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Cysteine-to-alanine replacements in the Escherichia coli SoxR protein and the role of the [2Fe-2S] centers in transcriptional activation

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ABSTRACT

The Escherichia coli soxRS regulon activates oxidative stress and antibiotic resistance genes in two transcriptional stages. SoxR protein becomes activated in cells exposed to excess superoxide or nitric oxide and then stimulates transcription of the soxS gene, whose product in turn activates ≥10 regulon promoters. Purified SoxR protein is a homodimer containing a pair of [2Fe-2S] centers essential for soxS transcription in vitro. The [2Fe-2S] centers are thought to be anchored by a C-terminal cluster of four cysteine residues in SoxR. Here we analyze mutant SoxR derivatives with individual cysteines replaced by alanine residues (Cys→Ala). The mutant proteins in cell-free extracts bound the soxS promoter with wild-type affinity, but upon purification lacked Fe or detectable transcriptional activity for soxS in vitro. Electron paramagnetic resonance measurements in vivo indicated that the Cys→Ala proteins lacked the [2Fe-2S] centers seen for wild-type SoxR. The Cys→Ala mutant proteins failed to activate soxS expression in vivo in response to paraquat, a superoxide-generating agent. However, when expressed to <5% of the cell protein, the Cys→Ala derivatives increased basal soxS transcription 2-4-fold. Overexpression of the Cys119→Ala mutant protein strongly interfered with soxS activation by wild-type SoxR in response to paraquat. These studies demonstrate the essential role of the [2Fe-2S] centers for SoxR activation in vivo; the data may also indicate oxidant-independent mechanisms of transcriptional activation by SoxR.

INTRODUCTION

Bacteria regulate many genes in response to imbalances in the production and disposal of reactive oxygen species; such conditions are often called ‘oxidative stress’ (1). In Escherichia coli, a set of <12 dispersed promoters under the control of the soxRS locus is activated when cells are exposed to sublethal levels of compounds, such as paraquat (PQ) (2, 3), which generate intracellular superoxide, or to nitric oxide (4, 5). This soxRS regulon controls antioxidant functions [e.g. superoxide dismutase or glucose-6-phosphate dehydrogenase (1)], repair of oxidative DNA damage [endonuclease IV (1)] and antibiotic resistance genes [micF (6) or acrAB (7)]. The soxRS system may also provide resistance to the toxicity of organic solvents and some heavy metals (8). The soxRS regulon is switched on in two transcriptional stages: existing SoxR protein is activated by an intracellular redox signal and triggers transcription of the soxS gene; the resulting SoxS protein binds and activates transcription from the various regulon promoters (1). SoxR protein is the master regulator of the soxRS response (9, 10). SoxR in vitro binds and strongly stimulates transcription of the soxS promoter by the exponential-phase RNA polymerase (RNAP) containing the 70S protein (11). The activity of SoxR as a transcription factor is completely dependent on the presence of non-heme iron in the protein (11). The metal is present in the active homodimeric SoxR protein as a pair of [2Fe-2S] clusters, which are in the oxidized form when the protein is isolated from cells (12, 13). SoxR activity can be regulated either by the assembly and disassembly of its iron–sulfur clusters (14, 15) or by oxidation–reduction of the [2Fe-2S] centers, with the oxidized protein being the transcriptionally active form in vitro (16, 17). Recent in vivo experiments support the interpretation that reduced SoxR is transcriptionally inactive (18).

From the foregoing it is clear that the [2Fe-2S] centers in SoxR are intimately involved in the protein’s function as a transcription regulator in vitro, but their in vivo importance has not been validated. Attempts to control iron availability are complicated by the essential roles of Fe in other proteins, and by the complex regulation governing the assimilation and storage of this metal (19). Initial spectroscopic analysis indicates that each [2Fe-2S] center in SoxR is anchored by four thiol ligands, which accounts for the four cysteine residues present in each protein monomer (12, 13). In this work we have tested the biological role of the [2Fe-2S] centers by mutating the individual cysteine residues of SoxR. These mutations eliminate the activation of SoxR by oxidative stress in vivo and prevent the assembly of stable iron–sulfur centers in the protein, but still allow a significant level of basal SoxR activity when the mutant proteins are expressed at high levels.

