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## **Pseudocalanus Copepods in the Bering Sea: Species Identification Intraspecific Diversity and Biogeography**

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PSEUDOCALANUS COPEPODS IN THE BERING SEA:  
SPECIES IDENTIFICATION, INTRASPECIFIC DIVERSITY AND  
BIOGEOGRAPHY

BY  
JENNIFER BAILEY

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## ABSTRACT

The calanoid copepod genus, *Pseudocalanus*, is an important secondary producer and widespread throughout the northern hemisphere. While ecologically important, previous work with this genus has been limited due to difficulties in morphometric identification. Here, *Pseudocalanus* species composition and distribution were examined in the Bering Sea through sequencing of the mitochondrial CO1 gene of 642 individuals from 10 locations. Phylogenetic trees of inter- and intraspecific diversity demonstrated that in the spring of 2010 (March-June), four species of *Pseudocalanus* co-existed at various locations across the shelf. Species composition in the middle shelf (50-100 m depth) was significantly different from composition on offshore domain ( $\geq 200$  m depth). Correlation of species composition with salinity indicated that environmental factors influence distribution. Differences between the middle shelf and offshore populations extended to the intraspecific level in two of the four species. Although high haplotypic diversity was observed in all *Pseudocalanus* spp., a weak structure between the middle shelf domain and offshore stations was only seen in *P. acuspes* and *P. minutus*. This indicates that while there may currently or previously have been some barriers, gene flow is possible across the shelf. Evidence of mixed intraspecific and divided interspecific distributions, in addition to correlation with environmental conditions, highlighted the importance of a species-specific approach to future research of this genus and will increase accuracy in the modeling of an ecosystem-wide response to climate change.

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## PREFACE

This thesis is composed of one manuscript, “*Pseudocalanus* copepods in the Bering Sea: Species identification, intraspecific diversity and biogeography”, which is formatted to be submitted to the Journal of Plankton Research. The manuscript addresses the question of *Pseudocalanus* spp. composition and distribution within the SE Bering Sea ecosystem.

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MANUSCRIPT:

*Prepared for submission to the Journal of Plankton Research*

***Pseudocalanus* copepods in the Bering Sea:**

**Species identification, intraspecific diversity and biogeography**

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**Abstract:**

The calanoid copepod genus, *Pseudocalanus*, consists of seven distinct and ecologically important species that are widespread and commonly co-occurring throughout temperate to arctic regions of the Northern Hemisphere. Due to difficulties in morphometric differentiation between species, very few studies have investigated the meso-scale distribution of these copepods, much less the factors influencing those distributions. Here, inter- and intraspecific diversity of *Pseudocalanus* was examined in the southeastern Bering Sea. The mitochondrial CO1 sequences of 642 individuals from 10 stations across the Bering Sea shelf were determined during the spring of 2010 (March-June). Four distinct species (*P. acuspes*, *P. mimus*, *P. minutus*, and *P. newmani*) were identified with an average genetic divergence of  $14.7\% \pm 4\%$ . Species were not evenly distributed across the Bering Sea shelf. Stations located in the middle shelf domain (50 - 100 m depth) were dominated by *P. acuspes* and had higher total abundance of *Pseudocalanus* copepods than stations located in the offshore domain (>200 m depth), which were dominated by *P. minutus*. A multi-dimensional scaling plot (MDS) of relative species composition data showed two groupings, each of 40% similarity. Based on relative species composition, offshore stations were significantly different from middle shelf stations (ANOSIM  $p = 0.01$ ). Haplotypic diversity was high in all species and in combination with a negative Tajima's  $D$  could indicate population expansion. A weak genetic structure was found between the middle shelf and offshore stations in *P. minutus* and *P. acuspes*. This indicates that while there may currently or previously have been some barriers preventing mating and the exchange of genes, gene flow is possible across the shelf. Correlation of species

composition with salinity and depth of the chlorophyll maximum suggests that distribution of species across the Bering Sea shelf is a result of environmental selection rather than isolation. Differential distribution of the four species and correlation with environmental conditions highlighted the importance of a species-specific approach to future research of this genus and will assist in creating greater accuracy in the modeling of an ecosystem-wide response to climate change.

## Introduction:

Copepods belonging to the genus *Pseudocalanus* are one of the most numerically dominant zooplankton in temperate-boreal waters of the Northern Hemisphere (Corkett and McLaren, 1978). Perhaps because of their high abundance and wide distribution, they serve an important role in the marine food web as secondary producers, directly connecting the primary production of microalgae with higher trophic levels (Coyle and Pinchuk, 2002). Ecologically and economically important species of juvenile fish, such as sprat and herring, rely upon these copepods as a significant prey item (Corkett and McLaren, 1978; Hinrichs et al., 2002; Möllmann and Köster, 2002), and while small in size, they may be more productive in coastal waters than larger copepods (e.g., *Calanus* spp.) (Corkett and McLaren, 1978).

Despite their importance, difficulties distinguishing between species morphometrically have limited interspecific research, creating a gap in knowledge of factors affecting the abundance and distribution of individual species (Frost, 1989; Bucklin et al., 1998). Within the *Pseudocalanus* genus, seven species have been recorded throughout the Northern Hemisphere, *P. acuspes*, *P. elongatus*, *P. major*, *P. moultoni*, *P. mimus*, *P. minutus*, and *P. newmani* (Frost, 1989). As these species are commonly co-occurring (Corkett and McLaren, 1978; Frost, 1989), most studies have focused on the genus as a whole (e.g., Coyle et al., 2011; Coyle et al., 2008; Coyle and Pinchuk, 2002; Vidal, 1980), and physiologies are largely unaccounted for at the species level (Frost 1989). Problems with grouping *Pseudocalanus* spp. for analysis of an ecosystem arise if individual species respond to unique environmental parameters for growth and survival. For example, there are differences in

overwintering strategies among *Pseudocalanus* spp. (Corkett and McLaren, 1978; McLaren et al., 1989). Species that have a primarily Arctic distribution, like *P. minutus* and *P. acuspes*, store large amounts of lipids compared to those with temperate to sub-arctic distributions (McLaren et al., 1989). Thus, different species may have different nutritional values that could affect the diet of juvenile fish.

Although species are difficult to identify morphometrically, there are clear genetic differences at the mitochondrial cytochrome *c* oxidase I gene (mtCOI) (Bucklin et al., 1998; Bucklin et al., 2003). Interspecific divergence in the mtCOI is estimated to range from 10-20% and intraspecific divergence from 0.5-2% (Bucklin et al., 2003). To date, six of the seven morphologically recognized species of *Pseudocalanus* have had the mtCOI gene sequenced and found significant divergence appropriate for a species-level classification (Bucklin et al., 2003). These sequences are now available on the international database, GenBank (Holmborn et al., 2010; Bucklin et al., 2003).

With the use of the mtCOI gene, an increased resolution of distribution patterns of individual species has now been established for Georges Bank, the Baltic Sea, the English Channel, the Black Sea, Svalbard fjords, and Balsfjord, Norway (Bucklin et al., 1998; Bucklin et al., 2001; McGillicuddy and Bucklin, 2002; Unal et al., 2006; Holmborn et al., 2010; Aarbakke et al., 2011). Most of these ecosystems contain only one member of this genus. The exceptions, Georges Bank and the NE Atlantic, have two and three species of *Pseudocalanus* respectively. By contrast, there are four species of *Pseudocalanus* in the Bering Sea: the primarily arctic *P. acuspes* and *P. minutus*, and the temperate/sub-arctic *P. newmani* and *P. mimus* (Frost 1989).

This multi-species system is ideal for field-based studies of ecological differences among *Pseudocalanus* spp.

The southeastern shelf of the Bering Sea is >500 km wide and accounts for over half the fish and shellfish landings in the United States (National Marine Fisheries Service, <http://www.st.nmfs.noaa.gov/commercial-fisheries/fus/fus11/index>). The inter-annual variations in the abundance of *Pseudocalanus* copepods have been shown to be a key factor determining the success of economically important fisheries such as walleye pollock (*Theragra chalcogramma*) (Napp et al., 2000; Napp and Hunt, 2001). In years when temperatures above the thermocline are warmer, *Pseudocalanus* copepods become the primary, if not exclusive prey items for larval fish in the region (Coyle et al., 2008).

Although no fine-scale mapping of *Pseudocalanus* spp. has been recorded for the Bering Sea, variations in the zooplankton community across the shelf and between warm and cold years indicate that there may be well-defined spatial and interannual distribution patterns for these smaller calanoid copepods (Cooney and Coyle, 1982). Water depth and spring/summer front systems divide the Bering Sea shelf into three domains: the inner (0-50 m), middle (50-100 m) and outer domain (>100 m depth) (Coachman, 1986). A generally weak cross-shelf advection results in distinct communities within these domains (Coachman, 1986). Larger copepods are found in the outer domains and offshore, whereas smaller copepods typically have higher abundances in the inner and middle domains (Coachman, 1986; Cooney and Coyle, 1982). During “cold” years, larger copepods dominate the zooplankton biomass, while smaller copepods are more abundant during “warmer” years (Coyle and

Pinchuck, 2002, Napp et al., 2002, Coyle et al., 2011, Stabeno et al., 2012).

