THE UNIVERSITY OF RHODE ISLAND

University of Rhode Island DigitalCommons@URI

Pharmacy Practice and Clinical Research Faculty Publications

Pharmacy Practice and Clinical Research

5-5-2015

Treatment Options for Carbapenem-Resistant Enterobacteriaceae Infections

Haley J. Morrill University of Rhode Island

M. Pogue

Keith S. Kaye

Kerry L. LaPlante University of Rhode Island, kerrylaplante@uri.edu

Follow this and additional works at: https://digitalcommons.uri.edu/php_facpubs

Citation/Publisher Attribution

Haley J. Morrill, Jason M. Pogue, Keith S. Kaye, Kerry L. LaPlante; Treatment Options for Carbapenem-Resistant *Enterobacteriaceae* Infections, *Open Forum Infectious Diseases*, Volume 2, Issue 2, 1 April 2015, ofv050, https://doi.org/10.1093/ofid/ofv050 Available at: https://doi.org/10.1093/ofid/ofv050

This Article is brought to you by the University of Rhode Island. It has been accepted for inclusion in Pharmacy Practice and Clinical Research Faculty Publications by an authorized administrator of DigitalCommons@URI. For more information, please contact digitalcommons-group@uri.edu. For permission to reuse copyrighted content, contact the author directly.

Treatment Options for Carbapenem-Resistant Enterobacteriaceae Infections

The University of Rhode Island Faculty have made this article openly available. Please let us know how Open Access to this research benefits you.

This is a pre-publication author manuscript of the final, published article.

Terms of Use

This article is made available under the terms and conditions applicable towards Open Access Policy Articles, as set forth in our Terms of Use.

Treatment Options for Carbapenem-Resistant Enterobacteriaceae Infections

Haley J. Morrill, Pharm.D.^{a,b}, Jason M. Pogue, Pharm.D., BCPS-ID^c, Keith S. Kaye, M.D., M.P.H.^d, and Kerry L. LaPlante, Pharm.D.^{a,b,e*}

^aVeterans Affairs Medical Center, Infectious Diseases Research Program, Providence, RI, USA ^bUniversity of Rhode Island, College of Pharmacy, Department of Pharmacy Practice, Kingston, RI, USA ^cDetroit Medical Center, Wayne State University, Department of Pharmacy Services, Detroit, MI, USA ^dDetroit Medical Center, Wayne State University, Division of Infectious Diseases, Detroit, MI, USA ^eWarren Alpert Medical School of Brown University, Division of Infectious Diseases, Providence, RI, USA

*Corresponding author: Kerry L. LaPlante, Pharm.D. Associate Professor, University of Rhode Island, College of Pharmacy; 7 Greenhouse Road Suite 295A, Kingston RI, 02881; Tel: 401-874-5560 e-mail: Kerrylaplante@uri.edu

Keywords: carbapenem-resistant enterobacteriaceae; carbapenemases; resistant infections; treatment; carbapenems

Running head: CRE treatment

Summary: This article provides a comprehensive review of currently available treatment options for infections due to carbapenem-resistant enterobacteriaceae (CRE).

Abstract.

Antimicrobial resistance in Gram-negative bacteria is an emerging and serious global public health threat. Carbapenems have been used as the "last-line" treatment for infections caused by resistant enterobacteriaceae, including those producing extended spectrum β-lactamases. However, enterobacteriaceae that produce carbapenemases, which are enzymes that deactivate carbapenems and most other β-lactam antibiotics, have emerged and are increasingly being reported worldwide. Despite increasing burden, the most optimal treatment for carbapenem-resistant enterobacteriaceae (CRE) infections is largely unknown. For the few remaining available treatment options, there is limited efficacy data to support their role in therapy. Nevertheless, current treatment options include the use of older agents, such as polymyxins, fosfomycin, and aminoglycosides, which have been rarely used due to efficacy and/or toxicity concerns. Optimization of dosing regimens and combination therapy are additional treatment strategies being explored. CRE infections are associated with poor outcomes and high mortality. Continued research is critically needed to determine the most appropriate treatment.

Introduction.

Antimicrobial resistance is globally recognized as one of the greatest threats to public health. Of particular concern, are infections caused by resistant Gram-negative bacilli, which are increasingly being reported worldwide. The escalating burden of Gram-negative antimicrobial resistance is largely due to ß-lactamases, which are enzymes that bind and deactivate ß-lactam antibiotics, rendering them ineffective. For years, carbapenems have been used successfully to treat infections due to resistant enterobacteriaceae, such as *Escherichia Coli* and *Klebsiella pneumoniae*, including those producing extended spectrum ß-lactamases (ESBLs; a subset of ß-lactamase enzymes which confer broad resistance to penicillins, cephalosporins, and the monobactam aztreonam).

However, recently enterobacteriaceae producing carbapenemases (known as carbapenem-resistant enterobacteriaceae [CRE]) have emerged, which confer broad resistance to most B-lactam antibiotics including "last-line" carbapenems. Carbapenem resistance can also be conferred when porin deficiencies, which allow decreased entry of the B-lactam into the cell membrane, are combined with ESBLs.[1] The prevalence of CRE infections has incresed over the last decade, especially in healthcare settings and CRE have been recognized by the United States Centers for Disease Control and Prevention as an urgent public health threat.[2, 3] The Ceners for Disease Control and Prevention estimates that more than 9,000 healthcareassocaited infections are caused by the two most common type of CRE, carbapenem-resistant Klebsiella species and carbapenem-resistant Escherichia species, each year in the United States.[3] CRE can cause a number of serious infection types (such as intra-abdominal infections, pneumonia, urinary tract infections, and device-associated infections) or asymptomatic colonization.[4-6] Each year approximtaley 600 deaths result from CRE infections. [3] CRE mortality rates are high and range from 18% to 60% depending on therapy.[7] This may be due to delayed time to active therapy, pharmacologic limitations of available treatment options, and that patients with CRE infections tend to be critically ill.