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MATERIALS AND METHODS

Strains and plasmids

This work employed the E. coli K-12 strains carrying single copy oUFon fusions present in lysogenized phage λ: strain TN521, a derivative of GC4468 (2) but ΔsoxRS::zjk2205-kan λ (soxR+ soxS2::lacZ) (20); and strain TN5311, as TN521 except λ (ΔsoxR soxS2::lacZ) (20) and containing F2 proAB lacFl lacZΔM15::TnJ0 (21), transferred from XLI-Blue (Stratagene, La Jolla, CA) by conjugation. The plasmid was from Stratagene and does not contain an inducible promoter for recombinant gene expression. The plasmid pSE380 contains the lacFl gene and the lac repressor-regulated trc promoter for controlled expression of recombinant genes (Invitrogen, San Diego, CA). The plasmid pKEN2 (a generous gift of G. Verdiene, Harvard University (11)) contains a lac-regulated derivative of the tac promoter for controlled expression of recombinant genes but does not contain its own lacFl gene; expression from pKEN2-derived plasmids is regulated by chromosomal lacI.

Construction of cysteine-to-alanine mutations

Four mutant soxR genes, each encoding an alanine residue substituting for a SoxR cysteine (residues 119, 122, 124 and 130), were constructed by using a plasmid containing the soxRS locus [pBD100 (22)] as a template for sequential rounds of polymerase chain reaction (PCR). For the first round of amplification, primers T3 (52-ATTAACCCCTCATAAAG-32) and T7 (52-AATACGACTCACTATAG-32), specific for sequences in the pBluescript KS vector, were used together with mutagenic primers converting the cysteine codon (TGT or TGC) to an alanine codon (GCT or GCC; underlined below).

a) C119A 52-GAACCTGAGCGGCTATTTGGGTGTTGG-32
b) C121A 52-CCACCAACCAATTCCCTGCTCCAGTTC-32
c) C122A 52-GATGTATTGCTGGCTGGCCGCTTT-32
d) C122A 52-AAAACGACCCACCACTATAC-32
e) C124A 52-TGGGCAAGGCCGCCAAGACCAAA-32
f) C124A 52-ATGGTTGTTTGCGCCCGCTTTTGCGGCA-32
g) C130A 52-CAGTGAAGCTCCGGCGTGCTT-32
h) C130A 52-TAGCAGACGGGGCGCTACGT-32

Overlapping PCR products were generated using the pairs of primers, pBD100 (10 ng) as the template, and U1 cloned Pfu DNA polymerase (Stratagene) and isolated from the reaction mixtures using a commercially available spin column (Qiagen, Chatsworth, CA). Approximately 10 ng of each pair of overlapping soxR fragments were then combined with 50 pmol of the T3 and T7 primers in a 100 μl reaction mixture. After denaturation and reannealing of the overlapping complementary cysteine coding regions, PCR yielded fragments (728 bp) containing the full-length mutated genes. These were purified and digested with EcoRI and HindIII (New England Biolabs, Beverley, MA), electrophoresed in a 1% agarose gel in Tris-acetate/EDTA buffer (23) and recovered from gel slices using DEAE membranes (Schleicher and Schuell, Keene, NH). The purified fragments were subcloned into EcoRI/HindIII-digested pBluescript and transformed into XLI-Blue cells. Transformants were selected on LB agar plates (21) containing 75 μg/ml ampicillin, 15 μg/ml tetracycline and the LacZ+ indicator 5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside (21). White colonies were selected and analyzed by PCR using the T3 and T7 primers to verify the presence of full-length soxR in the plasmids. Plasmid DNA was purified from positive clones and both strands of the soxR inserts completely sequenced by the dideoxy chain termination method (23) using Sequenase II (US Biochemical) and the following primers: T3, T7, SRK (52-GGGCAACACGCGCAAACGCT-32), SRKFP (52-AGC- GTTGGCGTTGGCC-32), E4 (52-GGGCGGATCCAGC- GGGAGATATAAA-32) and E5K (52-GGGCGGACGGCTGGTATGC- TCCAAAAAG-32).

