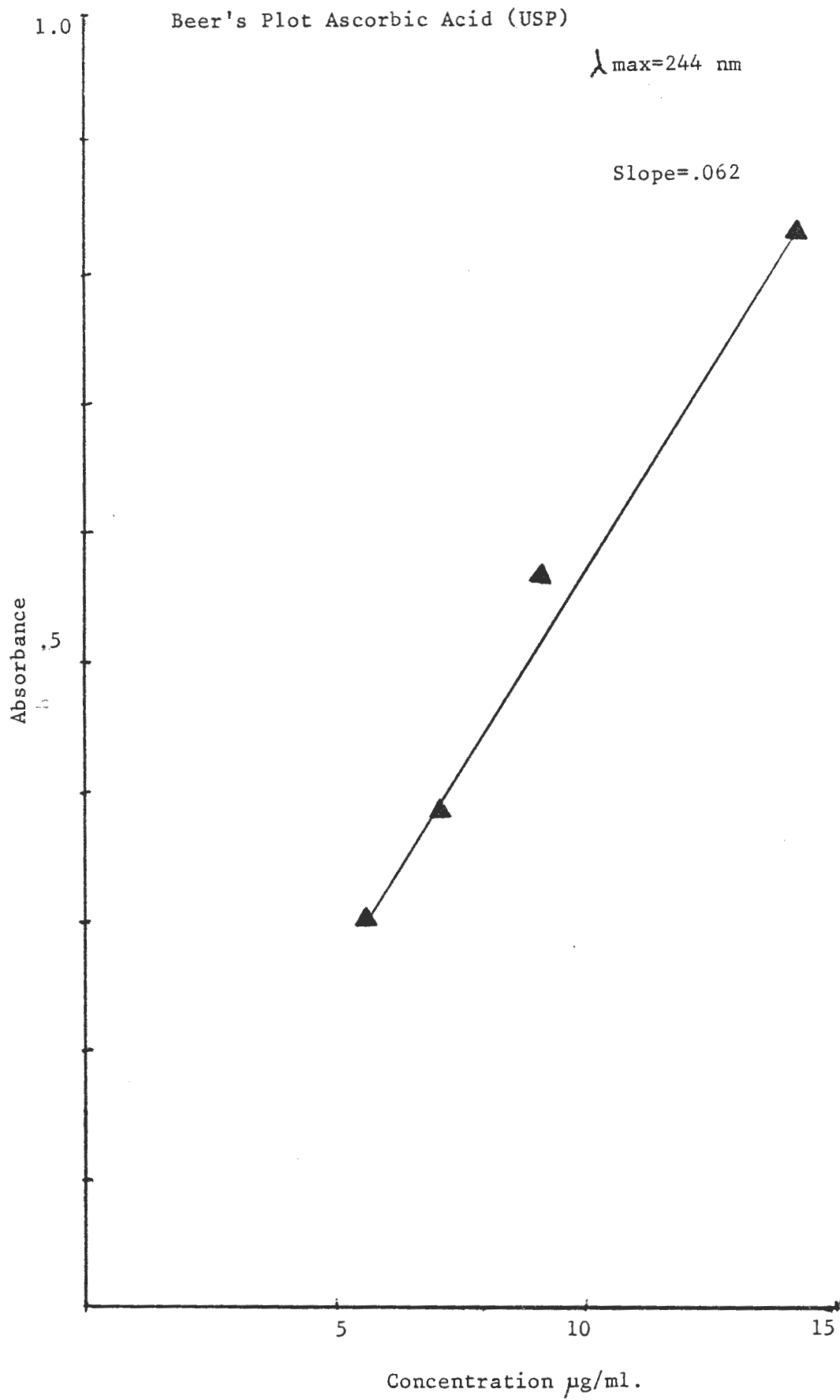
Spectrophotometer Reading For  
Granulated Ascorbic Acid (Roche)

Concentration( $\mu\text{g}/\text{ml}$ )	Absorbance
17.7	0.880
13.3	0.735
8.8	0.493
6.6	0.374



## Beer's Plot For Pyridoxine Hydrochloride(USP)





## Beer's Plot for Granulated Ascorbic Acid (Roche)