At this time there are a limited selection of treatment options for CRE infections. Clinicians have been forced to re-evaluate the use of agents, which have been historically rarely used due to efficacy and/or toxicity concerns, such as polymyxins, fosfomycin, and aminoglycosides. Additional CRE treatment strategies include optimization of dosing regimens and combination therapy. This review will focus on the current treatment options for CRE infections.

Overview of Carbapenem-resistant Enterobacteriaceae (CRE)Treatment.

There are numerous different types of carbapenemase enzymes, each conferring varying spectrums of resistance. An overview of the carbapenemase enzyme types with the greatest clinical importance can be found in Table 1. In general, the presence of a carbapenemase confers broad resistance to most ß-lactam antibiotics including penicillins, cephalosporins, and the monobactam aztreonam (excluding MBLs and OXAs).[1] *In vitro* activity of carbapenems in the setting of one of these enzymes is variable, and the exact role of carbapenems in infectious due to these organisms is controversial. To further complicate treatment, CRE often exhibit resistance to structurally unrelated antimicrobial classes such as aminoglycosides and fluoroquinolones.[8] However, aminoglycoside susceptibility can vary as a function of KPC strain type and co-existing aminoglycoside modifying enzymes, which are not tested in a traditional clinical laboratory. The emergence of resistance during therapy is another emerging concern.[9, 10]

Despite their increasing burden, the most optimal treatment for CRE infections is largely unknown. At this time, there is no published data from randomized controlled trials assessing antimicrobial treatment options for CRE infections. While important, in the United States at this time there may not be a sufficient amount of patients with serious CRE infections to conduct such a trial. Therefore, much of the existing evidence is from reviews of case reports, case series and small retrospective studies, which have a number of inherent limitations.[11, 12] A

potential CRE treatment algorithm and overviews of current treatment options can be found in Tables 2 and 3, respectively.

Carbapenems.

Pharmacokinetic data suggests that T>MIC targets can be achieved using high-dose prolonged-infusion carbapenems when carbapenem MICs are relatively low (<4µg/ml) or even moderately elevated (8-16µg/ml).[13-17] In a pharmacokinetic study of ten critically ill patients, high-dose meropenem (6000mg/day) administered by prolonged (over 4 hours)/continuous infusion had a high probability of target attainment (PTA) up to an MIC of 8-16µg/ml.[13] In another study, the PTA for an MIC of 4µg/ml increased with prolonged-infusion (over 3 hours) as compared to traditional-infusion (over 30 minutes); the PTA for prolonged-infusions were 100% (2000mg q8h) and 93% (1000mg q8h) as compared to 69% for traditional-infusion (1000mg q8h).[14] At an MIC of 8µg/ml, only high-dose prolonged-infusion meropenem had a high PTA(85%).

While pharmacokinetic data appears favorable, there is only limited clinical data assessing the efficacy of carbapenem monotherapy in the treatment of CRE infections. In a study that compiled data from eight clinical trials, in 44 patients treated with carbapenem monotherapy for infections due to carbapenemase-producing *K. pneumoniae*, treatment efficacy varied based on MIC.[17] The efficacy ranged from 69% (MIC $\leq 4\mu$ g/ml), 60% (MIC 8μ g/ml), to only 29% (MIC $>8\mu$ g/ml). Treatment efficacy when the MIC was $\leq 4\mu$ g/ml was similar to that observed in 22 patients with non-carbapenemase-producing *K. pneumoniae* infections (73%). The lowest mortality rate was observed in patients who received carbapenem-containing-combination treatment (MIC $\leq 4\mu$ g/ml). The mortality rate was lower for patients who received carbapenem-containing as compared to non-carbapenem regimens (12%[3/26] vs. 41%[46/112]; P=0.006).[17] In a recent review, the mortality rate associated with carbapenem monotherapy was unacceptably high (40.1%).[12] For patients with serious infections and/or

who are critically ill adding another active agent may increase the probability of clinical response.

Additionally, several retrospective studies have observed lower rates of mortality with carbapenem-based combination therapy as compared to non-carbapenem combination therapy.[17-20] The efficacy of carbapenem combination therapy also appears to be MIC dependent. In a large multi-center study where high-dose prolonged-infusion meropenem was used (2000mg administered over >3 hours q8h) mortality rates stratified by MIC were as follows: 13%(2/13) for $<4\mu g/ml$, 33% (1/3) for $8\mu g/ml$, and 35.2%(6/17) for $\geq 16\mu g/ml$.[19] In a large cohort study (see Table 4), the mortality rate associated with carbapenem-containingcombination therapy for carbapemase-producing K. pneumoniae bacteremia increased from 19.3% (MIC<8µg/ml) to 35.5% (MIC>8µg/ml).[20]. In a review of 20 clinical studies, carbapenem-containing-regimens were associated with lower mortality than non-carbapenemcontaining-regimens (18.8% vs. 30.7%).[12] While encouraging, it is important to note, that not all reports have focused on carbapenem-containing-regimens. A retrospective study conducted from a 10-bed intensive-care-unit (ICU) showed success in 24/26(92%) patients with KPC infections (16 ventilator-associated pneumonias [VAP], 7 bloodstream infections, 2 urinary tract infections [UTI], 1 peritonitis) with the use of carbapenem-sparing-combination therapy regimens.[21]

Double-carbapenem combination treatment may be an effective option for infections caused by pan-drug-resistant CRE, however data is limited to selected case reports.[22, 23] Experimental data has shown that the KPC enzyme may have increased affinity for ertapenem than other carbapenems, therefore when given together; KPC preferentially deactivates ertapenem, which hinders degradation and improves the activity of the concomitant carbapenem.[24, 25] In case reports, ertapenem plus either doripenem or meropenem has been used successfully to treat select pan-drug-resistant and colistin-resistant KPC-producing *K. pneumoniae* infections (bacteremia, VAP, and UTI). Double-carbapenem combination

treatment is a promising option, which may be most effective in combination with a third drug.[26]

