

11-23-2005

## Characterization of the Interactions Between the Active Site of a Protein Tyrosine Kinase and a Divalent Metal Activator

Xiaofeng Lin  
*University of Rhode Island*

Marina K. Ayrabetov  
*University of Rhode Island*

Gongqin Sun  
*University of Rhode Island, gsun@uri.edu*

Follow this and additional works at: [https://digitalcommons.uri.edu/cmb\\_facpubs](https://digitalcommons.uri.edu/cmb_facpubs)

---

### Citation/Publisher Attribution

Lin X, Ayrabetov M, and Sun G. Characterization of the interactions between the active site of a protein tyrosine kinase and a divalent metal activator. *BMC Biochem*, 6, 25, 2005.  
Available at: <http://dx.doi.org/10.1186/1471-2091-6-25>

This Article is brought to you by the University of Rhode Island. It has been accepted for inclusion in Cell and Molecular Biology Faculty Publications by an authorized administrator of DigitalCommons@URI. For more information, please contact [digitalcommons-group@uri.edu](mailto:digitalcommons-group@uri.edu). For permission to reuse copyrighted content, contact the author directly.

---

## Characterization of the Interactions Between the Active Site of a Protein Tyrosine Kinase and a Divalent Metal Activator

Creative Commons License



This work is licensed under a [Creative Commons Attribution 2.0 License](https://creativecommons.org/licenses/by/2.0/).

Research article

Open Access

## Characterization of the interactions between the active site of a protein tyrosine kinase and a divalent metal activator

Xiaofeng Lin, Marina K Ayrapetov and Gongqin Sun\*

Address: Department of Cell and Molecular Biology, University of Rhode Island, Kingston, RI 02881, USA

Email: Xiaofeng Lin - [xlin8985@postoffice.uri.edu](mailto:xlin8985@postoffice.uri.edu); Marina K Ayrapetov - [mayr1229@gmail.com](mailto:mayr1229@gmail.com); Gongqin Sun\* - [gsun@uri.edu](mailto:gsun@uri.edu)

\* Corresponding author

Published: 23 November 2005

Received: 31 August 2005

*BMC Biochemistry* 2005, **6**:25 doi:10.1186/1471-2091-6-25

Accepted: 23 November 2005

This article is available from: <http://www.biomedcentral.com/1471-2091/6/25>

© 2005 Lin and Sun; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background:** Protein tyrosine kinases are important enzymes for cell signalling and key targets for anticancer drug discovery. The catalytic mechanisms of protein tyrosine kinase-catalysed phosphorylation are not fully understood. Protein tyrosine kinase Csk requires two  $Mg^{2+}$  cations for activity: one (M1) binds to ATP, and the other (M2) acts as an essential activator.

**Results:** Experiments in this communication characterize the interaction between M2 and Csk. Csk activity is sensitive to pH in the range of 6 to 7. Kinetic characterization indicates that the sensitivity is not due to altered substrate binding, but caused by the sensitivity of M2 binding to pH. Several residues in the active site with potential of binding M2 are mutated and the effect on metal activation studied. An active mutant of Asn319 is generated, and this mutation does not alter the metal binding characteristics. Mutations of Glu236 or Asp332 abolish the kinase activity, precluding a positive or negative conclusion on their role in M2 coordination. Finally, the ability of divalent metal cations to activate Csk correlates to a combination of ionic radius and the coordination number.

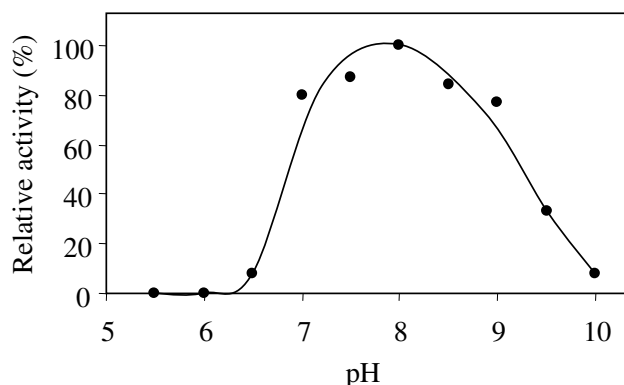
**Conclusion:** These studies demonstrate that M2 binding to Csk is sensitive to pH, which is mainly responsible for Csk activity change in the acidic arm of the pH response curve. They also demonstrate critical differences in the metal activator coordination sphere in protein tyrosine kinase Csk and a protein Ser/Thr kinase, the cAMP-dependent protein kinase. They shed light on the physical interactions between a protein tyrosine kinase and a divalent metal activator.

### Background

Protein tyrosine kinases (PTK)<sup>1</sup> are a large family of enzymes that transfer the  $\gamma$ -phosphate of ATP to tyrosine hydroxyl groups in proteins. By phosphorylation, PTKs regulate the conformation and function of their protein substrates [1]. This covalent modification is a fundamental mechanism of signal transduction in mammalian cells. Aberrant activation of many specific protein tyrosine kinases causes mishaps in cell signalling, and results in proliferative diseases, such as cancer [2]. Many protein

tyrosine kinases are considered as important targets for drug development against such diseases [3]. For full understanding of phosphorylation-mediated signalling and to provide a knowledge base for anti-PTK drug discovery, it is important to understand the catalytic mechanisms of protein tyrosine kinases.