The four mutant cysteine sequences were subcloned from the pBluescript vectors into pKEN2, generating the plasmids pKEN119A, pKEN122A, pKEN124A and pKEN130A. The C119A mutant gene was also subcloned into pSE380 to generate pSEC119A for subsequent studies. Despite several attempts, stable insertion of the other cysteine mutant genes into pSE380 was unsuccessful.

Preparation of whole cell homogenates and partially purified protein

Plasmids derived from pKEN2 were transformed into E. coli strain TN5311 and grown in 200 ml LB broth (21) containing 75 μg/ml ampicillin, 15 μg/ml streptomycin and 15 μg/ml kanamycin to OD600 = 0.5–1. Isopropyl-β-D-thiogalactopyranoside (IPTG) was then added to a final concentration of 0.5 mM, and the incubation was continued for 120 min at 37°C. The cells were harvested by centrifugation at 10,000 g at 4°C, resuspended and washed three times with ice-cold M9 salts (21). The final cell pellet was resuspended to a volume of 1.0 ml with 50 mM HEPES–NaOH, pH 7.6, 0.1 M NaCl, and lysed by agitation with glass beads (400 μl beads/ml) in a Mini-bead beater (Biospec Products, Bartlesville, OK) for 3 min. Following centrifugation at 10,000 g for 45 min at 4°C, the supernatants were collected and frozen at −20°C or −80°C until assay. Cell extracts were resolved on 15% SDS–polyacrylamide gels (24) and stained with Coomassie blue as previously described (11).

SoxR mutant proteins were also extensively purified using DE52–Sepharose and heparin–agarose column chromatography as described previously (11).

Iron and protein determinations

The Fe content of partially purified SoxR proteins (11) was determined by inductively coupled plasma emission spectrometry at the Chemical Analysis Laboratory, Institute of Ecology, University of Georgia (Athens, GA). SoxR concentrations were estimated by scanning densitometry analysis (Visage System, Millipore, Milford, MA) of Coomassie blue-stained SoxR in SDS–polyacrylamide gels (24) using a SoxR standard previously quantified by amino acid analysis.

DNA–protein binding

The binding affinity of wild-type and mutant SoxR proteins for the soxS promoter in vitro was analyzed by electrophoretic mobility shift assays, as previously described (9,11).

Preparation of cell suspensions

TN5311 transformed with the expression plasmid pKEN2 or its soxR-containing derivatives pK0XR [containing the wild-type soxR gene; (22)], pKENC119A, pKENC122A, pKENC124A or pKENC130A were grown overnight in LB broth containing ampicillin (100 μg/ml). A fresh 125-ml aliquot of the same
medium was incubated at 37°C with shaking at 250 r.p.m. for 110 min. IPTG was added to the final concentration of 0.5 mM, and the incubation continued at 37°C with shaking for an additional 120 min. The cells were harvested by centrifugation and resuspended in 0.5 ml of 50 mM HEPES–NaOH, pH 7.6, 0.1 M NaCl. Freshly dissolved dithionite, 0.1 M dithionite in 1 M HEPES–NaOH pH 7.6, was then added to the cell paste to a final concentration of 1 mM. Aliquots (300–400 µl cell paste) were immediately placed inside 4 mm EPR sample tubes, frozen in liquid nitrogen and kept at −80°C until analysis. The total SoxR concentration in the cells was determined by Western blot analysis of samples lysed directly in sample buffer (24), using previously quantified SoxR as a standard.

**Immunological methods**

Wild-type *E.coli* SoxR protein was purified to near homogeneity as described previously (11). The purified protein was emulsified in complete Freund’s adjuvant and injected subcutaneously into two New Zealand black rabbits. Starting three weeks after the primary injection, the rabbits received booster injections every 2 weeks with similar SoxR preparations in incomplete Freund’s adjuvant. Polycononal antisera were extracted periodically starting 4 weeks after the first injection, and SoxR-specific antibodies were enriched by affinity chromatography using SoxR-columns generated by coupling purified SoxR protein to HiTrap NHS-activated columns (Pharmacia) according to the conditions recommended by the supplier. Polycononal serum was applied to the columns and SoxR-specific antibodies eluted following standard procedures (25).