Polymyxins. Colistin (polymyxin E) and polymyxin B are considered to be the most active *in vitro* agents against CRE.[27] Polymyxin B and colistin differ by a single amino acid. A comparison of the two drugs can be found in Table 5. There are several potential advantages to the use of polymyxin B over colistin, many of which stem from the fact that colistin is administered as the inactive prodrug colistimethate (CMS). Only a small fraction of CMS is converted to colistin and this conversion is slow, with maximum concentrations occurring \geq 7 hours after administration.[28] As the conversion of CMS to colistin is slow and inefficient in patients with normal renal function the majority of CMS is cleared prior to conversion to colistin. Therefore, despite being dosed at a lower mg/kg/day dose, polymyxin B can achieve higher peak serum concentrations which are achieved much more rapidly than with colistin.[28, 29]

Renal dose adjustments are necessary for colistin/CMS but are not required polymyxin B.[26] The reason for this is that there is minimal renal clearance of colistin, but the prodrug CMS is predominately cleared renally.[26] As with colistin, polymyxin B undergoes extensive renal tubular reabsorption and is eliminated by mostly nonrenal clearance. Importantly, however, polymyxin B package insert dosing recommendations include vague renal dosing adjustments that have been followed in all of the polymyxin B literature to date. The efficacy and safety of non-renally adjusted polmyxin B remains unclear. The renal clearance of CMS allows an advantage over polymyxin B that a higher concentration of active drug in the urine is reached which would make colistin/CMS a viable UTI treatment alternative.[26, 30] Despite the potential advantages of polymyxin B use, the majority of clinical data to date for CRE infections has focused on the use of colistin.

The ideal dosages of colistin and polymyxin B are largely unknown, especially in the case of renal failure, renal replacement therapy, and critical illness.[31] Scientifically based

dosing recommendations can be found in Table 5.[28, 29] For serious infections caused by resistant Gram-negative pathogens, high total daily doses of colistin appear to be important to maximize treatment efficacy.[28, 32] In a retrospective study of 258 ICU patients treated with CMS, 21.7% of patients on the highest total daily dose (9 million IU/day) died as compared to 27.8% and 38.6% patients on lower doses of 6 and 3 million IU/day, respectively (p=0.0011).[33] In a retrospective study of 67 patients with Gram-negative bacteremia, the median colistin dose was higher in patients who achieved microbiological success (2.9 vs. 1.5mg/kg/day; P=0.011) and 7-day survival (2.7 vs 1.5mg/kg/day; P=0.007).[32] Another retrospective study found similar results with polymyxin B treatment.[34]

Historically, neurotoxicity was an important concern with the use of polymyxins, however with current formulations this side effect is reported less frequently. Patients discussed in the recent literature are more critically ill, ventilated, and sedated which might significantly limit the ability to detect neurotoxicity, which primarily manifested as parasthesias and ataxia. However, nephrotoxicity remains a concern as it continues to occur in >40% patients treated with polymyxins.[35] While nephrotoxicity has been reported with both colistin and polymyxin B use, recent evidence suggests that nephrotoxicity rates might be higher with colistin use than polymyxin B (50-60% vs. 20-40%).[35, 36] The use of colistin and polymyxin B at higher doses, which may be necessary for CRE infections, may be associated with a higher risk of nephrotoxicity.[32, 34] The better outcomes associated with high dose colistin, may come at the cost of worsening renal function.[32] In a retrospective study, a colistin dose of >5mg/kg of ideal body weight/day was independently predictive of the development of renal insufficiency.[37] For polymyxin B, a retrospective cohort study of 276 patients demonstrated that high doses (>200mg/day) were independently associated with lower mortality (adjusted OR 0.43; 95% CI 0.23–0.79).[34] However, the use of \geq 200mg/day was associated with a significantly higher risk of severe renal impairment (adjusted OR 4.51; 95% CI 1.58-12.90; P = 0.005). Even when controlling for the development of moderate to severe renal

dysfunction, multivariate analyses showed that doses ≥200mg/day were still associated with decreases in mortality.

Another concern with the use of polymyxins is on-treatment resistance development. Blood isolates from one patient infected with carbapenem-resistant *K. pneumoniae* and treated with polymyxin B monotheray, showed a significantly increased polymyxin B MIC in just 5 days (0.75µg/ml to 1,024µg/ml).[9] Additionally, there have been reports of colistin-resistant, carbapenem-resistant *K. pneumoniae* outbreaks.[38, 39] Therefore, polymyxins may be most effective as part of a combination for serious CRE infections.[31, 40] In a recent review which used compiled data on 889 patients with CRE infections (bacteremia, pneumonia, intraabdominal infections, UTIs, and surgical site infections), the mortality rate for colistin monotherapy was 42.8%.[12] A review of 55 studies found that clinical success was lower for colistin monotherapy as compared to colistin combination therapy for treatment of infections caused by KPC-producers (14% [1/7] vs. 73% [8/11]).[41] In a recent cohort study of 36 patients with blood stream infections due to CRE (all but two yielded both OXA-48 and CTX-M ESBLs), colistin based combination therapy was associated with better 28-day survival than non-colistin regimens (33.3% vs. 5.5%; p=0.018).[42]

Tigecycline. The majority of CRE isolates remain active against tigecyline *in vitro*, however resistance to tigecycline is increasing.[43-45] There are only limited clinical data to support use of tigecycline monotherapy for infections caused by CRE that demonstrate *in vitro* susceptibility.[19, 20, 41, 46, 47] In a small number of patients with carbapenem-resistant *K. pneumoniae*, 71.4% (5/7) patients had a favorable outcome with tigecycline treatment.[41] High mortality rates have been reported with the use of tigecycline monotherapy in the treatment of bloodstream infections due to carbapenem-resistant *K. pneumoniae* in two separate cohort studies (see Table 4).[19, 20] Additionally, despite *in vitro* susceptibility, on-treatment resistance emergence has been described.[10, 40, 48]