*Pseudocalanus* spp. have produced a rather mixed interannual signal with varied abundance patterns during “warm” and “cold” years (Coyle et al., 2008, Coyle et al., 2011). This confusion may be caused by the grouping of multiple species within this genus (Coyle et al., 2011).

In this study, we used the mtCO1 gene to determine the spatial distribution of *Pseudocalanus* spp. present in the SE Bering Sea during the spring of 2010 and investigated factors controlling their distribution. Variations in species composition were hypothesized to occur across the shelf. Due to the variety of environmental conditions existing in the Bering Sea, gene flow was examined by looking at intraspecific variation to determine if any barriers existed that might isolate groups of *Pseudocalanus* and prevent the exchange of genes between regions.

## **Methods:**

### ***Field Collection***

Zooplankton were collected at 10 stations from 2 cruises, the USCGC Polar Sea PSEA1001 and the R/V Thomas G. Thompson TN249, from across the SE Bering Sea shelf between March and June of 2010 (Table 1). Copepods were sampled with a 150 and 333  $\mu\text{m}$  mesh ring net deployed in vertical tows to a depth of 60 m or 1 meter from bottom (if depth <60m) at rates of 15 or 20  $\text{m}\cdot\text{min}^{-1}$ . Net tow duration was  $\leq 5$  min. Half of the tow contents was filtered through a 150 $\mu\text{m}$  sieve and rinsed into 113.4 g jars filled with 95% ethanol (Bucklin et al., 2001). Twelve to twenty-four



hours after collection, samples were filtered into fresh ethanol to limit the diluting effects of organism dehydration.

At each station, temperature, salinity, pressure, bottom depth, and fluorescence were collected using a CTD profiler (Seabird 19plus). Extracted chlorophyll *a* was measured at the surface and chlorophyll maximum using standard fluorometric techniques (Lomas et al. 2012), and ice coverage was recorded and later retrieved from the ship logs.

***Morphometrics, DNA extraction, amplification, and sequencing:***

Adult female *Pseudocalanus* were isolated at random from the zooplankton samples (72-100 station<sup>-1</sup>). Each copepod was rinsed in fresh 95% ethanol and photographed. The urosome length, width, depth, and the prosome length were measured from photographs with ImageJ (Abramoff et al., 2004). DNA was extracted using the 96 well plate DNeasy Tissue Kit (Qiagen Inc.) following manufacturer's instruction with an overnight lysis incubation.

Three gene loci (the nuclear small subunit 18S ribosomal DNA [rDNA], the mitochondrial cytochrome *c* oxidase subunit I [mtCOI], and the nuclear internal transcribed spacer regions [nITS, ITS1-5.8S-ITS2]) were amplified by polymerase chain reaction (PCR). All reactions were run in a 20 µl PCR consisting of 1X Bio-X-Act master mix (Bioline UK Ltd.), 0.5 µM of each forward and reverse primers (APPENDIX A), and approximately 100 ng of DNA template diluted with ultra-pure water in a thermocycler (Eppendorf Mastercycler).

The 18S rDNA was amplified using eukaryotic universal primers (Medlin et al., 1988) and thermocycling protocol of 94°C for 30 s, followed by 35 cycles of 94°C for 30 s, 60°C for 1 min, and 72°C for 2 min, followed by 72°C for 10 min. The mitochondrial COI gene was amplified using the forward primer L1384-COI (Machida et al., 2004) and the reverse primer HCO 2198 (Folmer et al., 1994), with thermocycling protocol of 94°C for 1 min, 35 cycles of 94°C for 30 s, 49°C for 30 s, and 72°C for 50 s, followed by 72°C for 5 min. Finally, the nITS region (ITS1-5.8S-ITS2) was amplified with the universal primers ITS-4 and ITS-5 (White et al., 1990) under the following thermocycling conditions: 95°C for 1 min, 35 cycles of 95°C for 30 s, 50°C for 30 s, and 72°C for 50 s, followed by 72°C for 7 min. PCR amplification was confirmed on a 1% agarose gel with SYBR Green (Applied Biosystems) staining and successful amplicons were purified with ethanol precipitation (APPENDIX B).

Sequencing for the 18S gene and nITS region was performed using the universal eukaryotic primers for single stranded sequencing (Medlin et al., 1988) and universal ITS4 and ITS5 primers (White et al., 1990) respectively. The mtCOI sequencing was performed in both directions for 62 copepods. The remaining copepods were sequenced only in the forward direction due to full gene coverage with the L1384-COI primer (Machida et al. 2004). All sequencing was performed at the Rhode Island Genomics and Sequencing Center with a 3130X1 (ABI Inc.) sequencer. The mtCOI gene was sequenced for a total of 642 copepods. Thirty copepods were sequenced to test divergence for the 18S gene, and the nITS region was sequenced for 40 copepods. Copepods were selected for nuclear sequencing to compare with unique sequence groups found with the mtCOI gene.

### ***Sequence Data Analysis:***

Sequences were trimmed and aligned with Clustal w in MegAlign (DNASTAR Lasergene 8). Phylogenetic relationships for all three loci were assessed with MEGA V.5 (Tamura et al., 2007) using a Kimura 2-parameter substitution model (Kimura, 1980) for neighbor-joining (NJ), and maximum likelihood (M-L) methods. Bootstrap values, UPGMA, NJ and ML trees were calculated with PHYLIP v. 3.69 (1000, 1000, and 100 pseudoreplicates respectively) (Felsenstein, 2009).

A phylogenetic tree of all unique mitochondrial CO1 sequences was created using FastTree (Price et al., 2009) as implemented in QIIME (Caporaso et al., 2010), and visualized with FigTree v1.3.1 (Rambaut, 2009). The tree included the six *Pseudocalanus* sequences available on GenBank (*P. acuspes* [HM770074], *P. minutus* [HM770076], *P. newmani* [AF332796], *P. mimus* [AF513651], *P. moultoni* [AF332795], and *P. elongatus* [HM770078]). The resulting tree was used to determine the species identity of each copepod sequence. Sequences that were <4% divergent were considered to be a species and were grouped with the corresponding GenBank sequence. Relative abundances of species were then calculated for each station by tallying mtCO1 groupings. Absolute species abundance was calculated for TN249 stations by multiplying relative abundance percentages by total adult female *Pseudocalanus* abundance counts recorded using a 25-cm diameter CalVet system (CalCOFI Vertical Egg Tow; Pers. Comm. A. Pinchuk, UAF; Smith et al. 1985).

DNA polymorphism was examined for the mtCO1 gene by calculating nucleotide diversity ( $\pi$ ), haplotype diversity ( $H$ ), and Tajima's  $D$  in DnaSP v5.10.01

(Librado and Rozas, 2009). Statistical significance of Tajima's  $D$  was tested by permutations with 1000 replicates. Population structure of individual species was determined across the shelf with a one-way analysis of molecular variance (AMOVA) and 1000 permutations in ARLEQUIN v3.5.1.3 (Excoffier and Lischer, 2010), where populations were defined as sampling stations and groups were defined as middle shelf domain (stations <100 m deep) and offshore domain (stations  $\geq$ 200 m deep) (Weir and Cockerham 1984, Excoffier et al. 1992, Weir 1996). Thus, for analysis using F-statistics,  $F_{ST}$  is defined as the genetic variation within a station relative to the total genetic variation,  $F_{SC}$  is the genetic variation within a station relative to the variation between the middle domain and offshore domain, and  $F_{CT}$  is the genetic variation between the middle domain and offshore domain relative to the total genetic variation.

### ***Morphometric Analysis***

Multivariate analysis of variance (MANOVA) and quadratic discrimination function analyses (QDFA) statistics were used with mtCO1 species assignments to test the effectiveness of size measurements in species identification. Measurements of urosome length, width, depth (EL, EW, ED), and prosome length (PL) were transformed (natural log) and analyzed as prediction variables. Wilks' Lambda was used as a test statistic to evaluate the MANOVA and QDFA analyses. Pillai's trace was additionally included given its reliability with relatively small sample sizes (Olson, 1976). Jackknife values were recorded from the QDFA to estimate variance. All morphometric analyses were performed with SYSTAT v.13 (Wilkinson, 1998).

## ***Environmental Correlation***

Resemblance matrices for both species abundance data (Bray-Curtis Similarity) and environmental data (Normalized Euclidean Distance for salinity, temperature, depth of the chlorophyll maximum, total chlorophyll, and dissolved oxygen) were created with PRIMER v6 software (Clarke and Ainsworth 1993, Clarke and Gorley 2006). Temperature, dissolved oxygen, chlorophyll and salinity were taken from the depth of the chlorophyll maximum where *Pseudocalanus* spp. may be feeding and the surface layer. Multidimensional Scaling (MDS) plots created with a Bray-Curtis Similarity matrix and overlaid clusters were created for station species composition to visualize station groupings. These groupings were tested using ANOSIM with all possible permutations (210). Abundance data and environmental patterns were compared with the global BEST test (Clarke and Gorley 2006). Within the BEST application, the BIO-ENV procedure was run using the Spearman coefficient (Kendall 1970) as a rank correlation method to search all possible combinations of variables. The BIO-ENV procedure was run for 999 permutations to test statistical significance of resulting correlations.

