Reproductive Biology and Natural History of the White-lipped Pit Viper (*Trimeresurus albolabris* Gray, 1842) in Hong Kong

Anne Devan-Song  
*University of Rhode Island*

Paolo Martelli

*See next page for additional authors*

Follow this and additional works at: https://digitalcommons.uri.edu/nrs_facpubs

Citation/Publisher Attribution
Available at: http://www.herpconbio.org/Volume_12/Issue_1/Devan-Song_etal_2017.pdf

This Article is brought to you for free and open access by the Natural Resources Science at DigitalCommons@URI. It has been accepted for inclusion in Natural Resources Science Faculty Publications by an authorized administrator of DigitalCommons@URI. For more information, please contact digitalcommons@etal.uri.edu.
**Reproductive Biology and Natural History of the White-lipped Pit Viper (Trimeresurus albolabris Gray, 1842) in Hong Kong**

**Anne Devan-Song**\(^1,3,4^\), **Paolo Martelli**\(^2\), and **Nancy E. Karraker**\(^1,3\)

1School of Biological Sciences, University of Hong Kong, Kadoorie Biological Sciences Building, Pokfulam, Hong Kong  
2Veterinary Department, Ocean Park Corporation, 180 Wong Chuk Hang Road, Aberdeen, Hong Kong  
3Current address: Department of Natural Resources Science, University of Rhode Island, 1 Greenhouse Road, Coastal Institute, Kingston, Rhode Island 02881, USA  
4Corresponding author, email: devansong@uri.edu

**Abstract.**—The White-lipped Pit Viper (Trimeresurus albolabris) is broadly distributed through South and Southeast Asia, and very little is known about the natural history of the species. In this study, we report on the ecology, demography, reproductive states, and growth rates of T. albolabris in Hong Kong, China. The sex ratio is even in Trimeresurus albolabris populations. This species displays sexual dimorphism from birth. Females attain larger sizes, and all males, including neonates, displaying a white supralabial stripe on the head and have longer relative tail lengths. Males attain sexual maturity at about 410 mm snout-vent length (SVL). Females reach sexual maturity at about 460 mm SVL, but only begin reproducing at about 520 mm SVL. Mating occurs between August to October, coinciding with the onset of spermatogenesis. Trimeresurus albolabris displays post-nuptial vitellogenesis, and long-term sperm storage in females likely occurs over the winter. Females gestate over the summer and parturition occurs in late July to August. Females reproduce once every two or more years, except for very large females that may reproduce annually. Brumation by females occurs within the first three weeks of December, but snakes will emerge to bask on sunny days. Understanding its demography and reproductive ecology can provide insights into the conservation and management of T. albolabris populations in the region and increase our understanding of sexual dimorphism, reproduction, and evolution of the cryptic Trimeresurus species complex.

**Key Words.**—brumation; demography; diet; growth rates; parasites; sexual dimorphism; sexual maturity; Viperidae

---

**INTRODUCTION**

The vast majority of our understanding of snake ecology originates from temperate species (Vitt 1983; Brown and Shine 2002), and there is still a paucity of ecological data on tropical snakes compared to their temperate counterparts (Böhml et al. 2013). This lack of data prevents many patterns in the ecology of tropical snake groups, such as comparative analyses of reproductive cycles, life-history attributes, or population dynamics from being identified or recognized (Almeida-Santos and Salomao 2002; Scartozzoni et al. 2009). South and Southeast Asia, for example, harbor a high diversity and endemicity of herpetofauna (Böhml et al. 2013; Das and van Dijk 2013; Ganesh et al. 2013), yet even snakes that are relatively easily encountered are not well-studied, such as members of the diverse Trimeresurus complex (Serpentes: Viperidae: Crotalinae). Significant research has been conducted on Trimeresurus species on the subjects of venom proteomics (Du et al. 2002; Rojnuckarim et al. 2006; Soogarun et al. 2008), toxicology and bite pathology (Chanhome et al. 2002; Chotenimitkhun and Rojnuckarin 2008; He et al. 2008), and phylogeny (Malhotra and Thorpe 2000; Giannasi et al. 2001; David et al. 2011); however, relatively little has been published on the natural history, demography, and reproductive biology of Trimeresurus.