Tigecycline may be most effective when used at higher doses and/or in combination for serious CRE infections, and depending on the source of the infection.[40, 49, 50] However, high dose tigecycline may only transiently lead to increased plasma concentrations, as higher doses may lead to increased intracellular accumulation and tissue distribution.[49] In 30 complex patients with severe intra-abdominal infections due to KPC-producing *K. pneumoniae,* high dose tigecycline in combination with colistin was associated with lower mortality as compared to approved dose tigecycline plus colistin.[51] In a review which used compiled data on patients with various types of CRE infections, the mortality rate with tigecycline monotherapy was 41.1%.[12] A carbapenem-sparing regimen of tigecycline plus either gentamicin or colistin was effective in 92%(24/26) of ICU patients treated for KPC infections.[21]

Fosfomycin. Limited data has demonstrated fosfomycin has activity against KPC-producing *K*. *pneumoniae* and NDM-1-producing enterobacteriaceae.[52, 53] Fosfomycin achieves high urinary concentrations for prolonged time periods (after a single 3 gram dose peak urine concentrations of >4000µg/ml are obtained and above MIC concentrations persist for 72 hours).[54] Select case reports have demonstrated success of oral fosfomycin for treating UTIs caused by fosfomycin susceptible KPC- and NDM-producing enterobacteriaceae.[55, 56] Two patients with OXA-48-producing *K. pneumoniae* UTIs were successfully treated with oral fosfomycin and colistin.[57]

In Europe an intravenous fosfomycin formulation is available. In a small (n=11) European study, intravenous fosfomycin (2-4 g q6h) in combination was associated with good bacteriological and clinical outcomes in all patients for various carbapenem-resistant *K. pneumoniae* infections (bacteremia, VAP, UTI, wound infections).[58] In a report of three cases of KPC-producing *K. pneumoniae* bacteremia, intravenous fosfomycin was used as an adjunct "last-resort" treatment which initially led to bacteremia control, however ultimately all three patients failed treatment due to relapse and resistance development.[59] The use of

intravenous fosfomycin monotherapy for the treatment of systemic infections may be limited due to the potential for the development of drug resistance during treatment.[60]

Aminoglycosides. Gentamicin is generally the most active aminoglycoside *in vitro* against carbapenem-resistant *K. pneumoniae*, however amikacin can be most active against other CRE.[46, 61, 62] Data on the use of aminoglycosides as monotherapy is limited and aminoglycosides monotherapy appears to be most efficacious in the treatment of UTIs.[12, 41, 63] In a retrospective cohort study of cases of carbapenem-resistant *K. pneumoniae* bacteriuria, treatment with an *in vitro* active aminoglycoside was associated with a significantly higher rate of microbiologic clearance as compared to either polymyxin B or tigecycline.[63] In multivariate analysis, aminoglycoside treatment was independently associated with microbiologic clearance.

Aminoglycoside therapy may be most appropriate as a component of combination therapy for infections, especially UTIs, caused by CRE.[64-66] In the largest CRE bacteremia cohort study to date, similar mortality rates were observed for aminoglycoside monotherapy (22.2%) and combination therapy (26.5%), however only a small number of patients (n=9) were treated with monotherapy as compared to 68 patients treated with aminoglycoside combination therapy.[20] In a review of 24 cases of aminoglycoside combination therapy (most often with colistin, carbapenems, fluoroquinolones, and tigecycline), all patients who failed aminoglycoside based combination therapy had bloodstream infections.[66] In a review of 20 clinical studies, the combination of an aminoglycoside and a carbapenem had the lowest mortality rate (11.1%).[12]

Combination Therapy. Combination therapy for CRE infections may decrease mortality as compared to monotherapy. It is also an important empiric consideration when a CRE is suspected.[18, 19, 31] Benefits of combination therapy include reduction of initial inappropriate

antimicrobial therapy, potential synergistic effects, and suppression of emerging resistance.[31] As monotherapy options all have significant limitations (pharmacokinetics, toxicity, emergence of resistance), combination therapy can be an attractive option to optimize therapy. However, with combination therapy there is the potential for an increased risk for the development of *Clostridium difficile* infection, colonization/infection with other resistant bacteria, and adverse effects such as nephrotoxicity.[11, 31] Combination therapy leads to increased antimicrobial pressure and may potentiate the development of antimicrobial resistance. The benefits of combination therapy may outweigh the risks, and many experts recommend combination therapy as opposed to monotherapy for the treatment of severe CRE infections.[31, 40]

As previously described, emerging clinical evidence suggests that treatment with combination therapy may be beneficial for serious CRE infections.[12, 18-21, 41, 42, 67-69] In the most comprehensive review to date, which included data on 889 patients with CRE infections, combination therapy with two or more in vitro active agents was associated with lower mortality than treatment with a single in vitro active agent (27.4%[121/441] vs. 38.7%[134/346], p<0.001).[12] Monotherapy resulted in mortality rates that were not significantly different from those in patients treated with inappropriate therapy with no in vitro active agents (46.1%[48/102]). Another comprehensive review found similar mortality results Several observational studies have assessed the efficacy of (18.3% vs. 49.1%).[31] combination therapy versus monotherapy in the treatment of bloodstream infections due to carbapenemase-producing K. pneumoniae (mostly KPC-producers).[18-20, 67] A summary of these studies can be found in Table 4. In the first study, all patients who received combination therapy had favorable outcomes, while 46.7% patients who received active monotherapy died.[67] The next retrospective cohort study also demonstrated lower mortality rate with combination treatment (usually a carbapenem with colistin or tigecycline) compared with monotherapy.[18] A larger multi-center retrospective cohort study also found similar results.[19] Interestingly, meropenem, colistin, tigecycline combination was associated with a significant

reduction in mortality even in patients who received inappropriate empiric therapy (14% vs. 61%). In the most recent and largest cohort study to date, combination therapy again was associated with lower mortality than monotherapy (27.2% vs. 44.4%).[20] Combination therapy was an independent predictor of survival; which was mostly due to the effectiveness of carbapenem-containing regimens.