## **Results:**

### ***Interspecific Diversity***

Groupings in mtCO1 gene trees generated from UPGMA, Neighbor-Joining, and Maximum Likelihood methods for 493 bp sequences of 632 individuals indicated that four *Pseudocalanus* spp. were present in the Bering Sea: the temperate *P. newmani* and *P. mimus*, and the arctic *P. minutus* and *P. acuspes* (Figure 1). High

bootstrap values (>70%) confirmed species groupings. Divergence in the mtCO1 gene among the four *Pseudocalanus* spp. ranged from 10.7% between *P. acuspes* and *P. mimus* to 18.7% between *P. minutus* and *P. newmani*. The average overall divergence among all four species was  $14.7\% \pm 4\%$ .

Nuclear divergence was more limited in both the 18S gene and nITS region than in the mtCO1 gene. The 18S rDNA of 62 individuals representing all four mtCO1-defined species were identical along the 1566 bp sequence. Gene trees (UPGMA, NJ, and ML) from the 727 bp length sequences covering ITS1-5.8S-ITS2 of 33 individuals representing four species and divergent haplotypes showed limited divergence with no defined groups or agreement with the mtCO1 sequence groupings (Figure 2). The average overall divergence in the nITS region was  $1.3\% \pm 1.1\%$ .

### ***Morphometric Analysis***

Four hundred and eighty two individuals, with representatives from every station, were photographed and measured. Pillai's trace results showed a significant difference in size (prosoma and urosoma length, width and depth) between species for both MANOVA and QDFA models with  $p < 0.001$  for both models. The classification success of species based on these measurements was 68% for *Pseudocalanus acuspes*, 43% for *P. mimus*, 40% for *P. minutus*, and 25% for *P. newmani*. When the species data was combined, overall success of classification was 43%.

### ***Geographic Distribution***

Multiple species (2-4) coexisted at all ten stations in the Bering Sea (Figure 3). *P. minutus* dominated the offshore stations and *P. acuspes* dominated the middle domain stations. Limited numbers of *P. mimus* were found, but those sampled appeared primarily in the offshore regions. *P. newmani* was fairly well mixed among all stations. Absolute abundance of adult female *Pseudocalanus* for TN249 stations agreed with patterns observed in relative abundance data (Figure 4). Stations in the middle domain (50-100 m) had higher total abundance than stations in the offshore domain ( $\geq 200$  m depth). Due to differences in sampling methods between cruises, abundance data for the PSEA1001 cruise were not used in comparison between stations.

A multidimensional scaling plot (MDS) of relative species composition data showed two groupings of 40% similarity (Figure 5). One group consisted of stations all located on the middle shelf, in water less than 100 m deep. The other grouping consists of offshore stations ( $\geq 200$  m depth) and station 156 based on relative species composition. Offshore stations were significantly different from middle shelf stations (ANOSIM,  $p = 0.01$ ).

One anomaly found in spatial distribution was at station 156, where 2% of all sampled *Pseudocalanus* spp. were infected with the parasite *Hemiarthrus abdominalis*. This parasite ranged in size from 300-700  $\mu\text{m}$  in length and was found loosely attached to the dorsal side of live and preserved *Pseudocalanus* spp.

### ***Intraspecific Diversity***

A total of 137 unique mtCO1 haplotypes were identified from 632 individuals. Among these, 42 haplotypes were classified as haplotypes of *P. newmani*, 25 as *P. minutus*, 21 as *P. mimus*, and 40 as *P. acuspes*. Nucleotide and haplotype diversity varied among all four species with the two temperate species, *P. newmani* and *P. mimus*, having the highest diversity values (Table 2a). All Tajima's *D* values departed from the standard neutral model (Tajima, 1989). In *P. newmani* and *P. mimus*, Tajima's *D* was negative but not significant ( $0.10 > p > 0.05$ ). For *P. minutus* and *P. acuspes*, Tajima's *D* values were negative and significant ( $p < 0.05$ ), suggesting population expansion in these two species. Results from the AMOVA test showed a greater percentage of variation within populations than between groups, where populations are defined as stations and groups are defined as the middle domain and offshore (Table 2b). The fixation indices,  $F_{SC}$  and  $F_{ST}$ , were not significant for all four species ( $p > 0.05$ ), indicating that there is no genetic structure among populations within groups or among populations and among groups. The  $F_{CT}$  was small, but significant for *P. acuspes* and *P. minutus*. This indicates that a weak genetic structure exists across the shelf in arctic species.

### ***Correlation of Environmental Data with Species Distribution***

Environmental data was compiled for the TN249 and PSEA1001 cruises (Table 3). The BIO-ENV procedure showed that optimal correlation ( $\rho$ ) between environmental parameters and species composition resulted from a combination of depth of the chlorophyll maximum and surface salinity. This pairing of environmental



parameters had a statistically significant correlation with species distribution ( $\rho = 0.576$ ). The addition of other environmental variables including surface temperature and surface dissolved oxygen to this combination caused an immediate drop in correlation values (Table 4). Total chlorophyll (measured at the chlorophyll maximum) and % ice coverage were not in the top five correlation combinations. Temperature, salinity, and dissolved oxygen measured at the chlorophyll maximum depth did not register in the top 10 BIO-ENV combinations.

### **Discussion:**

Mitochondrial CO1 sequencing conducted across the shelf demonstrated that four *Pseudocalanus* spp., the arctic *P. acuspes* and *P. minutus*, and the temperate *P. mimus* and *P. newmani*, are present in the SE Bering Sea. This array of species matches the morphometric findings of Frost (1989) and is the greatest number of co-existing *Pseudocalanus* species in any region that has been confirmed with the use of genetic tools. Nuclear divergence of 0% in the 18S gene and  $1.3\% \pm 1.1\%$  in the nITS region suggests that species divergence may be more recent for *Pseudocalanus* spp. than other copepods. While the 18S gene is generally only useful in identifying between copepod genera (Bucklin et al. 2003), the divergence found in the nITS region of the *Pseudocalanus* was significantly lower than other copepods like *Acartia tonsa*, which have a divergence in the nITS region of 39% between clades (Chen and Hare, 2008). An average nITS divergence of  $1.3\% \pm 1.1\%$  for all *Pseudocalanus* spp. indicates this is not a useful region for species identification. The mitochondrial CO1 gene was the only marker successful in identifying species.

With the use of the mtCO1 gene, each station was shown to consist of 2-4 co-existing species. The distribution and abundance of these species varied widely across the shelf. There was a greater overall *Pseudocalanus* abundance in stations in the middle domain than in those beyond the offshore domain, which is consistent with the findings of previous studies that observed spring zooplankton distributions (e.g. Cooney and Coyle, 1982; Smith and Vidal, 1986). At the species level, *P. acuspes* and *P. minutus* were dominant but differed in their distribution across the shelf. *P. acuspes* was almost exclusively found in the middle domain, and although *P. minutus* was found at almost all stations, it makes up the largest portion of the species complex in the offshore stations. While *P. mimus* was also found primarily in offshore stations, the low number of individuals collected does not allow for definitive conclusions about their distribution. This species is thought to have a greater abundance on the shelf in the Gulf of Alaska based on detailed morphometric analyses (Frost, 1989; Napp et al., 2005) and may be present in the Bering Sea as a result of northward current flow across the shelf break. In contrast, *P. newmani* was fairly evenly distributed throughout the middle domain and offshore stations. Because this study only covered one spring season, it is unclear what seasonal and interannual variations of species presence and distribution patterns exist. The year of 2010 was classified as a “cold year” (Stabeno et al., 2012), which could explain the dominance of the primarily arctic species. These are the only two species that have been shown to store large amounts of lipids during the winter months (McLaren et al., 1989) and thus have been suggested to respond in a similar manner to larger calanoid copepods like

*Calanus marshallae/glacialis*, which increase in abundance during cooler years (Coyle et al., 2011).

The differences in species composition between the middle domain and offshore stations were statistically supported in similarity groupings. The one exception to the trends in both abundance and distribution was station T156, a middle domain station, which grouped at 40% similarity with offshore stations. Station P3, a station in the same area as T156 but sampled in March rather than June, was grouped with the middle domain stations. One potential explanation for this difference in grouping could be the effect of the parasite *Hemiarthrus abdominalis*, which was only found at station T156. While this parasite was only loosely attached to the *Pseudocalanus* urosome and did not appear to be feeding directly on the copepod, the size of the parasite ( $446\ \mu\text{m} \pm 72\ \mu\text{m}$ ) may have impacted swimming behavior and/or location in the water column, thus impacting feeding and survival. Any preference of *Pseudocalanus* host species could be a biotic cause of differences in species composition at this location.