Members of the genus Trimeresurus occupy a wide range of habitats and exhibit variability in habits, ecology, and reproductive strategy (Pope 1935; McDiarmid et al. 1999; Orlov et al. 2002). One of the widespread members of this genus is the White-lipped Pit Viper (Trimeresurus [Cryptelytrops] albolabris), also locally known in South China as the Bamboo Pit Viper. Trimeresurus albolabris is a small, sexually dimorphic, arboreal pit viper that ranges from India to Bangladesh, Indochina, southern China, Malaysia, and Indonesia (Orlov et al. 2002). In northern Vietnam, copulation by T. albolabris has been observed between late March to late May, a captive female was reported to...
store sperm for a year, and neonates are born between late July and early August with litter sizes of four to 14 (Orlov et al. 2002; Kamelin and Lukin 2003). However, given the lability in reproductive strategies in single species across a geographic range (Aldridge and Duvall 2002; Sasa et al. 2009; Siqueira et al. 2013) and that environmental factors correlate strongly with reproductive cycles (Tsai and Tu 2000, 2001; Sasa et al. 2009), it cannot be assumed that other populations throughout its range display the same reproductive cycle seen in North Vietnam.

In Hong Kong, *T. albolabris* is one of the most commonly encountered snakes (Karsen et al. 1998) and is by far the most prevalent cause of medically significant snakebites (Cockram et al. 1990; Hon et al. 2004; Shek et al. 2009). *Trimeresurus albolabris* is described as a habitat generalist, occurring in most terrestrial vegetation types at all elevations in Hong Kong (Karsen et al. 1998), but no quantitative information is available on ecological aspects of this species in south China. The objective of this study was to shed light on demographic parameters, extent of sexual dimorphism, and reproductive ecology of *T. albolabris* in Hong Kong. In addition, we documented observations on brumation, parasites, and diet.

### Materials and Methods

#### Study site

We conducted this study in Hong Kong (22°09’ to 22°37’N, 113°50’ to 114°30’E), a special autonomous region of China spanning 1,100 km² (Fig. 1). Hong Kong harbors a high-density urban area but also 23 national parks and expanses of rural land containing small traditional villages. Hong Kong occurs within the tropics and exhibits a seasonal monsoon climate of hot, humid summer with high precipitation from May to September and cool, dry, and mild winter from November to February (Dudgeon and Corlett 1994). We sampled snakes within active and abandoned agricultural land, secondary forest, shrubland, and grassland.

#### Field methods

We used visual encounter surveys to locate *T. albolabris* along 32.1 km of hiking trails at 10 sites within Hong Kong (Fig. 1). We conducted surveys from April to December 2012 and June to December 2013. In 2013, we surveyed each of the 10 sites three to nine times. However, we did not quantify survey efforts in 2012, and we captured only vipers > 50 g in body mass in that year. We captured all individuals encountered in the 2013 visual encounter surveys.
We also captured vipers encountered outside of timed surveys in 2013, but we did not include these individuals in the calculation of capture rates.

We anesthetized all vipers with Isoflurane, an inhalant anesthesia, for measurements described below, but we used only a subset of vipers for radio telemetry. We implanted snakes with SB-2 or BD-2 radio transmitters (Holohil Systems Ltd, Carp, Ontario, Canada) that weighed ≤ 6% of the body mass of an individual. We waited 2 d for telemetered vipers to recover from surgery and at least 3 h for non-telemetered snakes to recover from anesthesia. We released all vipers within 5 m of capture site. We relocated telemetered snakes two to three times a week, with a mixture of day and night tracking, and we recorded their locations with a global positioning system. Individuals that survived 14–20 weeks had their transmitters replaced with new ones. At the end of each year, we captured all surviving telemetered vipers, removed their transmitters, and released them within 5 m of location of last capture.

We measured body mass, snout-vent length (SVL), and tail length of anesthetized vipers. Injuries and truncated tails were noted. We determined sex by probing the cloaca to detect hemipenes. If present. We conducted ultrasound scans on the ovaries of gravid, telemetered individuals after obtaining visual indications of substantial weight loss. For gravid vipers, we calculated change in body condition from the period post-parturition until brumation. We defined onset of brumation as the range of dates between last locating a snake above ground and first locating it underground beneath leaf litter, or in a rock wall crevice, even if the snake was found to emerge from refugia to bask or shift brumation sites. If vipers were hiding in rocky crevices, we removed loose rocks to uncover vipers during brumation to determine depth of burrow or depth below ground.