For immunoblotting, samples of cell suspensions or purified SoxR were electrophoresed in SDS–polyacrylamide gels, and transferred to nitrocellulose membranes (Schleicher and Schuell, Keene, NH) with a TE series Transphor electrophoresis unit ( Hoefer Scientific, San Francisco, CA). The filters were probed with the affinity-purified antiserum and bound antibody was detected with alkaline phosphatase-conjugated anti-rabbit IgG antibodies (Promega, Madison, WI).

**EPR spectroscopy**

X-band EPR spectra were recorded at 20 K on a Bruker model ESP300 spectrometer maintained at constant temperature, with an Oxford Instruments model ESR910 continuous flow cryostat as described previously (14,15). The amount of reduced SoxR was determined by comparison in the same experiment to standardized Fe-SoxR samples after reduction with dithionite (13,14). The high EPR background noise of the cell paste was greatly reduced by electronically subtracting from the spectra for SoxR-containing samples the spectrum of TN5311-pKEN2 cells, which do not express SoxR.

**In vitro transcription**

The activity of wild-type and mutant SoxR proteins was determined by *in vitro* transcription of the *soxS* gene by commercial *E.coli* RNA polymerase accompanied by the indicated amounts of SoxR, as described previously (13,14).

**Galactosidase assays**

The ability of the mutant proteins to stimulate *soxS* transcription *in vivo* was assessed using TN521 and TN5311. TN5311 cells were transformed either with the pBluescript-based plasmids for low-level SoxR expression, or with the pKEN2-based plasmids for high-level expression. TN521 cells were also transformed with the pSEC119A plasmid. Overnight cultures of the indicated strains were diluted 1:100 (or in some experiments 1:1000) into fresh LB broth containing 50 µg/ml ampicillin and treated as follows. For TN5311 containing pBluescript-based plasmids, PQ was added to a final concentration of 100 µM to one aliquot at OD600 of 0.4, and incubation at 37°C continued 60 min. For TN5311 containing pKEN2-based plasmids, when the cell density approached OD600 of 0.1, IPTG was added to a final concentration of 0.5 mM and after 60 min PQ was added as above. For TN5311 containing pSE380 or pSEC119A, IPTG was added (to 0.5 mM) at OD600 of 0.05. Aliquots of the cultures were removed at 0, 30 or 60 min following IPTG addition, PQ added (final concentration 250 µM) and the incubation continued 30 min before harvesting for the assay. β-Galactosidase activity was assayed in SDS/CHCl₃-treated cells as described by Miller (21).

**RESULTS**

Both strands of the four *soxR* Cys→Ala mutant genes were sequenced to verify the presence of the mutant codons. Each of the four mutant alleles contained the desired mutations (TGT→GCT or TGC→GCC) (Fig. 1) that reprogrammed the cysteine codon to an alanine codon. The full-length sequences of the four mutant constructs showed that no additional mutations were introduced by the site-specific procedure.

**In vitro properties of SoxR Cys→Ala mutant proteins**

SoxR proteins for *in vitro* characterization were overexpressed in TN5311 cells (OsoxRS) transformed with pKEN2-derived
mutants lacked the near-axial resonance spectrum characteristic of reduced wild-type Fe-SoxR (12–15), while such a spectrum was easily seen in cells overexpressing the wild-type protein (Fig. 3).

For the wild-type protein overexpressed in vivo in the absence of PQ treatment, 36% was detected as the reduced Fe-SoxR form, consistent with other experiments performed in a similar manner (18).

From the foregoing results, there was no indication that the mutant proteins contained [2Fe-2S] centers in vivo, although the unlikely possibility remains that such centers are present but remain in an EPR-silent (oxidized?) state. In vitro transcription assays (14) showed that the Cys→Ala mutant proteins were devoid of transcriptional activity for the soxS gene, and behaved similarly to apo-SoxR (Fig. 4). As found for the crude extracts (Fig. 2B), binding activity of the purified mutant proteins for the soxS promoter was indistinguishable from that of wild-type SoxR (data not shown). Thus, the Cys→Ala mutant proteins neither have tightly bound [2Fe-2S] centers nor function as specific transcription factors for the soxS gene in vitro.