Emerging treatment. An overview of emerging treatment options can be found in Table 6. The Food and Drug Administration approved ceftazidime-avibactam in February 2015 for the treatment of complicated intra-abdominal infections (cIAI) and complicated urinary tract infections (cUTI).[70] It is expected that ceftazidime-avibactam will be available in the second quarter of 2015, however ceftazidime-avibactam received a priority review based on Phase II data, and as such should be reserved for patients with limited or no alternative treatment options.[70]

Ceftazidime-avibactam is combination of an established broad-spectrum cephalosporin (ceftazidime) and a novel β -lactamase inhibitor (avibactam) with activity against class A, class C, and some class D β -lactamases.[71, 72] Avibactam has activity against KPC-type carbapenemases and some OXA enzymes, however it has no activity against metallo- β - lactamases (such as NDM-1).[71, 72] In two Phase II trials, efficacy and safety rates were similar for ceftazidime-avibactam versus comparator drugs for the treatment of clAI and cUTI.[73, 74] For clAI, favorable clinical response rates were observed for ceftazidime-avibactam (2000/500 mg IV q8h) plus metronidazole (500 mg IV q8h) as compared to meropenem (1000 mg IV q8h; 91.2% [62/68] vs. 93.4% [71/76], p=0.06).[74] For cUTI, favorable clinical response rates were observed for ceftazidime-avibactam (500/125 mg IV q8h) as compared to imipenem (500 mg IV q6h; 85.7% [24/28] vs. 80.6% [29/36], p=0.06).[73] The most common adverse drug reactions (>10%) in trials were vomiting, nausea, constipation, and anxiety.[70] In a Phase III trial, clinical cure rates for ceftazidime-avibactam were lower for

patients with a creatinine clearance between 30 to 50 ml/min.[70] Additionally, seizures have been reported with the use of ceftazidime and as with other β-lactam antibiotics there is a risk for serious hypersensitivity.[70] Phase III trails are underway assessing ceftazidime-avibactam for the treatment of cIAI, cUTI, and nosocomial pneumonia and results will likely be available in late 2015.[70]

Conclusions. The burden of antimicrobial resistance among Gram-negative pathogens, particularly carbapenem-resistant enterobacteriaceae is increasing rapidly worldwide. Treatment options for serious CRE infections remains extremely limited at this time. Optimization of dosing of currently available agents and combination therapy may be the most appropriate treatment strategies at this time. However, continued research is desperately needed, in particular randomized controlled trials, to determine the most appropriate treatment for serious CRE infections.

Funding.

The views expressed in this article are those of the authors and do not necessarily reflect the position or policy of the United States Department of Veterans Affairs. This work was supported, in part, by the Office of Research and Development, Department of Veterans Affairs. This work was unfunded. Haley Morrill, Jason Pogue, and Keith Kaye have no conflicts to disclose. Kerry LaPlante has received research funding or acted as an advisor or consultant for Astellas, Cubist, Davol, Forest, and Pfizer Inc.

References.

- 1. Nordmann P, Dortet L, Poirel L. Carbapenem resistance in Enterobacteriaceae: here is the storm! Trends in molecular medicine **2012**; 18(5): 263-72.
- 2. Centers for Disease C, Prevention. Vital signs: carbapenem-resistant Enterobacteriaceae. MMWR Morbidity and mortality weekly report **2013**; 62(9): 165-70.
- 3. Centers for Disease Control and Prevention (CDC). Antibiotic resistance threats in the United States, 2013. Atlanta: CDC; 2013. Available from: <u>http://www.cdc.gov/drugresistance/threat-report-2013/pdf/ar-threats-2013-508.pdf</u>.
- 4. Hossain A, Ferraro MJ, Pino RM, et al. Plasmid-mediated carbapenem-hydrolyzing enzyme KPC-2 in an Enterobacter sp. Antimicrobial agents and chemotherapy **2004**; 48(11): 4438-40.
- 5. Peleg AY, Franklin C, Bell JM, Spelman DW. Dissemination of the metallo-beta-lactamase gene blaIMP-4 among gram-negative pathogens in a clinical setting in Australia. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2005**; 41(11): 1549-56.
- 6. Leavitt A, Navon-Venezia S, Chmelnitsky I, Schwaber MJ, Carmeli Y. Emergence of KPC-2 and KPC-3 in carbapenem-resistant Klebsiella pneumoniae strains in an Israeli hospital. Antimicrobial agents and chemotherapy **2007**; 51(8): 3026-9.
- Akova M, Daikos GL, Tzouvelekis L, Carmeli Y. Interventional strategies and current clinical experience with carbapenemase-producing Gram-negative bacteria. Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases 2012; 18(5): 439-48.
- 8. Bratu S, Tolaney P, Karumudi U, et al. Carbapenemase-producing Klebsiella pneumoniae in Brooklyn, NY: molecular epidemiology and in vitro activity of polymyxin B and other agents. The Journal of antimicrobial chemotherapy **2005**; 56(1): 128-32.
- 9. Lee J, Patel G, Huprikar S, Calfee DP, Jenkins SG. Decreased susceptibility to polymyxin B during treatment for carbapenem-resistant Klebsiella pneumoniae infection. Journal of clinical microbiology **2009**; 47(5): 1611-2.
- 10. Anthony KB, Fishman NO, Linkin DR, Gasink LB, Edelstein PH, Lautenbach E. Clinical and microbiological outcomes of serious infections with multidrug-resistant gram-negative organisms treated with tigecycline. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2008**; 46(4): 567-70.
- 11. Paul M, Carmeli Y, Durante-Mangoni E, et al. Combination therapy for carbapenem-resistant Gram-negative bacteria. The Journal of antimicrobial chemotherapy **2014**.
- 12. Tzouvelekis LS, Markogiannakis A, Piperaki E, Souli M, Daikos GL. Treating infections caused by carbapenemase-producing Enterobacteriaceae. Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases **2014**.
- 13. Roberts JA, Kirkpatrick CM, Roberts MS, Robertson TA, Dalley AJ, Lipman J. Meropenem dosing in critically ill patients with sepsis and without renal dysfunction: intermittent bolus versus continuous administration? Monte Carlo dosing simulations and subcutaneous tissue distribution. The Journal of antimicrobial chemotherapy **2009**; 64(1): 142-50.
- 14. Kuti JL, Dandekar PK, Nightingale CH, Nicolau DP. Use of Monte Carlo simulation to design an optimized pharmacodynamic dosing strategy for meropenem. Journal of clinical pharmacology **2003**; 43(10): 1116-23.
- 15. Bulik CC, Nicolau DP. In vivo efficacy of simulated human dosing regimens of prolonged-infusion doripenem against carbapenemase- producing Klebsiella pneumoniae. Antimicrobial agents and chemotherapy **2010**; 54(10): 4112-5.