Parasites and diet.—We obtained fecal samples from anesthetized vipers to identify fecal parasites occurring in *T. albolabris* found in Hong Kong. We used direct smears and fecal flotation to identify parasite ova under a light microscope. We retrieved coelomic, subcutaneous, and intra-organ parasites from vipers during necropsies if we encountered viper carcasses. If mammalian hairs were detected in the feces, we identified hair to family level using light microscopy. Hair identification was aided with mammal hair samples from specimens at the Lee Kong Chian Natural History Museum, National University of Singapore.

Reproduction.—We gently palpated anesthetized male snakes both posterior and anterior to the cloaca without everting hemipenes to determine if semen was present. We conducted ultrasound scans on the ovaries of anesthetized, telemetered female snakes to enumerate and measure follicles or embryos. For telemetered females that survived until December of either year, we obtained these measurements in June and July, as well as December. We estimated the date range of parturition of gravid, telemetered individuals after obtaining visual indications of substantial weight loss. We measured mass loss through parturition by weighing postpartum snakes in situ. We also documented any intraspecific interaction between telemetered and non-study vipers that appeared to be mating.

Brumation.—We recorded microhabitat (ground/ foliage/ canopy/ tree or rock wall crevice/ burrow) and height above ground with each relocation of telemetered snakes. We defined onset of brumation as the range of dates between last locating a snake above ground and first locating it underground beneath leaf litter, or in a rock wall crevice, even if the snake was found to emerge from refugia to bask or shift brumation sites. If vipers were hiding in rocky crevices, we removed loose rocks to uncover vipers during brumation to determine depth of burrow or depth below ground.

Statistical analyses.—We compared sex ratios of *T. albolabris* using Chi-square. To assess normality of datasets, we used the Shapiro-Wilk test. Because male and female SVL were not normally distributed (*W* = 0.875, *P* = 0.015), we compared means using a Mann-Whitney U test. We then used Levene’s test to determine if variance in scaled mass index of reproductive females was different from that of immature females, using the smallest gravid viper found (524 mm SVL) as an empirically determined threshold between the two groups, and removing eight female neonates from the analysis. We used a Johnson-Neyman procedure (D’Alonzo 2004) to identify the non-significant regions of male vs. female mean tail length differences across total lengths (i.e., at which SVL relative tail length in males and females overlapped). We chose this test as an alternative to Analysis of Covariance, as the latter is uninformative beyond unequal slopes, while the Johnson-Neyman procedure (D’Alonzo 2004) allows for comparison of differences along the entire length of both slopes. We removed individuals with injured or truncated tails from relative tail length analyses. We used linear regression to quantify the relationship between SVL and litter size. We tested significance at α = 0.05. For snakes that we captured and measured multiple times, we used only measurements from initial capture for analyses, with the exception of growth rates and follicle measurements. We used only snakes from 2013 in statistical analyses, but we included snakes from 2012 in growth and reproductive data. We conducted all statistical analyses in R v. 3.1.2 (R Development Core.
Team, Vienna, Austria) except for the Johnson-Neyman procedure, which we calculated by hand.

**Results**

During 204.5 person-hours of visual encounter surveys in 2013, we captured 90 vipers, resulting in a capture rate of 0.43 vipers per person-hour of survey effort. We recaptured only five of the 90 vipers. Of these 90 vipers, we did not obtain SVL and tail length for 18 individuals captured in June. We included these 18 vipers in capture rate and sex ratio analyses but not in any analyses involving SVL, tail length, or total length. We encountered an additional 19 vipers incidentally, and we took all measurements of these vipers and included them in analyses involving SVL, tail and total length, but not capture rates.

We radio-tracked 20 snakes, which included nine females in 2012 (three of which survived), seven females in 2013 (five of which survived), and four males in 2013 (two individuals survived). We lost two snakes mid-season, and causes of death were attributed mostly to predation and vehicle strikes. We observed no direct causes of mortality from anesthesia, transport, or surgery. Pathology was limited to localized sepsis at the incision site, while infected subcutaneous tissue and muscle became encapsulated and surrounded by healthy tissue.