In vivo properties of the Cys→Ala mutant proteins

In view of the lack of transcriptional activity of the mutated SoxR proteins in vivo, we wished to determine whether the Cys→Ala mutant proteins retained ability to induce regulated expression of the soxS gene in vivo. For this purpose we used a CsoR− strain harboring a single copy of the soxS::lacZ fusion (TN5311), in which expression of β-galactosidase activity encoded by the fusion depends on the in vivo activity of SoxR expressed in trans (20,26). The in vivo effectiveness of the mutant SoxR proteins was initially examined using constructs in the pBluescript vector, which lacks a strong inducible promoter and expresses wild-type SoxR at a level similar to cells containing a single soxR allele in the genome (E.H. and B.D., unpublished data). All four mutant proteins failed to increase β-galactosidase activity above the basal level observed with the vector alone, even when the cells were grown in the presence of PQ, which strongly activated soxS expression in cells with wild-type SoxR (<14-fold; Fig. 5A).

The results were somewhat different when intracellular SoxR concentrations were elevated (to <5% of the total cell protein) using the high level expression vector pKENC122A (11,13) (see Fig. 2A for SoxR protein levels). Wild-type SoxR (expressed from this vector) yielded <19-fold soxS activation in response to PQ (Fig. 5B), giving an induced level of β-galactosidase expression nearly twice that seen for cells bearing the wild-type pBluescript construct (Fig. 5A). Thus, the normal intracellular concentration of SoxR may be limiting for soxS activation in some situations. In contrast, cells overexpressing the mutant proteins from the pKENC2 vector exhibited elevated soxS expression that was not affected by PQ (Fig. 5B). For three of the mutant proteins (C119A, C122A and C124A) the level of soxS::lacZ expression was <4-fold higher than the basal (uninduced) expression mediated by wild-type SoxR; for the C130A protein, basal expression was <2-fold higher (Fig. 5B). The Cys→Ala mutant proteins have therefore lost responsiveness to the redox signal generated by PQ, but can exert a higher basal transcriptional activity than wild-type SoxR.

It seemed possible that expression of the mutant proteins might interfere with wild-type SoxR, either through the formation of heterodimers or by competition for binding the soxS promoter. At low levels of mutant SoxR expression (from the pBluescript vectors encoding either wild-type SoxR (pKOXR) or the respective mutant proteins encoded by pKENC119A, pKENC122A, pKENC124A and pKENC130A, SDS–PAGE analysis of cell extracts demonstrated the presence of similar amounts of the wild-type and the four cysteine mutant proteins, each with M_r = 17000 (identical to that of wild-type SoxR; Fig. 2A).

We sought to determine whether elimination of the cysteine residues might impair the ability of the mutant proteins to bind soxS promoter DNA in vitro in electrophoretic mobility shift assays (EMSA) (11). Cell-free extracts containing wild-type and mutant SoxR proteins displayed similar levels of soxS binding activity in vitro that increased with the amount of extract protein added (Fig. 2B). The mutant and wild-type proteins therefore bind the soxS target with similar affinities, since the proteins were present at similar levels in the extracts (Fig. 2A).

Since the Cys→Ala mutant SoxR proteins retained DNA binding activity, all four proteins were extensively purified (to ≥90% homogeneity), in parallel with wild-type SoxR and in similar yield (data not shown). The purified mutant proteins each lacked the visible absorbance characteristic of oxidized Fe–SoxR and had no significant Fe detectable by inductively coupled plasma emission spectrometry (<0.1 atom Fe per SoxR monomer; data not shown). It seemed possible that the Cys→Ala mutant proteins might contain iron in vivo, but lose the metal during purification. Analysis by EPR spectroscopy of intact cells overexpressing the various proteins showed that all the Cys→Ala

Figure 2. (A) Coomassie blue-stained SDS–polyacrylamide gel of protein extracts from TN531 cells overexpressing wild-type and mutant SoxR proteins. SoxR, 1 or 0.5 μg purified wild-type SoxR protein. Cell extracts (15 μg): WT, pKOXR plasmid; C1, pKENC119A plasmid; C2, pKENC122A plasmid; C3, pKENC124A plasmid; C4, pKENC130A plasmid.