- Li C, Kuti JL, Nightingale CH, Nicolau DP. Population pharmacokinetic analysis and dosing regimen optimization of meropenem in adult patients. Journal of clinical pharmacology 2006; 46(10): 1171-8.
- 17. Daikos GL, Markogiannakis A. Carbapenemase-producing Klebsiella pneumoniae: (when) might we still consider treating with carbapenems? Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases **2011**; 17(8): 1135-41.
- Qureshi ZA, Paterson DL, Potoski BA, et al. Treatment outcome of bacteremia due to KPCproducing Klebsiella pneumoniae: superiority of combination antimicrobial regimens. Antimicrobial agents and chemotherapy **2012**; 56(4): 2108-13.
- 19. Tumbarello M, Viale P, Viscoli C, et al. Predictors of mortality in bloodstream infections caused by Klebsiella pneumoniae carbapenemase-producing K. pneumoniae: importance of combination therapy. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2012**; 55(7): 943-50.
- 20. Daikos GL, Tsaousi S, Tzouvelekis LS, et al. Carbapenemase-producing Klebsiella pneumoniae bloodstream infections: lowering mortality by antibiotic combination schemes and the role of carbapenems. Antimicrobial agents and chemotherapy **2014**; 58(4): 2322-8.
- 21. Sbrana F, Malacarne P, Viaggi B, et al. Carbapenem-sparing antibiotic regimens for infections caused by Klebsiella pneumoniae carbapenemase-producing K. pneumoniae in intensive care unit. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2013**; 56(5): 697-700.
- 22. Giamarellou H, Galani L, Baziaka F, Karaiskos I. Effectiveness of a double-carbapenem regimen for infections in humans due to carbapenemase-producing pandrug-resistant Klebsiella pneumoniae. Antimicrobial agents and chemotherapy **2013**; 57(5): 2388-90.
- 23. Ceccarelli G, Falcone M, Giordano A, et al. Successful ertapenem-doripenem combination treatment of bacteremic ventilator-associated pneumonia due to colistin-resistant KPC-producing Klebsiella pneumoniae. Antimicrobial agents and chemotherapy **2013**; 57(6): 2900-1.
- 24. Bulik CC, Nicolau DP. Double-carbapenem therapy for carbapenemase-producing Klebsiella pneumoniae. Antimicrobial agents and chemotherapy **2011**; 55(6): 3002-4.
- 25. Wiskirchen DE, Crandon JL, Nicolau DP. Impact of various conditions on the efficacy of dual carbapenem therapy against KPC-producing Klebsiella pneumoniae. International journal of antimicrobial agents **2013**; 41(6): 582-5.
- 26. Zavascki AP, Bulitta JB, Landersdorfer CB. Combination therapy for carbapenem-resistant Gramnegative bacteria. Expert review of anti-infective therapy **2013**; 11(12): 1333-53.
- 27. Gales AC, Jones RN, Sader HS. Contemporary activity of colistin and polymyxin B against a worldwide collection of Gram-negative pathogens: results from the SENTRY Antimicrobial Surveillance Program (2006-09). The Journal of antimicrobial chemotherapy **2011**; 66(9): 2070-4.
- Garonzik SM, Li J, Thamlikitkul V, et al. Population pharmacokinetics of colistin methanesulfonate and formed colistin in critically ill patients from a multicenter study provide dosing suggestions for various categories of patients. Antimicrobial agents and chemotherapy 2011; 55(7): 3284-94.
- 29. Sandri AM, Landersdorfer CB, Jacob J, et al. Population pharmacokinetics of intravenous polymyxin B in critically ill patients: implications for selection of dosage regimens. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2013**; 57(4): 524-31.
- 30. Bergen PJ, Landersdorfer CB, Zhang J, et al. Pharmacokinetics and pharmacodynamics of 'old' polymyxins: what is new? Diagnostic microbiology and infectious disease **2012**; 74(3): 213-23.