*Trimeresurus albolabris* exhibited a sex ratio of 1.08 females to 1 male, which did not significantly deviate from an even ratio ($\chi^2 = 0.154$, df = 1, $P = 0.695$). All males exhibited a faint to prominent lateral white stripe on the head and body, whereas no females displayed such markings on the head. However, it was not uncommon to find females with a white to yellow lateral stripe on the body. Ventral scale color varied from white to bright yellow to light green or white in both males and females. The mean SVL of females (469.5 ± 22.7 [SD] mm; range, 203.0–810.0 mm) was significantly greater than the mean SVL of males (347.8 ± 17.8 mm; range, 180.0–512.0 mm; $U = 471$, $P < 0.001$). Variance of scaled mass index for non-reproductive female body condition was significantly lower from that of reproductive females ($F_{1,37} = 5.64$, $P = 0.023$, Fig. 2).

At all sizes, males had longer relative tail lengths (mean = 0.199 ± 0.002) than females (mean = 0.144 ± 0.002), including neonates (Fig. 3). The limit of non-significant values fell between -1.7 and 195.8 mm total length, which is smaller than the smallest neonate encountered (Fig. 3).

Adult males decreased in body condition over the active season, as did two females (Fig. 4). Three non-gravid females (455 mm, 580 mm and 642 mm in SVL) increased in body condition between parturition and brumation, and the larger two initiated vitellogenesis (Fig. 4). Only one female (710 mm SVL) that was gravid in the June/July period increased in body condition over the active season, and this same individual initiated vitellogenesis (Fig. 4).

**Reproduction.**—The smallest gravid female we encountered was 524 mm in SVL and 87 g in mass, and the largest gravid viper encountered was 710 mm in SVL and 245 g in mass. The smallest sexually mature female (determined by presence of ovarian follicles via ultrasonography) was 463 mm in SVL. Two of six
(33.3%) adult female vipers encountered during the gestation period (June-July) in 2013 were gravid, and two of four (50%) adult female vipers encountered during that same period in 2012 were gravid. Litter size was strongly positively related to SVL ($r^2 = 0.98$, $F_{1,3} = 145.9$, $df = 4$, $P = 0.001$, Fig. 5). The largest gravid viper gestated at least 20 embryos that overlapped within the body cavity, whereas smaller vipers had only five to six non-overlapping embryos.

Parturition occurred between 15 July and 9 August in both years. Due to the low probability of obtaining direct visual observations of individuals when tracking vipers in dense vegetation, the exact dates of parturition are unknown for all vipers. No post-parturition parental care was observed, and we did not observe any neonates in the vicinity of the female post-parturition. Females were emaciated following parturition and lost approximately 3 g in mass per embryo. A necropsy of a telemetered female snake that was killed by a vehicle a few weeks after parturition revealed depleted fat reserves. Another female died of apparent starvation three months after parturition.

Follicle size at the end of the active season was reduced to the size measured during the inactive season ($< 5$ mm) in vipers that had been gravid in the summer of that season, except for one individual that had enlarged follicles in December (Fig. 6). This individual, tracked June-December 2012, was a very large female with substantial body mass even post-parturition and gained body mass over the active season after giving birth. Small adults retained primary follicles throughout the study period (Fig. 6). Large adults that were not gravid in the summer and gained substantial mass over the active season (July to November) exhibited increased follicle size of as much as 7–15 mm by the end of the year, which corresponds to stage II–III vitellogenesis in the closely related *Trimeresurus (Viridovipera) stejnegeri* (Tsai and Tu 2001).

Over the course of 2 y, we observed five instances of adult males attending large females between 17 August and 15 October. In all observations, males were perched on the same branch approximately 10–50 cm away from the female with its head orientated toward the female. We did not observe any two individuals of this solitary species within such close proximity outside of the putative mating season, even on surveys where we captured a high number of vipers within a short length of forest trail. We did not observe copulation or any other interactions. On one of the five above-mentioned occasions, two adult males were attending one large female. We captured five adult males 412–512 mm in SVL between 5 September and 15 October 2013 that secreted semen when palpated. We captured one of them while attending a large female, and the other four were solitary when captured.