(B) Binding activity for soxS promoter DNA. SoxR-containing cell extracts. Lane 1, DNA alone; lane 2, vector control (5 μg); lanes 3 and 4, wild-type SoxR extract (WT); lanes 5 and 6, SoxR-C119A extract (C1); lanes 7 and 8, SoxR-C122A extract (C2); lane 9 or 10, SoxR-C124A extract (C3); lanes 11 and 12, SoxR-C130A extract (C4); odd numbered lanes correspond to 0.5 μg extract protein, even numbered lanes to 5 μg. C, SoxR–DNA complex; D, free DNA.
expression
mutant
completely eliminated
of the
overexpressing wild
both the vector control (pSE380) and the strain
harbors
overexpressing one of the mutant proteins
vectors), expression of soxS by wild-type SoxR in response to PQ
was unaffected (data not shown).

An additional study was conducted to assess the effect of
overexpressing one of the mutant proteins (C119A) on the
inducibility of soxS in TN521 (soxR+) cells. Because this strain
harbors only the lacI* allele, we used the vector pSE380 (bearing
lacI*: see Materials and Methods) to achieve tightly regulated
SoxR expression inducible by IPTG. After 1000-fold dilution of
overnight cultures into fresh medium and outgrowth for 60 min,
IPTG was added, followed at various times by a 30 min exposure
to PQ (250 μM) to measure activation of soxS transcription. The
induction of soxS2::lacZ by PQ was essentially the same for
both the vector control (pSE380) and the strain
overexpressing wild-type SoxR (from pSXIR) throughout the
course of the experiment (Fig. 6). In contrast, the expression
of the C119A protein strongly interfered with soxS induction,
even at the earliest time point (some leaky expression of SoxR
is expected even in the absence of IPTG). PQ inducibility was
completely eliminated after only a 30 min pre-induction of the
mutant protein with IPTG (Fig. 6). As seen in Figure 5B, basal
expression of

®-galactosidase from soxS2::lacZ was increased to <1000 U
in cells overexpressing the C119A protein.

**DISCUSSION**

Our previous work established that the SoxR [2Fe-2S] centers
are essential for the protein’s transcriptional activity at soxS in vitro
(11,13–15). Here, we have extended the demonstration of this
requirement to the activity of SoxR in vivo by engineering mutant
genes that direct the synthesis of SoxR derivatives with individual
cysteine residues replaced by alanines. For at least the C119A
dervative, high-level synthesis interfered with the normal activity
of SoxR during activation by PQ. Despite their inability to be
activated by oxidative stress, overexpression of the Cys→Ala
SoxR proteins revealed an unexpectedly high level of basal
transcriptional activity in vivo that was not detected in vitro.

Removal of the SoxR [2Fe-2S] clusters in vitro by aerobic
exposure of the protein to 2-mercaptoethanol (11,13) or the
biological thiol glutathione (15) eliminates the transcriptional
activity at soxS without any apparent effect on SoxR protein
stability, oligomeric state or binding affinity for the soxS
promoter. Elimination of detectable metal binding by Cys→Ala
replacements effectively blocked SoxR activation in response to
PQ and did not significantly affect protein expression in vivo or
soxS binding in vitro. These observations provide new evidence
for post-translational activation of SoxR, and show that this
activation in vivo depends critically on the integrity of the
[2Fe-2S] centers. The results also support the conclusion from
EPR spectroscopy that all four SoxR cysteine residues are
involved in anchoring the metal center. Efforts to assemble
[2Fe-2S] centers into the Cys119→Ala protein in vitro were
unsuccessful (14).

Re-insertion in vitro of [2Fe-2S] centers into wild-type apo-
SoxR restored full transcriptional activity (14). However, the
properties of the Cys→Ala mutant proteins alone do not resolve
whether the critical step for SoxR activation in vivo is synthesis
of the [2Fe-2S] centers or a subsequent redox reaction. We have
previously suggested (13,14) that these two steps might be linked
if the stability of the metal centers is influenced by oxidation and

*Figure 3.* EPR analysis of SoxR in vivo. The EPR spectra of cell suspensions were recorded as described previously for purified Fe-SoxR (13). The intracellular SoxR concentration [SoxR] was estimated by immunoblotting of whole cell extracts (see Materials and Methods). The content of reduced [2Fe-2S] SoxR in the cells was estimated from the amplitude of the EPR resonance at g1 (15) normalized to a known preparation of purified SoxR.