- 31. Petrosillo N, Giannella M, Lewis R, Viale P. Treatment of carbapenem-resistant Klebsiella pneumoniae: the state of the art. Expert review of anti-infective therapy **2013**; 11(2): 159-77.
- 32. Vicari G, Bauer SR, Neuner EA, Lam SW. Association between colistin dose and microbiologic outcomes in patients with multidrug-resistant gram-negative bacteremia. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2013**; 56(3): 398-404.
- 33. Falagas ME, Rafailidis PI, Ioannidou E, et al. Colistin therapy for microbiologically documented multidrug-resistant Gram-negative bacterial infections: a retrospective cohort study of 258 patients. International journal of antimicrobial agents **2010**; 35(2): 194-9.
- 34. Elias LS, Konzen D, Krebs JM, Zavascki AP. The impact of polymyxin B dosage on in-hospital mortality of patients treated with this antibiotic. The Journal of antimicrobial chemotherapy **2010**; 65(10): 2231-7.
- 35. Akajagbor DS, Wilson SL, Shere-Wolfe KD, Dakum P, Charurat ME, Gilliam BL. Higher incidence of acute kidney injury with intravenous colistimethate sodium compared with polymyxin B in critically ill patients at a tertiary care medical center. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2013**; 57(9): 1300-3.
- 36. Phe K, Lee Y, McDaneld PM, et al. In vitro assessment and multicenter cohort study of comparative nephrotoxicity rates associated with colistimethate versus polymyxin B therapy. Antimicrobial agents and chemotherapy **2014**; 58(5): 2740-6.
- 37. Pogue JM, Lee J, Marchaim D, et al. Incidence of and risk factors for colistin-associated nephrotoxicity in a large academic health system. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2011**; 53(9): 879-84.
- 38. Marchaim D, Chopra T, Pogue JM, et al. Outbreak of colistin-resistant, carbapenem-resistant Klebsiella pneumoniae in metropolitan Detroit, Michigan. Antimicrobial agents and chemotherapy **2011**; 55(2): 593-9.
- 39. Antoniadou A, Kontopidou F, Poulakou G, et al. Colistin-resistant isolates of Klebsiella pneumoniae emerging in intensive care unit patients: first report of a multiclonal cluster. The Journal of antimicrobial chemotherapy **2007**; 59(4): 786-90.
- 40. Lynch JP, 3rd, Clark NM, Zhanel GG. Evolution of antimicrobial resistance among Enterobacteriaceae (focus on extended spectrum beta-lactamases and carbapenemases). Expert opinion on pharmacotherapy **2013**; 14(2): 199-210.
- 41. Hirsch EB, Tam VH. Detection and treatment options for Klebsiella pneumoniae carbapenemases (KPCs): an emerging cause of multidrug-resistant infection. The Journal of antimicrobial chemotherapy **2010**; 65(6): 1119-25.
- 42. Balkan, II, Aygun G, Aydin S, et al. Blood stream infections due to OXA-48-like carbapenemaseproducing Enterobacteriaceae: treatment and survival. International journal of infectious diseases : IJID : official publication of the International Society for Infectious Diseases **2014**; 26C: 51-6.
- 43. Daikos GL, Markogiannakis A, Souli M, Tzouvelekis LS. Bloodstream infections caused by carbapenemase-producing Klebsiella pneumoniae: a clinical perspective. Expert review of anti-infective therapy **2012**; 10(12): 1393-404.
- 44. Sader HS, Farrell DJ, Flamm RK, Jones RN. Variation in potency and spectrum of tigecycline activity against bacterial strains from U.S. medical centers since its approval for clinical use (2006 to 2012). Antimicrobial agents and chemotherapy **2014**; 58(4): 2274-80.
- 45. Livermore DM, Warner M, Mushtaq S, Doumith M, Zhang J, Woodford N. What remains against carbapenem-resistant Enterobacteriaceae? Evaluation of chloramphenicol, ciprofloxacin, colistin, fosfomycin, minocycline, nitrofurantoin, temocillin and tigecycline. International journal of antimicrobial agents **2011**; 37(5): 415-9.

- 46. Souli M, Galani I, Antoniadou A, et al. An outbreak of infection due to beta-Lactamase Klebsiella pneumoniae Carbapenemase 2-producing K. pneumoniae in a Greek University Hospital: molecular characterization, epidemiology, and outcomes. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2010**; 50(3): 364-73.
- 47. Nguyen M, Eschenauer GA, Bryan M, et al. Carbapenem-resistant Klebsiella pneumoniae bacteremia: factors correlated with clinical and microbiologic outcomes. Diagnostic microbiology and infectious disease **2010**; 67(2): 180-4.
- 48. Peleg AY, Potoski BA, Rea R, et al. Acinetobacter baumannii bloodstream infection while receiving tigecycline: a cautionary report. The Journal of antimicrobial chemotherapy **2007**; 59(1): 128-31.
- 49. Cunha BA. Pharmacokinetic considerations regarding tigecycline for multidrug-resistant (MDR) Klebsiella pneumoniae or MDR Acinetobacter baumannii urosepsis. Journal of clinical microbiology **2009**; 47(5): 1613.
- 50. De Pascale G, Montini L, Pennisi MA, et al. High dose tigecycline in critically ill patients with severe infections due to multidrug-resistant bacteria. Critical care **2014**; 18(3): R90.
- 51. Di Carlo P, Gulotta G, Casuccio A, et al. KPC 3 Klebsiella pneumoniae ST258 clone infection in postoperative abdominal surgery patients in an intensive care setting: analysis of a case series of 30 patients. BMC anesthesiology **2013**; 13(1): 13.
- 52. Tuon FF, Rocha JL, Formighieri MS, et al. Fosfomycin susceptibility of isolates with blaKPC-2 from Brazil. The Journal of infection **2013**; 67(3): 247-9.
- 53. Perry JD, Naqvi SH, Mirza IA, et al. Prevalence of faecal carriage of Enterobacteriaceae with NDM-1 carbapenemase at military hospitals in Pakistan, and evaluation of two chromogenic media. The Journal of antimicrobial chemotherapy **2011**; 66(10): 2288-94.
- 54. Popovic M, Steinort D, Pillai S, Joukhadar C. Fosfomycin: an old, new friend? European journal of clinical microbiology & infectious diseases : official publication of the European Society of Clinical Microbiology **2010**; 29(2): 127-42.
- 55. Kitchel B, Sundin DR, Patel JB. Regional dissemination of KPC-producing Klebsiella pneumoniae. Antimicrobial agents and chemotherapy **2009**; 53(10): 4511-3.
- 56. Peirano G, Ahmed-Bentley J, Woodford N, Pitout JD. New Delhi metallo-beta-lactamase from traveler returning to Canada. Emerging infectious diseases **2011**; 17(2): 242-4.
- 57. Navarro-San Francisco C, Mora-Rillo M, Romero-Gomez MP, et al. Bacteraemia due to OXA-48carbapenemase-producing Enterobacteriaceae: a major clinical challenge. Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases **2013**; 19(2): E72-9.
- 58. Michalopoulos A, Virtzili S, Rafailidis P, Chalevelakis G, Damala M, Falagas ME. Intravenous fosfomycin for the treatment of nosocomial infections caused by carbapenem-resistant Klebsiella pneumoniae in critically ill patients: a prospective evaluation. Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases **2010**; 16(2): 184-6.
- 59. Karageorgopoulos DE, Miriagou V, Tzouvelekis LS, Spyridopoulou K, Daikos GL. Emergence of resistance to fosfomycin used as adjunct therapy in KPC Klebsiella pneumoniae bacteraemia: report of three cases. The Journal of antimicrobial chemotherapy **2012**; 67(11): 2777-9.
- Falagas ME, Kastoris AC, Kapaskelis AM, Karageorgopoulos DE. Fosfomycin for the treatment of multidrug-resistant, including extended-spectrum beta-lactamase producing,
 Enterobacteriaceae infections: a systematic review. The Lancet infectious diseases 2010; 10(1): 43-50.