Although biotic factors such as parasitic infection, predation and prey availability all have the potential to greatly impact species distribution, the influence of environmental abiotic factors such as temperature and salinity were the focus of this study. A correlation between species composition and the pairing of salinity and the depth of the chlorophyll maximum indicates that the environment may play a key role in determining distribution patterns. Though the relationship to the depth of the chlorophyll maximum is complex and most likely influenced by a combination of environmental conditions, salinity has been previously suggested to influence

zooplankton distribution in the Bering Sea. A study conducted by Smith and Vidal (1984) concluded that salinity and lack of advection across the shelf, not temperature, were the critical factors in zooplankton community structure. There are two possible hypotheses that could explain the link between salinity and *Pseudocalanus* spp. structure. The first is that the environment is selecting for certain species. Stations that had higher salinity (31-32.5 psu) were composed of greater percentages of *P. minutus* and *P. newmani*, while stations with lower salinity (30-31 psu), were dominated by *P. acuspes*. If the species have different critical salinity ranges, this could explain their differential distribution. In the Baltic Sea, a decreasing success in spring maturation and reproduction in *P. elongatus* has been linked to decreasing salinity from increased precipitation (Möllmann et al., 2003). A similar relationship may exist between salinity and the four species of the Bering Sea. An alternative hypothesis to explain this link is that salinity is an indicator of water mass, and species composition will be more similar in water masses that are linked by current flow. If this were the case, however, we would expect to see a similar correlation with temperature, which is not found. Although temperature and salinity are conservative at depth, they are subject to evaporation and air heat exchange at the surface where we observed the strongest correlation with species composition. It appears unlikely that the correlation with salinity is entirely related to the water mass. In order to more thoroughly test this hypothesis, data would need to be collected on the source populations of each of these species and how the currents distribute them across the shelf. This would require a species-specific look at developmental stages as well as populations outside of the Bering Sea. With either hypothesis, it does appear that

salinity is more closely correlated with species distribution than temperature and other abiotic factors.

The correlation found between salinity and species composition was not observed at the subspecies level. There was, however, a cross-shelf difference in the haplotypic distribution of the Arctic species, *P. minutus* and *P. acuspes*. A weak but significant genetic structure between the middle domain stations and the offshore stations demonstrated that there may currently or previously have been some environmental (e.g. salinity) or physical barriers to genetic exchange (e.g., Goetze, 2005; Chen and Hare, 2008; Chen and Hare, 2011; Aarbakke et al., 2011). In combination with negative Tajima's *D* values, which signify population expansion (Tajima, 1989), it is possible that this structure is a relict of the last glacial period when flow through the Bering Strait was extremely limited. In the last glacial cycle, the Strait was nearly closed at multiple times, with the last closure ~70kyr BP (Hu et al., 2010). One explanation for the weak structure and expanding population of the two Arctic species is that populations of these species were previously isolated, on either side of the Bering Strait, and have only recently (within the last 70kyr BP) been able to exchange genes. This may additionally be the cause of the lower haplotypic and nucleic diversity observed in the arctic relative to the temperate species. The temperate species, *P. newmani* and *P. mimus*, which do not have significant structure or Tajima's *D*, would not have been subject to this isolation given their already more southern distribution. Regardless of whether structure in the Bering Sea is a relict from the last glacial period or represents current conditions, it is clear that there is some genetic exchange within the ecosystem. Existing structure is weak to non-

existent in all four species and both haplotypic and nucleic diversity are high relative to *Pseudocalanus* spp. from other ecosystems. In the Baltic Sea, haplotypic diversity of *P. elongatus* was 0.024, dramatically lower than even that measured in *P. acuspes* ( $H = 0.436$ ). The Bering Sea is clearly a mixing zone for different populations of both arctic and temperate species of *Pseudocalanus*. The high haplotypic diversity could be a result of the mixing of distinct lineages from different ocean basins (Pacific/Atlantic) in this region and/or the recent exchange of genes following a period of isolation (e.g. Bering Strait closure).

With a better understanding of species distribution and environmental impacts, more accurate modeling may be able to identify the importance of individual species as a prey for larval fish. As the southeastern Bering Sea has previously demonstrated a rapid response to atmospheric perturbations (Napp and Hunt, 2001) and an increase in the frequency of these warm-water years is expected (Coyle and Pinchuck, 2002), studies indicate that *Pseudocalanus* copepods may become an even more important prey item in the Bering Sea ecosystem (Coyle et al., 2008; Coyle et al., 2011). Future research in the Bering Sea will be able to look at seasonal as well as interannual changes in the distribution patterns established in this study to better predict the impacts of changes in the climate. A better grasp of species-specific abundance and distribution will further assist in the accuracy of fisheries modeling. Pollock have a “higher energy density” in their first winter if they are able to consume mesoplankton with higher lipid levels in a previous cool summer (Coyle et al., 2011; Hunt et al., 2011). If *Pseudocalanus* spp. with lower lipid content (*P. mimus* and *P. newmani*) become the primary Bering Sea species during warm summers, Pollock and other

foraging fish species may be negatively impacted and have a lower survival rate the following winter. This study has established the species composition and distribution for a ‘cool’ spring season as well as possible environmental forcing factors and thus provides a baseline for future studies to investigate seasonal and temporal changes within the Bering Sea.

On a global scale, this research highlights the importance of investigating the role of *Pseudocalanus* spp. in an ecosystem, from a species-specific perspective. The Bering Sea is only one of several ecosystems that house multiple species within this genus. If salinity is an important determinant in habitat and niche partitioning, changes to the environment might cause a redistribution that would not be seen if the genus were investigated as a whole. Large shifts in the biogeographic distribution of zooplankton have already been observed since the 1960s as a result of climate change (e.g., Beaugrand et al. 2009). It is possible that shifts will occur within a genus allowing some species to move into new territories and others to retreat northward. Recent advances in molecular protocols have allowed for a more detailed investigation into the present and future of this widespread and ecologically important copepod genus.

Table 1: Summary of TN239 and PSEA1001 cruise stations, sampling, and DNA analysis.



Cruise	Station No.	Date Sampled	Latitude (°N)	Longitude (°W)	Water Depth (m)	Pseudocalanus Abundance (individuals · m <sup>-3</sup> )	Number of Copepods Analyzed	
							mtCOI	nITS
PSEA1001	3	13-Mar-10	62.04	175.20	80.0	-	65	-
PSEA1001	68	2-Apr-10	59.88	171.66	70.0	-	38	-
TN249	49	19-May-10	59.90	178.90	485.6	42.4	100	8
TN249	55	21-May-10	58.20	174.24	381.4	62.9	65	2
TN249	87	29-May-10	55.43	168.06	204.5	38.6	62	3
TN249	99	31-May-10	56.85	164.51	73.0	91.1	73	-
TN249	156	5-Jun-10	62.19	175.15	78.7	121.3	53	5
TN249	163	7-Jun-10	59.89	178.90	657.4	40.0	64	3
TN249	175	9-Jun-10	59.90	172.20	73.0	311.4	57	-
TN249	179	10-Jun-10	58.83	168.16	46.0	105.5	63	11

Figure 1: Neighbor-Joining tree of the 137 unique mtCO1 sequences generated from 642 individuals. Sequences of *Pseudocalanus* retrieved from GenBank were included in the tree for species classification. Bootstrap values for UPGMA, NJ and ML analyses are shown (UPGMA/NJ/ML). An asterisk “\*” indicates bootstrap values of 100%. A dash “-” indicates a value less than 50%. Abundances of haplotypes are denoted with symbols, a circle (•) represents 10 individuals, a diamond (♦) represents an additional 5+ individuals, and outline around the circle (⊙) marks the point of 100 individuals. Nodes without a symbol represent singletons.