**Brumation.**—We identified brumation refugia as: (1) small crevices in the ground covered by leaves, logs, or other plant debris, (2) crevices inside low human-made rock walls, and (3) small mammal burrows in soil, or under loose stones, boulders, or tree roots. We found a few vipers as deep as 0.5 m below the surface. The onset of brumation was 6–18 December in 2012 and 4–12 December in 2013. In December of both years, vipers emerged to bask on warm, sunny days in low shrubs, on the ground, or occasionally in trees. Each viper was
not confined to one brumation site and shifted between brumation sites within a few meters from each other. We did find any communal brumation sites. Neither of the two surviving males brumated for the duration of this project and both were still active and above ground in low shrubs in December.

**Parasites and diet.**—We identified *Kalicephalus* sp., *Strongyloides* sp., *Capillaria* sp., *Oxyuris* sp., ascarids, and other hookworm ova in the feces of 21 vipers. We detected an unidentified coccidian parasite in fecal matter of several vipers. We found ascarid larvae buried subcutaneously and within the coelomic cavities of several vipers. In one instance, a necropsy we performed on a female revealed ascarid infection throughout the coelomic cavity, fat bodies, kidneys, and liver (Fig. 7A). We also detected loose subcutaneous parasites (Fig. 7B). This individual had been suffering from severe hepatitis with lesions around an inflamed, mottled, and pale liver, with numerous hepatic nodular masses with parasites inside the cysts (Fig. 7C and 7D). The cause of death was due to asphyxiation during regurgitation and not infection, although it appeared that the viper would have eventually succumbed to the infection. The fish and bird tapeworm *Ligula intestinalis*, not previously reported to infect *T. albolabris*, was also found subcutaneously and retrieved from an anesthetized viper.

The only predation event we saw was that of an individual *T. albolabris* eating a Chinese Gecko (*Gekko chinensis*). This individual was found with a food bolus much larger than the size of the gecko 7 d later, although this second prey item was not identified. Examination of fecal matter revealed Muridae hairs in several vipers and Soricidae hair in one viper.

**Discussion**

Because of a low recapture rate of 5.5%, population parameters could not be measured based on the mark-recapture data, and more exhaustive sampling efforts or different methods to sample vipers are necessary, if studies aim to quantify populations. Low detection and
recapture rates and difficulties measuring population parameters are not unusual in studies of snakes (Parker and Plummer 1987; Steen 2010). The ratio of male to female vipers showed that \textit{T. albolabris} populations exhibit an even sex ratio in Hong Kong. We captured too few neonates in 2013 to detect a difference in the sex ratio (5 F:11 M) of juveniles.

\textbf{Sexual dimorphism, sexual maturity, and growth.—} Based on our data, we determined that it is possible to determine the sex of \textit{T. albolabris} at all ages by any one of the following: males have a diagnostic white stripe on the head and females do not; male relative tail length is higher than mean female tail length at all total lengths of snakes; and presence of hemipenes is evident in all sizes of male snakes. Sexual dimorphism in relative tail length of juvenile and adult snakes is well documented (Klauber 1943; Shine et al. 1999), and in \textit{T. albolabris}, this is evident at the neonate stage, not just juvenile and adult stage. Based on our measurement and reproductive data, we estimated the sizes at sexual maturity for both males and females. Sexual maturity in males probably occurs at or before about 410 mm SVL, as the smallest male found retaining semen in our study was 412 mm. Female \textit{T. albolabris} are sexually mature at about 460 mm SVL, but probably do not start reproducing before about 520 mm SVL. This is supported by three lines of evidence: (1) the smallest sexually mature female measured 463 mm in SVL, but the smallest gravid viper found was 524 mm, (2) large variation in body condition in relation to SVL only appears in females after they have attained about 525 mm in SVL, and (3) high variation in body condition of telemetered female vipers appears to be related to reproductive state. Reproduction in sexually mature females is therefore probably only initiated when body reserves exceed a threshold value, as in other snakes (Bonnet et al. 2002).

\textbf{Reproduction.—} Based on our data, we constructed a probable reproductive cycle for \textit{T. albolabris} in Hong Kong (Fig. 8). Our study provides evidence of gestation in June (or earlier) to July, parturition in July to August, suggests that mating takes place in August to October coinciding with the presence of semen in males, and shows that onset of vitellogenesis in females occurs before December. These observations suggest a post-nuptial vitellogenesis (vitellogenesis following mating) and subsequent long-term sperm storage over winter, both of which are characteristics of temperate snakes (Schuett 1992; Aldridge and Duvall 2002; Shine 2003). We hypothesize that vitellogenesis extends into the following year based on post-nuptial vitellogenesis and long-term sperm storage, and we estimated that the ovulation period occurs between March to June based on gestation, vitellogenesis, and parturition dates (Fig. 8). However, as we did not conduct surveys or track snakes from January to May 2013, we are unable to rule out the possibility of two mating seasons of \textit{T. albolabris} in Hong Kong, as seen in some temperate pit vipers in \textit{Agkistrodon} and \textit{Crotalus} (Aldridge and Duvall 2002).