- 61. Livermore DM, Mushtaq S, Warner M, et al. Activity of aminoglycosides, including ACHN-490, against carbapenem-resistant Enterobacteriaceae isolates. The Journal of antimicrobial chemotherapy **2011**; 66(1): 48-53.
- 62. Castanheira M, Sader HS, Jones RN. Antimicrobial susceptibility patterns of KPC-producing or CTX-M-producing Enterobacteriaceae. Microbial drug resistance **2010**; 16(1): 61-5.
- 63. Satlin MJ, Kubin CJ, Blumenthal JS, et al. Comparative effectiveness of aminoglycosides, polymyxin B, and tigecycline for clearance of carbapenem-resistant Klebsiella pneumoniae from urine. Antimicrobial agents and chemotherapy **2011**; 55(12): 5893-9.
- 64. Benenson S, Navon-Venezia S, Carmeli Y, et al. Carbapenem-resistant Klebsiella pneumoniae endocarditis in a young adult. Successful treatment with gentamicin and colistin. International journal of infectious diseases : IJID : official publication of the International Society for Infectious Diseases **2009**; 13(5): e295-8.
- 65. Bratu S, Brooks S, Burney S, et al. Detection and spread of Escherichia coli possessing the plasmid-borne carbapenemase KPC-2 in Brooklyn, New York. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America **2007**; 44(7): 972-5.
- 66. Lee GC, Burgess DS. Treatment of Klebsiella pneumoniae carbapenemase (KPC) infections: a review of published case series and case reports. Annals of clinical microbiology and antimicrobials **2012**; 11: 32.
- 67. Zarkotou O, Pournaras S, Tselioti P, et al. Predictors of mortality in patients with bloodstream infections caused by KPC-producing Klebsiella pneumoniae and impact of appropriate antimicrobial treatment. Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases **2011**; 17(12): 1798-803.
- 68. Jernigan MG, Press EG, Nguyen MH, Clancy CJ, Shields RK. The combination of doripenem and colistin is bactericidal and synergistic against colistin-resistant, carbapenemase-producing Klebsiella pneumoniae. Antimicrobial agents and chemotherapy **2012**; 56(6): 3395-8.
- 69. Falagas ME, Lourida P, Poulikakos P, Rafailidis PI, Tansarli GS. Antibiotic treatment of infections due to carbapenem-resistant enterobacteriaceae: systematic evaluation of the available evidence. Antimicrobial agents and chemotherapy **2014**; 58(2): 654-63.
- 70. Actavis plc. Actavis Receives U.S. FDA Approval for AVYCAZ[™] (CEFTAZIDIME-AVIBACTAM). 2015 Feb 25, Available at: <u>http://www.prnewswire.com/news-releases/actavis-receives-us-fda-approval-for-avycaz-ceftazidime-avibactam-300041592.html</u>. Accessed 2015 March 3.
- 71. Castanheira M, Farrell SE, Krause KM, Jones RN, Sader HS. Contemporary diversity of betalactamases among Enterobacteriaceae in the nine U.S. census regions and ceftazidimeavibactam activity tested against isolates producing the most prevalent beta-lactamase groups. Antimicrobial agents and chemotherapy **2014**; 58(2): 833-8.
- 72. Aktas Z, Kayacan C, Oncul O. In vitro activity of avibactam (NXL104) in combination with betalactams against Gram-negative bacteria, including OXA-48 beta-lactamase-producing Klebsiella pneumoniae. International journal of antimicrobial agents **2012**; 39(1): 86-9.
- 73. Vazquez JA, Gonzalez Patzan LD, Stricklin D, et al. Efficacy and safety of ceftazidime-avibactam versus imipenem-cilastatin in the treatment of complicated urinary tract infections, including acute pyelonephritis, in hospitalized adults: results of a prospective, investigator-blinded, randomized study. Current medical research and opinion **2012**; 28(12): 1921-31.
- 74. Lucasti C, Popescu I, Ramesh MK, Lipka J, Sable C. Comparative study of the efficacy and safety of ceftazidime/avibactam plus metronidazole versus meropenem in the treatment of complicated intra-abdominal infections in hospitalized adults: results of a randomized, double-blind, Phase II trial. The Journal of antimicrobial chemotherapy **2013**; 68(5): 1183-92.