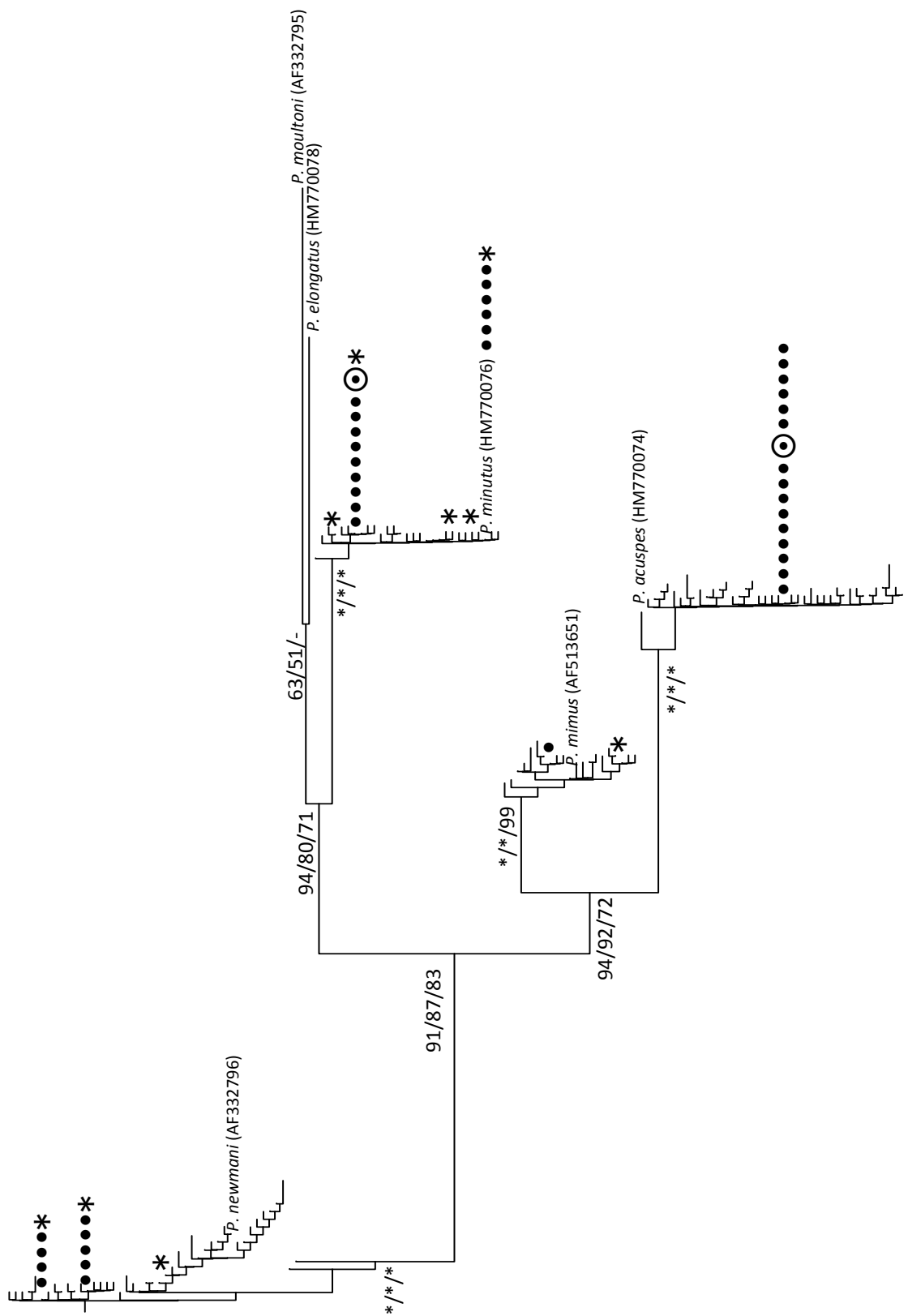


Figure 2: Neighbor-Joining tree of the 6 unique nITS ((ITS1-5.8S-ITS2) sequences generated from 40 individuals. The species name at the end of each node represents the mtCO1 defined species of the individual copepod that the unique nITS sequence was extracted from. The *Calanus pacificus* nITS sequence was included as an outgroup for tree rooting. Bootstrap values for UPGMA, NJ and ML analyses are shown (UPGMA/NJ/ML). An asterisk “\*” indicates bootstrap values of 100%.

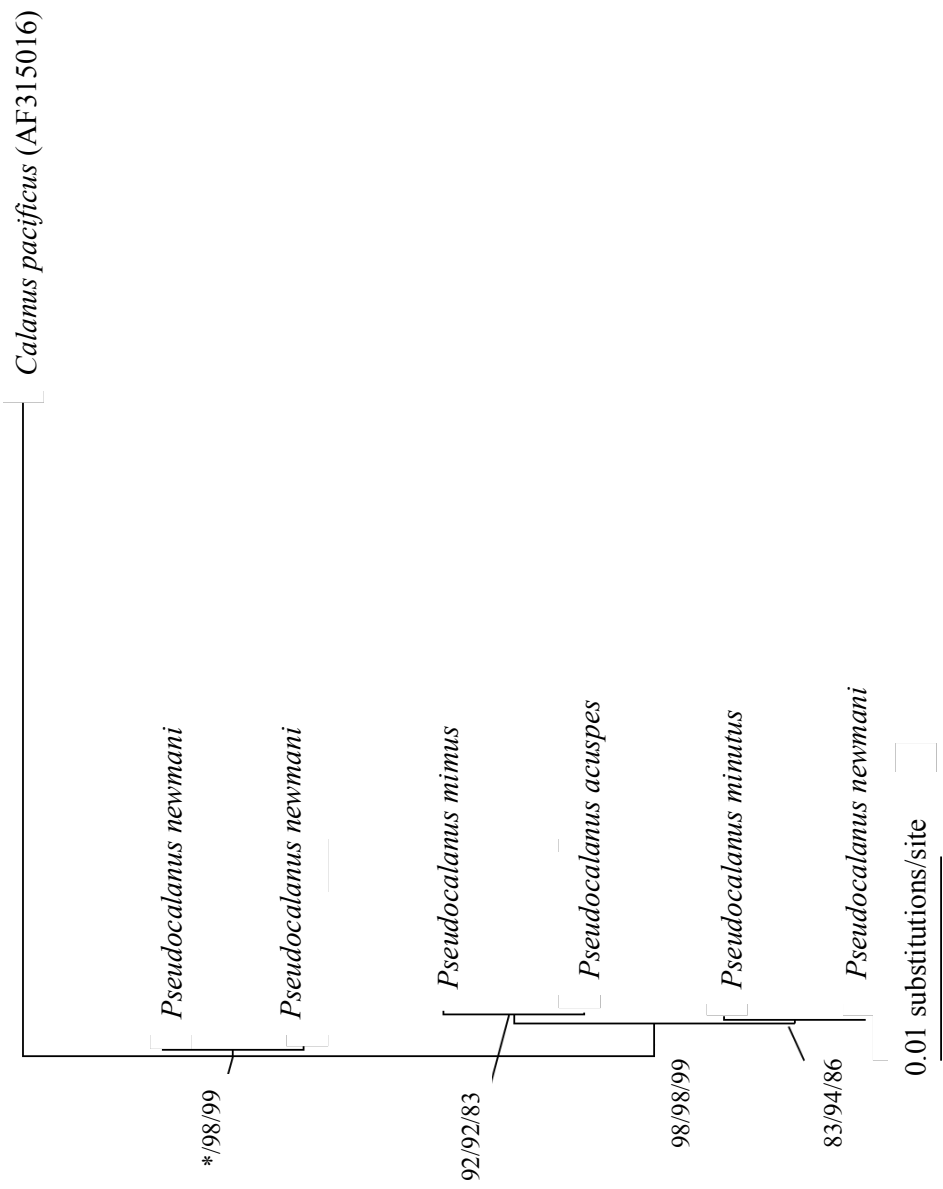


Figure 3: Geographic distribution of *Pseudocalanus* spp. in the Bering Sea based upon mtCO1 sequencing. The total number of individuals ( $n$ ) sequenced at each station is listed beneath each pie chart. Station names beginning with “T” are from the May/June 2010 TN249 cruise. Stations beginning with “P” are from the March/April 2010 PSEA1001 cruise. N values indicate the number of sequences used to create the corresponding pie charts.