*Figure 4.* SoxR-dependent in vitro transcription of soxS. Fe-SoxR (active), apo-SoxR (inactive) or Cys→Ala mutant proteins (5 ng each) were incubated with 100 nM RNAP in in vitro transcription reactions containing a plasmid carrying both the SoxR-dependent soxS gene and the SoxR-independent bla gene as indicated in Materials and Methods. The corresponding primer extension reactions (13,14) for the soxS and bla transcripts are indicated.
in Figure 5. Expression of soxS::lacZ as a function of mutant SoxR expression in strain TN5311 (ΔsoxR). (A) Low level expression via uninduced pBluescript vectors or (B) high level expression via induced pKEN2 vectors. +, incubation with PQ; -, non-exposed cells. β-galactosidase activity is expressed as Miller units (21).

In conclusion, SoxR appears to require formation of two Fe-S centers for complete function. Regarding the second possible mode of interference, wild-type SoxR and the Cys-Ala mutant proteins bound the soxS promoter equally well in vitro (Fig. 2B). However, RNAP binds the soxS promoter cooperatively with activated SoxR (11), an effect that could disfavor competition by low levels of the Cys-Ala mutant proteins.

The second unexpected feature of the Cys-Ala mutant forms of SoxR was the relatively high basal soxS transcription they exert upon overexpression in vitro, which contrasts with the essentially undetectable activity of these proteins in vivo. It remains possible that the Cys-Ala mutant proteins cause transcriptional activation higher than that of apo-SoxR, but at a level still undetectable in our in vitro assay. Alternatively, the in vitro activity of the mutant proteins in the absence of a redox signal could suggest the existence of additional controls on soxS activation that are overcome by high concentrations of the Cys-Ala derivatives but not non-activated wild-type SoxR. The known negative autoregulation by SoxS (27) is evidently not responsible for this effect, which we observed in SoxS deficient cells. Previous genetic studies (2,3) identified regulatory mutations only in soxR and soxS (9,10), but a smaller effect on soxS expression caused by other mutations might have been missed. One possible negative regulator is Rob protein, which is abundant (≤5000 molecules/cell; (28)) and has DNA binding specificity that overlaps that of SoxS (29–31). MarA protein also exhibits overlapping specificity with SoxS (32), but its expression is kept low through repression by the MarR protein (33).

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Figure 6. Interference of SoxR-C119A protein with wild-type SoxR in vitro. The bottom lines indicate the plasmid present (pSE380, control; pSXR, wild-type SoxR; pSEC119, SoxR-C119A) in strain TN521 (soxR::soxS2::lacZ), the time of IPTG treatment to induce recombinant SoxR, and exposure to PQ (+). β-Galactosidase activity is shown in units (21). All values represent the mean of three independent samples, except pSEC119G min/no PQ, for which only two samples were available.

in Figure 5. Expression of soxS::lacZ as a function of mutant SoxR expression in strain TN5311 (ΔsoxR). (A) Low level expression via uninduced pBluescript vectors or (B) high level expression via induced pKEN2 vectors. +, incubation with PQ; -, non-exposed cells. β-galactosidase activity is expressed as Miller units (21).

For wild-type SoxR, the quantity of reduced [2Fe-2S] centers detected in untreated cells corresponded to only 36% of the total SoxR protein (Fig. 3). The EPR-silent forms include both oxidized and apo-SoxR (12,13), with only the former active in vitro (11,16,17). The in vitro EPR analysis shown here and elsewhere (18) thus supports the conclusion that reduced Fe-SoxR is transcriptionally inactive in cells. Some oxidation evidently occurs during sample preparation by the method employed here: using a modified method (H.D. and B.D., manuscript in preparation), we have measured levels of reduced wild-type SoxR representing ≥95% of the total SoxR.

Two additional features of the SoxR Cys-Ala mutants merit mention. First, during activation by PQ, none of the substituted forms, when expressed at approximately wild-type levels, competed detectably with wild-type SoxR expressed from a single-copy gene; such interference was observed only upon high level expression of the C119A derivative. In view of their evidently normal stability and DNA binding affinity, the relative

reduction. However, recent experiments (16,17) indicate that SoxR with reduced [2Fe-2S] centers is stable, but transcriptionally inactive in vitro. In vivo studies of the properties of constitutive forms of SoxR extend this model to living cells (18).

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