Along with ratios of non-gravid to gravid adult female vipers during gestation periods, follicle measurements
and growth rates suggest that females reproduce biennially or even less frequently. Biennial or less frequently reproducing snakes are unable to complete all events necessary for yearly reproduction within 12 mo (Bull and Shine 1979) and are more common in temperate than in tropical species. Females that were gravid in June and July reverted to primary follicles in December of that year, while large non-gravid females commenced vitellogenesis by December. However, in one case, a gravid female appeared to undergo vitellogenesis in the same year after parturition. Notably, this was the largest female tracked and was the second largest female captured in this study. Very large females may therefore have sufficient fat reserves and efficient postpartum body condition recovery to reproduce in consecutive years, as has been observed in captive *T. albolabris* (Kamelin and Lukin 2003). Reproductive output in *T. albolabris* may therefore be determined by body reserves, similar to a typical Capital Breeder, such as *Vipera aspis* (Bonnet et al. 2002).

Our results highlight lability in reproductive traits of the same species across its geographic range. *Trimeresurus albolabris* in northern Vietnam appears to mate in spring (Orlov et al. 2002), despite being capable of long-term sperm storage (Kamelin and Lukin 2003). In contrast, *T. albolabris* in Hong Kong probably mates at the end of summer in the year preceding ovulation and fertilization. Interestingly, the latitudes and climates of North Vietnam and Hong Kong are not starkly different, and variation in reproductive strategies even within the geographic range of a species is usually accompanied by distinctions in environmental factors (Aldridge and Duvall 2002; Sasa et al. 2009; Siqueira et al. 2013). *Trimeresurus albolabris* exhibits relatively high variability in reproductive traits within a small geographic zone with little climatic variation.

**Natural history.**—Brumation is an important life-history event in ectotherms living in seasonal climates and can be critical for survival during the winter (Gregory 1982). Communal brumation is common in many snake species (Hirth 1966; Rudolph et al. 2007; Gienger and Beck 2011). It is characteristic of species that inhabit areas with more severe winters (Shine 1979) and may also reflect scarcity of suitable refugia (Harvey and Weatherhead 2006). Conversely, solitary brumation, such as is seen in *Sistrurus c. catenatus* (Harvey and Weatherhead 2006), suggests refugia are
not limiting. Based on our field research, we speculate that refugia are not limiting for *T. albolabris*, in which individuals brumate in multiple refugia over the season. The use of shallow refugia (0–0.5 m below surface) by *T. albolabris* also indicates overwintering areas are relatively warmer in Hong Kong compared with those available to temperate species (Shine 1979) and facilitates the basking behavior that we observed on warmer days.

All fecal parasite genera that we discovered in *T. albolabris* have been previously documented in the species. *Kalicephalus laticaudae*, *Capillaria* sp. and *Oxyurus* sp. were found in the feces of captive *T. albolabris* in Thailand (Chaiyabutr and Chanhome 2002). However, we were unable to identify the coccidian parasite present. Subcutaneous ascarid larvae also appear to be important parasites as they were found in fairly high numbers in several individuals. Brief notes in the literature suggest frogs (such as *Polypedates* spp.) form the bulk of the diet of *T. albolabris* (Orlov et al. 2002; Kamelin and Lukin 2003). Pope (1935) documented *Calotes versicolor* (Agamidae) and rats as commonly found prey items, and small mammals may be important sources of prey especially for larger vipers. High abundance of geckos encountered during visual encounter surveys suggests that geckos may be a food source.

In this study, we have constructed the reproductive cycle of *T. albolabris* in south China. We also have demonstrated sexual differences in behavior and substantial sexual dimorphism, even at the neonate stage. Knowledge of the natural history, reproductive ecology, and demography of *T. albolabris* contributes to baseline ecological data of this poorly studied species, which is the major cause of snake bites in Hong Kong. This knowledge will be useful for the conservation and management of this venomous snake.