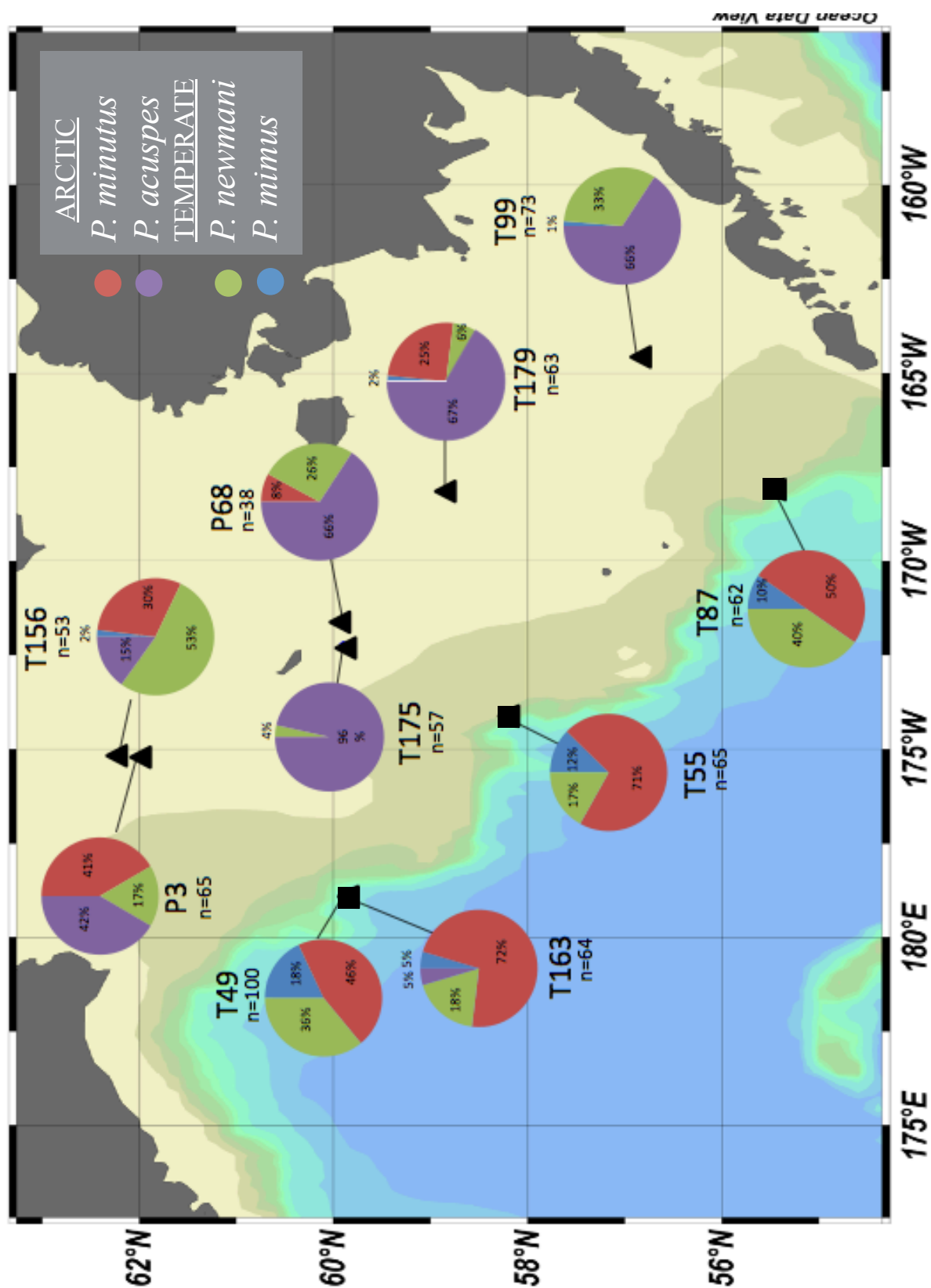


Figure 4: Abundance of *Pseudocalanus* spp. from TN249 stations. Stations are grouped by location on the shelf (Middle Domain and Outer Shelf/Slope Domain).



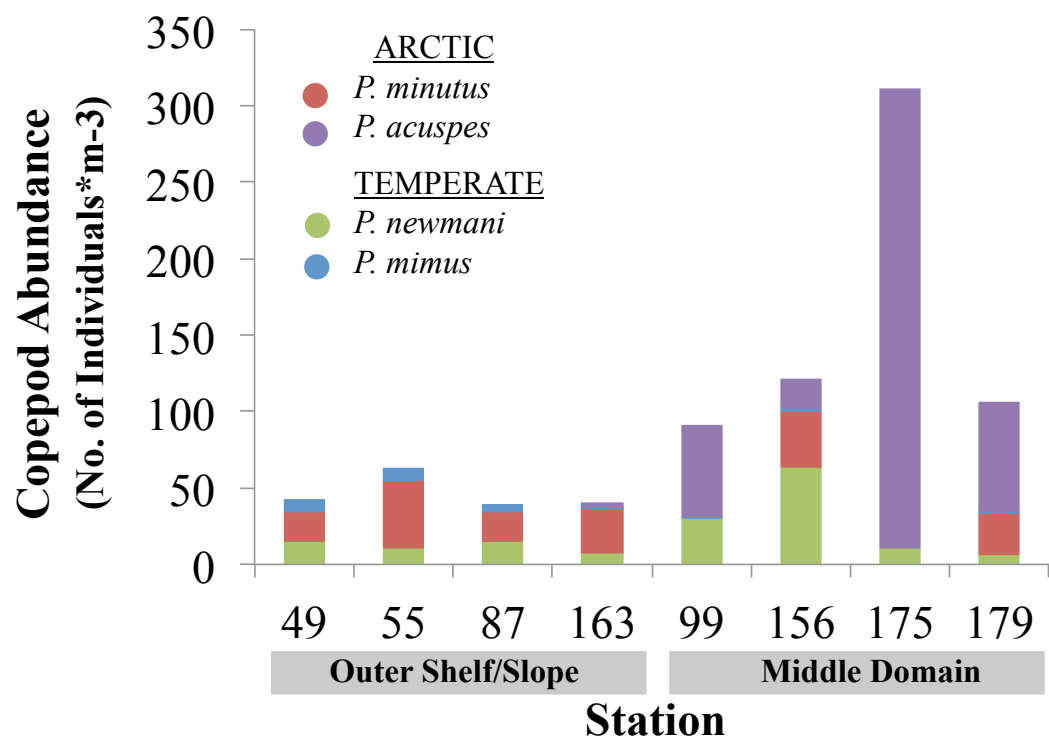


Figure 5: Multidimensional scaling plot showing similarity among stations based relative species composition. Stations in the middle domain (50 – 100m deep) are symbolized by triangles ( $\Delta$ ), and stations in the offshore domain ( $\geq 200$  m depth) are symbolized by squares ( $\square$ ). The results of Cluster Analysis are overlaid on the MDS plot as similarity circles showing stations that are 40% and 60% similar.

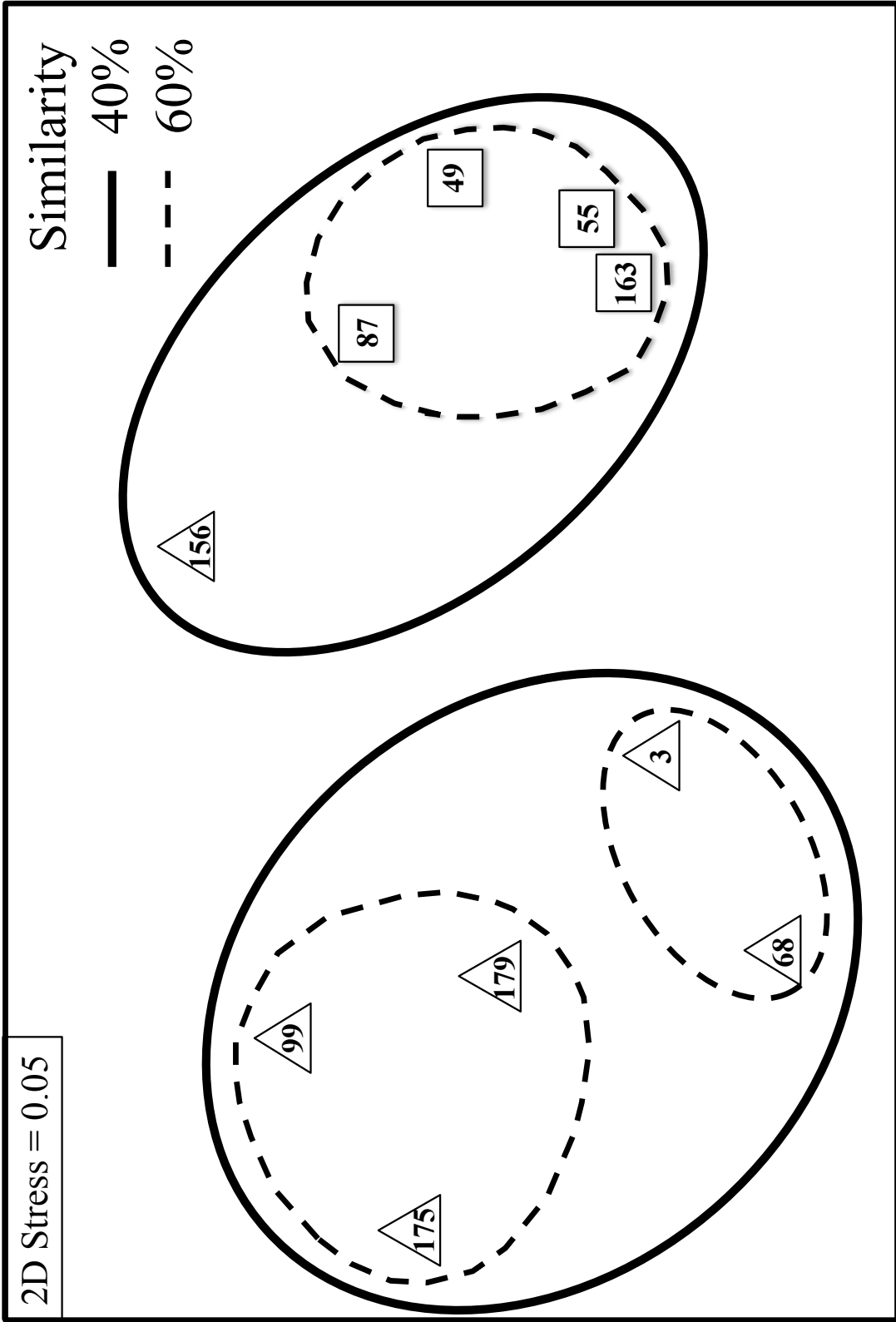


Table 2: Summary of intraspecific genetic data. a. Statistics of the mtCO1 gene of each of the four *Pseudocalanus* spp. Species are grouped by their dominant distribution region (i.e. Arctic and Temperate-Boreal). Statistical significance is listed below statistics values. b. AMOVA statistics investigating the genetic differentiation within stations, among stations, and between inner and middle shelf (50-100 m deep) vs. offshore stations ( $\geq 200$  m depth).

a.

Species	TEMPERATE		ARCTIC	
	<i>P. newmani</i>	<i>P. mimus</i>	<i>P. acuspes</i>	<i>P. minutus</i>
No. of Sequences	163	38	205	226
No. of Haplotypes	42	21	40	25
Nucleotide Diversity ( $\pi$ )	5.84E-03	7.85E-03	1.50E-03	2.01E-03
Standard Deviation	6.00E-04	6.10E-04	2.10E-04	1.30E-04
Haplotype Diversity ( $H$ )	0.786	0.915	0.436	0.670
Standard Deviation	0.024	0.034	0.045	0.023
Tajima's $D$	-1.57	-1.59	-2.59	-2.03
$P$ Value	0.10> $P$ >0.05	0.10> $P$ >0.05	$P$ <0.001	$P$ <0.05

b.

Species		<i>P. newmani</i>	<i>P. mimus</i>	<i>P. acuspes</i>	<i>P. minutus</i>
Percentage Variance	Within Populations	88.8%	99.1%	84.9%	92.0%
	Among Populations	15.5%	-3.75%	0.470%	-1.03%
	Between Groups	-4.28%	4.65%	14.7%	9.02%
Fixation Indices	FSC	0.150	-0.040	0.010	-0.010
	( $P$ -Value)	( $P=0.87$ )	( $P=0.87$ )	( $P=0.13$ )	( $P=0.13$ )
	FST	0.112	8.94E-3	0.151	0.073
	( $P$ -Value)	( $P=0.72$ )	( $P=0.72$ )	( $P=0.23$ )	( $P=0.23$ )
	FCT	-0.042	0.046	0.147	0.090
	( $P$ -Value)	( $P=0.06$ )	( $P=0.06$ )	( $P=0.00$ )	( $P=0.00$ )

Table 3: Summary of environmental data

Cruise	Station No.	Depth of Chl Max (m)	Surface Temperature (°C)	Surface Salinity (psu)	Total Chl (µg · L <sup>-1</sup> )	Ice Coverage (%)
TN249	49	1	0.03	31.85	23.8	0
	55	12	1.44	31.42	0.868	0
	87	11	4.09	32.28	5.47	0
	99	21	1.41	31.58	4.15	0
	156	32	2.34	31.76	4.29	0
	163	15	4.43	31.91	4.19	0
	175	37	2.03	30.03	0.993	0
	179	29	1.05	30.71	1.21	0
	3	0	-1.71	31.62	0.160	100
PSEA1001	68	0	-1.67	31.18	0.380	100

Table 4: The top 5 combinations of abiotic factors that most strongly correlate (Spearman's rank correlation coefficient) with the species composition patterns in the SE Bering Sea. An "X" designates which environmental factors were combined to achieve the corresponding correlation value. Abiotic values measured at the chlorophyll maximum depth are not included due to their absence in the BIO-ENV results.



Spearman Rank CORRELATION ( $\rho$ )	Temperature	Salinity	Dissolved Oxygen	Total Chlorophyll	Depth of Chl Max	% Ice Cover
0.576		X			X	
0.466	X	X			X	
0.344	X				X	
0.304			X		X	
0.302	X		X		X	

## LITERATURE CITED

- Aarbakke, O.N.S., Bucklin, A., Halsband, C. and Norrbin, F. (2011) Discovery of *Pseudocalanus moultoni* (Frost, 1989) in Northeast Atlantic waters based on mitochondrial CO1 sequence variation. *J. Plankton Res.*, **33**(10), 1487-1495.
- Abramoff, M.D., Magalhaes, P.J. and Ram, S.J. (2004) Image Processing with ImageJ. *Biophotonics International*, **11**(7), 36-42.
- Beaugrand, G., Luczak, C. and Edwards, M. (2009) Rapid biogeographical plankton shifts in the North Atlantic Ocean. *Gl. Ch. Biol.*, **15**, 1790-1803.
- Bucklin, A. Bentley, A.M. and Franzen, S.P. (1998) Distribution and relative abundance of *Pseudocalanus moultoni* and *P. newmani* (Copepoda: Calanoida) on Georges Bank using molecular identification of sibling species. *Mar. Biol.*, **132**, 97-106.
- Bucklin, A., Gaurnieri, M., McGillicuddy, D.J. and Hill, R.S. (2001) Spring evolution of *Pseudocalanus* spp. Abundance on Georges Bank based on molecular discrimination of *P. moultoni* and *P. newmani*. *Deep-Sea Res. II*, **48**, 589-608.
- Bucklin, A., Frost, B.W., Bradford-Grieve, J., Allen, L.D. and Copley, N.J. (2003) Molecular systematic and phylogenetic assessment of 34 calanoid copepod species of the Calanidae and Clausocalanidae. *Mar. Biol.*, **142**, 333-343.
- Caporaso J.G., Kuczynski J., Stombaugh J., Bittinger K., Bushman F.D., Costello E.K., Fierer N., Gonzalez Pena A., Goodrich J.K., Gordon J.I., Huttley G.A., Kelley S.T., Knights D., Koenig J.E., Ley R.E., Lozupone C.A., McDonald D., Muegge B.D., Pirrung M., Reeder J., Sevinsky J.R., Turnbaugh P.J., Walters W.A., Widmann J., Yatsunenko T., Zaneveld J. and Knight R. (2010) QIIME allows analysis of high-throughput community sequencing data. *Nature Methods*, **7**(5), 335-336.
- Chen, G. and Hare, M.P. (2008) Cryptic ecological diversification of a planktonic estuarine copepod, *Acartia tonsa*. *Mol. Ecol.*, **17**, 1451-1468.
- Chen, G. and Hare, M.P. (2011) Cryptic diversity and comparative phylogeography of the estuarine copepod *Acartia tonsa* on the US Atlantic coast. *Mol. Ecol.*, **20**, 2425-2441.
- Clarke, K.R., and Ainsworth, M. (1993) A method of linking multivariate community structure to environmental variables. *Mar. Ecol. Prog. Ser.*, **92**, 205-219.
- Clarke, K.R., and Gorley, R.N. (2006) PRIMER v6: User Manual/Tutorial. PRIMER-E, Plymouth.

- Coachman, L.K. (1986) Circulation, water masses, and fluxes on the southeastern Bering Sea shelf. *Cont. Shelf Res.*, **5**(1/2), 23-108.
- Cooney, R.T. and Coyle, K.O. (1982) Trophic Implications of Cross-Shelf Copepod Distributions in the Southeast Bering Sea. *Mar. Biol.*, **70**, 187-196.
- Corkett, C.J. and McLaren, I.A. (1978) The biology of *Pseudocalanus*. *Adv. Mar. Biol.*, **15**, 1-231.
- Coyle, K.O. and Pinchuk, A.I. (2002) Climate-related differences in zooplankton density and growth on the inner shelf of the southeaster Bering Sea. *Prog. Oceanogr.*, **55**, 177-194.
- Coyle, K.O., Pinchuk, A.I., Eisner, L.B. and Napp, J.M. (2008) Zooplankton species composition, abundance and biomass on the eastern Bering Sea shelf during summer: The potential role of water-column stability and nutrients in structuring the zooplankton community. *Deep-Sea Res. II*, **55**, 1775-1791.
- Coyle, K.O., Eisner, L.B., Mueter, F.J., Pinchuk, A.I., Janout, M.A., Ciciel, K.D., Farley, E.V. and Andrews, A.G. (2011) Climate change in the southeastern Bering Sea: impact on pollock stocks and implications for the oscillating control hypothesis. *Fish. Oceanogr.*, **20**(2), 139-156.
- Excoffier, L., Smouse, P.E., Quattro, J.M. (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics*, **131**, 479–491.
- Excoffier L. and Lischer H.E.L. (2010) Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Mol. Ecol. Res.*, **10**, 564–567.
- Felsenstein, J. (2009) (Phylogeny Inference Package) Version 3.6. Distributed by the Author. Department of Genome Sciences, University of Washington, Seattle.
- Folmer, O., Black, M., Hoen, W., Lutz, R. and Vrijenhoek, R. (1994) DNA primers for amplification of mitochondrial cytochrome *c* oxidase subunit I from diverse metazoan invertebrates. *Mol. Mar. Biol. Biotech.*, **3**, 294-299.
- Frost, B.W. (1989) A taxonomy of the marine copepod genus *Pseudocalanus*. *Can. J. Zool.*, **67**, 525-551.
- Goetze, E. (2005) Global population genetic structure and biogeography of the oceanic copepods *Eucalanus hyalinus* and *E. spinifer*. *Evolution*, **59**(11), 2378-2398.