**Acknowledgments.**—We are grateful to Dave Willott for assistance with site selection and entry. We appreciate the use of equipment and space at Freshwater Conservation Laboratory at the University of Hong Kong, and from David Dudgeon and Lily Ng. Field assistance was provided by Mary-Ruth Low, Lee Tirrell, Ray So, Sam Yue, Katharine MacIntosh, Matthew Strimas-Mackey, Lily Glidden, Rebecca O’Brien, Annamarie Saenger, and Nicholas Jakubek. We thank Nimal Fernando, Lee Foo Khong, Wendy Chan, Geraldine Kwok, Jodi Sievwright, and Joey Heung at the Veterinary Department of Ocean Park Corporation for assistance with surgeries. We appreciate manuscript edits from Jason Kolbe, Liliana Gonzalez, Sinlan Poo, and Margarete Walden. We also thank Gary Ades, Paul Crow, and Yorkie Wong of Kadoorie Farm and Botanic Garden and Robert Ferguson, Sally Ferguson, Jack Ferguson, Kevin Caldwell, Elsa Caldwell, Dan Rosenberg, Stephen Loman, Will Sargent, and Richard Wright for assistance with site selection and permissions to conduct research on private property. We thank Stephen Goldberg for parasite identification. Research was approved by Animals Control of Experiments Ordinance 11-1083, Department of Health, Collection Permit 40, Agriculture Fisheries and Conservation Department, and Veterinary Department, Ocean Park Corporation. We are grateful for funding provided by the Research Grants Council of Hong Kong (Grant # 767109 and 765510M), USDA National Institute of Food and Agriculture - McIntire-Stennis Program (RI00MS-978-INT), and the College of the Environment and Life Sciences, University of Rhode Island.

**Literature Cited**

Devan-Song et al.—Reproductive biology of *Trimeresurus albolabris*.


alborabris) venom protein by LC/MS-MS. Journal of Biochemical and Molecular Toxicology 22:225–229.


Anne Devan-Song is a Research Associate in the Department of Natural Resources Science at the University of Rhode Island, Kingston, USA. She received her B.Sc. at the National University of Singapore, and her M.Sc. from the University of Rhode Island. Her graduate research has focused on issues of wildlife translocation and effects of human activities on pit viper ecology and movements in South China. Currently, Anne is completing a herpetofaunal inventory in southeastern Virginia, USA. (Photographed by Scott Buchanan).

Paolo Martelli graduated as a Veterinarian in 1991. He began his career as a zoo veterinarian in 1993 at the Singapore Zoo and Night Safari where he worked until 2004, leaving as Director (veterinary). Since 2005 he has worked at Ocean Park Hong Kong where he is Director of Veterinary Services. In 2003 he received his Certificate in Zoological Medicine (RCVS). Paolo is on the board of the Asian Society of Conservation Medicine and is a recognized Specialist for Conservation Medicine (ASCM). Paolo is vice-chair of the Veterinary Science thematic group of the IUCN-SSV-Crocodile Specialist Group and an ordinary member of the IUCN-SSC-Pangolin Specialist group and Wildlife Health Specialist group, and is a founding member of the Tomistoma Task Force. In the course of his work, he has also led or participated in field projects in South and South East Asia and Southern Africa. He has also assisted or consulted to institutions, projects, and individuals with a variety of medical, surgical, or husbandry needs in many countries. (Photographed by Karthiyani Krishnasamy).

Nancy Karraker, Ph.D., is an Associate Professor at University of Rhode Island, Kingston, USA, in the Department of Natural Resources Science, where she teaches Herpetology, Wetland Ecology, and a field course on Biodiversity Conservation in Indonesia. She is an Honorary Assistant Professor at the University of Hong Kong where she taught for five years. She received a B.Sc. in Wildlife Management, M.Sc. in Natural Resources, and M.A. in English (Teaching of Writing) from Humboldt State University, Arcata, California, USA, and a Ph.D. in Conservation Biology from State University of New York College of Environmental Science and Forestry, Syracuse, USA. Her research focuses on the ecology and conservation of amphibians, reptiles, and wetlands in Southeast Asia and the Northeastern U.S. (Photographed by Anne Devan-Song).