- Hinrichsen, H.H., Mollman, C., Voss, R., Koster, F.W. and Kornilovs, G. (2002) Biophysical modeling of larval Baltic cod (*Gadus morhua*) growth and survival. *Can. J. Fish. Aquat. Sci.*, **59**(12), 1858-1873.
- Hu, A., Meehl, G.A., Otto-Bliesner, B.L., Waelbroeck, C., Han, W. Loutre, M.-F., Lambeck, K., Mitrovica, J.X. and Rosenbloom, N. (2010) Influence of Bering Strait flow and North Atlantic circulation on glacial sea-level changes. *Nat. Geosci.*, **3**, 118-121.
- Hunt Jr., G.L., Coyle, K.O., Eisner, L.B., Farley, E.V., Heintz, R.A., Mueter, F., Napp, J.M., Overland, J.E., Ressler, P.H., Salo, S. and Staben, P.J. (2011) Climate impacts on eastern Bering Sea foodwebs: a synthesis of new data and an assessment of the Oscillating Control Hypothesis. *ICES J. Mar. Sci.*, **68**(6), 1230-1243.
- Kendall, M.G. (1970) Rank Correlation Methods. Griffin, London.
- Kimura, M. 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J. Mol. Evol.*, **16**, 111-120.
- Librado, P. and Rozas, J. (2009) DnaSP v5: A software for comprehensive analysis of DNA polymorphism data. *Bioinformatics*, **25**, 1451-1452 | doi: 10.1093/bioinformatics/btp187.
- Lomas, M.W, Moran, S.B, Casey, J.R., Bell, D.W., Tiahlo, M., Whitefield, J., Kelly, R.P., Mathis, J.T. and Cokelet, E.D. (2012) Spatial and Seasonal variability of primary production on the Eastern Bering Sea shelf. *Deep-Sea Res. II*, **65-70**, 126-140.
- Machida, R.J., Miya, M.U., Yamauchi, M.M., Nishida, M. and Nishida, S. (2004) Organization of the Mitochondrial Genome of Antarctic Krill *Euphausia superba* (Crustacea: Malacostraca). *Mar. Biotech.*, **6**, 238-250.
- McGillicuddy, D.J. and Bucklin, A. (2002) Intermingling of two *Pseudocalanus* species on Georges Bank. *J. Mar. Res.*, **60**, 583-604.
- McLaren, A., Laberge, E., Corkett, C.J. and Sévigny, J.-M. (1989) Life cycles of four species of *Pseudocalanus* in Nova Scotia. *Can. J. Zool.*, **67**, 552-558.
- Medlin, L.K., H.J. Elwood, S. Stickel and M.L. Sogin. (1988) The characterization of enzymatically amplified eukaryotic 16S-like rRNA-coding regions. *Gene*, **71**, 491-499.

- Möllmann, C. and Köster, F.W. (2002) Population dynamics of calanoid copepods and the implications of their predation by clupeid fish in the Central Baltic Sea. *J. Plankton Res.*, **24**(10), 959-977.
- Möllmann, C., Kornilovs, G., Fetter, M., Köster, F.W. and Hinrichsen, H.-H. (2003) The marine copepod, *Pseudocalanus elongatus*, as a mediator between climate variability and fisheries in the Central Baltic Sea. *Fish. Ocean.*, **12**, 360-368.
- Napp, J.M., Kendall, A.W. Jr. and Schumacher, J.D. (2000) A synthesis of biological and physical processes affecting the feeding environment of larval walleye pollock (*Theragra chalcogramma*) in the eastern Bering Sea. *Fish. Oceanogr.*, **9**, 147-162.
- Napp, J.M. and Hunt, G.L. Jr. (2001) Anomalous conditions in the south-eastern Bering Sea 1997: linkages among climate, weather, ocean and biology. *Fish. Oceanogr.*, **10**, 61-68.
- Napp, J.M., Baier, C.T., Brodeur, R.D., Coyle, K.O., Shiga, N. and Mier, K. (2002) Interannual and decadal variability in zooplankton communities of the southeast Bering Sea shelf. *Deep-Sea Res. II*, **49**, 5991-6008.
- Napp, J.M., Hopcroft, R.R., Baier, C.T. and Clarke, C. (2005) Distribution and species-specific egg production of *Pseudocalanus* in the Gulf of Alaska. *J. Plank. Res.*, **27**, 415-426.
- Olson, C.L. (1976) On choosing a test statistic in multivariate analysis of variance. *Psychol. Bull.*, **83**, 579-586.
- Price M.N., Dehal P.S. and Arkin A.P. (2010) FastTree 2-Approximately Maximum-Likelihood Trees for Large Alignments. *Plos One*, **5**(3).
- Rambaut A (2009) FigTree v1.3.1: Tree Figure Drawing Tool. Available from <http://treebioedacuk/software/figtree/>. Accessed 2012 January 20.
- Smith, P.E., Flerx, W. and Hewitt, R.P. (1985) The CalCOFI vertical egg tow (CalVET) net. In Lasker, R. (Ed.), An Egg Production Method for Estimating Spawning Biomass of Pelagic Fish: Application to the northern anchovy *Engraulis Mordax*, pp. 22-33 *NOAA Technical Report NMFS 36*, U.S Department of Commerce.
- Smith, S.L. and Vidal, J. (1984) Spatial and temporal effects of salinity, temperature and chlorophyll on the communities of zooplankton in the southeastern Bering Sea. *J. Mar. Res.*, **42**, 221-257.

- Smith, S.L. and Vidal, J. (1986) Variations in the distribution, abundance, and development of copepods in the southeastern Bering Sea in 1980 and 1981. *Cont. Shelf Res.*, **5**, 215-239.
- Stabeno, P.J., Kachel, N.B., Moore, S.E., Napp, J.M., Sigler, M., Yamaguchi and A., Zerbini, A.N. (2012) Comparison of warm and cold years on the southeastern Bering Sea shelf and some implications for the ecosystem. *Deep-Sea Res. II*, **65-70**, 31-45.
- Steele, M., Ermold, W. and Zhang J. (2008) Arctic Ocean surface warming trends over the past 100 years. *Geophys. Res. Lett.*, **35**, doi:10.1029/2007GL031651.
- Tajima, F. (1989) Statistical Method for Testing the Neutral Mutation Hypothesis by DNA Polymorphism. *Genetics*, **123**, 585-595.
- Tamura, K. Dudley, J., Nei, M., and Kumar, S. (2007) *MEGA4*: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *Mol. Biol. Evol.*, **24**, 1596-1599. (Publication PDF at <http://www.kumarlab.net/publications>).
- Unal, E., Frost, B.W., Armbrust, V. and Kideys, A.E. (2006) Phylogeography of *Calanus hlegolandicus* and the Black Sea copepod *Calanus euxinus*, with note on *Pseudocalanus elongatus* (Copepoda, Calanoida). *Deep-Sea Res. II*, **53**, 1961-1975.
- Vidal, J. (1980) Physioecology of Zooplankton. II. Effects of Phytoplankton Concentration, Temperature, and Body Size on the Development and Molting Rates of *Calanus pacificus* and *Pseudocalanus* sp. *Mar. Biol.*, **56**, 135-146.
- Weir, B.S. (1996) Genetic Data Analysis II: Methods for Discrete Populaiton Genetic Data. Sinauer Assoc. Inc; Sunderland, MA, USA.
- Weir, B.S. and Cockerham, C.C. (1984) Estimating F-statistics for the analysis of population structure. *Evolution*, **38**, 1358–1370.
- White, T.J., Burns, T., Lee, S., and Taylor, J. (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: *PCR Protocols: A Guide to Methods and Applications* (eds Innis, M.A., Gelfand, D.H., Shinsky, J.J., White, T.J.), pp. 315-322. Academic Press, San Diego, California.
- Wilkinson, L. (1998) Systat 8.0: Statistics, SPSS, Chicago, Illinois, USA.

# APPENDIX A: Primer Sequences

PRIMER	TARGET REGION	SEQUENCE (5'-3')	SOURCE
EukA	18S rRNA (F)	AAC CTG GTT GAT CCT GCC AGT	Medlin et al., 1988
EukB	18S rRNA (R)	TGA TCC TTC TGC AGG TTC ACC TAC	Medlin et al., 1988
L1384-CO1	mtCO1 (F)	GGT CAT GTA ATC ATA AAG ATA TTG	Machida et al., 2004
HCO2198	mtCO1 (R)	TAA ACT TCA GGG TGA CCA AAA AAT CA	Folmer et al., 1994
ITS4	ITS1-5.8S-ITS2 (F)	TCC TCC GCT TAT TGA TAT GC	White et al., 1990
ITS5	ITS1-5.8S-ITS2 (R)	GGA AGT AAA AGT CGT AAC AAG G	White et al., 1990

## APPENDIX B:

### Ethanol Precipitation Protocol

1. Place 25  $\mu$ l of PCR reaction (30 – 5 for gel) in a 1.5 ml labeled microcentrifuge tube
2. Add 2.5  $\mu$ l 3M Sodium Acetate and vortex
3. Add 55  $\mu$ l ice cold pure (~95%) ethanol and vortex
4. Store on ice for 10 minutes, or if more convenient in the freezer for up to 12 hours
5. Centrifuge for 10 minutes at top speed (1350) in microcentrifuge with hinge facing out
6. Carefully decant supernatant, use a sterile toothpick to release surface tension and drain last droplets, tap upside down on paper towel
7. Add 500  $\mu$ l 70% ethanol and rock once gently
8. Centrifuge for an additional 5 minutes at top speed again hinges out
9. Remove supernatant as above
10. Leave open in the clean hood for 20 min to 45 minutes to evaporate last traces of ethanol, check by sniffing
11. Dissolve the DNA in 50  $\mu$ l pure H<sub>2</sub>O by running along the side by the hinge, vortex