C-terminal Src kinase (Csk) is a cytoplasmic PTK that phosphorylates Src family kinases (SFKs) and down-regulates their kinase activities [4,5]. The mechanistic basis of



**Figure 1**  
**pH optimum of Csk.** The Csk kinase activity is assayed as described in the Methods section. The activity at different pH is normalized to that at pH 8.

catalysis by Csk and PTKs in general is still poorly understood. Csk-catalyzed phosphorylation reaction obeys a ternary complex mechanism, likely with rapid and random binding of ATP-Mg and the phosphate-accepting substrate [6]. In addition to a  $Mg^{2+}$  cation (M1) as part of the ATP-Mg complex, Csk requires another  $Mg^{2+}$  ion (M2) for optimal kinase activity [7,8]. Kinetic studies demonstrate that M2 is an essential activator [7]. Because the affinity of Csk for the metal activator at 2.3 mM falls within the range of the cellular  $Mg^{2+}$  concentration, this activation may play a regulatory role in the kinase function [7,9].

Even though  $Mg^{2+}$  is likely the physiological activator, several other divalent metal cations can substitute for  $Mg^{2+}$  and activate Csk to various levels [8,10]. For example,  $Mn^{2+}$  can replace  $Mg^{2+}$  and results in higher activity of Csk, while  $Co^{2+}$ ,  $Ni^{2+}$  are not as effective as  $Mg^{2+}$  as an activator.  $Zn^{2+}$  can also substitute for  $Mg^{2+}$  in binding to the M2 binding site, but it cannot serve as an activator. Thus,  $Zn^{2+}$  acts as an inhibitor of Csk activity competitive against M2 [8]. Another intriguing property of the Csk-metal interaction is that these substitution metals all bind to Csk considerably stronger than the physiological activator,  $Mg^{2+}$ . While Csk binds to  $Mg^{2+}$  with an  $AC_{50}$  of 2.3 mM, the other metal cations all bind to Csk with  $AC_{50}$  or  $IC_{50}$  in the low  $\mu M$  range. Among all divalent metal cations tested,  $Zn^{2+}$  has the highest affinity for Csk, with an  $IC_{50}$  of 0.5  $\mu M$  [8].

The requirement of two divalent metal cations for full activity by Csk appears to represent a general catalytic requirement by all PTKs. Several PTKs from different families, such as v-Fps [11], Yes [12], Src [13], Lck [14], insulin receptor kinase [15] and epidermal growth factor

receptor [16], all require two  $Mg^{2+}$  cations for full activity. The insulin receptor kinase has been co-crystallized with both a peptide substrate and an ATP analog [17]. In the active site, two  $Mg^{2+}$  are observed, providing direct structural evidence for the presence of two  $Mg^{2+}$  ions in PTK catalysis. Kinetic analysis reveals that the metal cation activator might participate in catalysis by different mechanisms for different PTKs. For example, M2 activates Csk and Src by increasing the  $k_{cat}$  without affecting the  $K_m$  for ATP [7]. However, M2 activates IRK [18] and v-Fps [11] by decreasing the  $K_m$  for ATP without affecting the  $k_{cat}$ . The mechanistic basis for such kinetic differences has not been determined.

Interestingly, a protein Ser/Thr kinase, the cAMP-dependent protein kinase (PKA), also binds to two divalent metal cations in the active site during catalysis [19]. However, the second  $Mg^{2+}$  inhibits the kinase activity [20]. Crystallization of PKA complexed with catalytic ligands reveals that two  $Mg^{2+}$  cations are present in the active site [21].

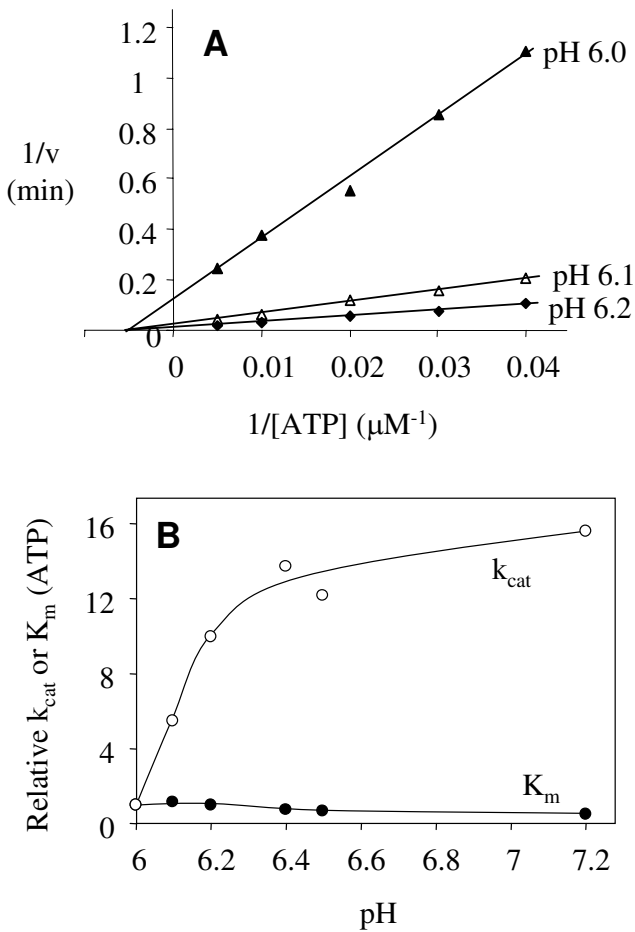
In the current study, we characterized the parameters for the interactions between Csk and M2, such as activity sensitivity to pH, required physical parameters of the divalent metal cation activators, and potential M2 coordinating residues. Mutagenic studies eliminated a residue as a potential ligand for M2, but could not determine if two other residues are involved due to lack of activity in all mutants varying these residues.

## Results

### **Csk activity is sensitive to pH in the range of 6 to 7**

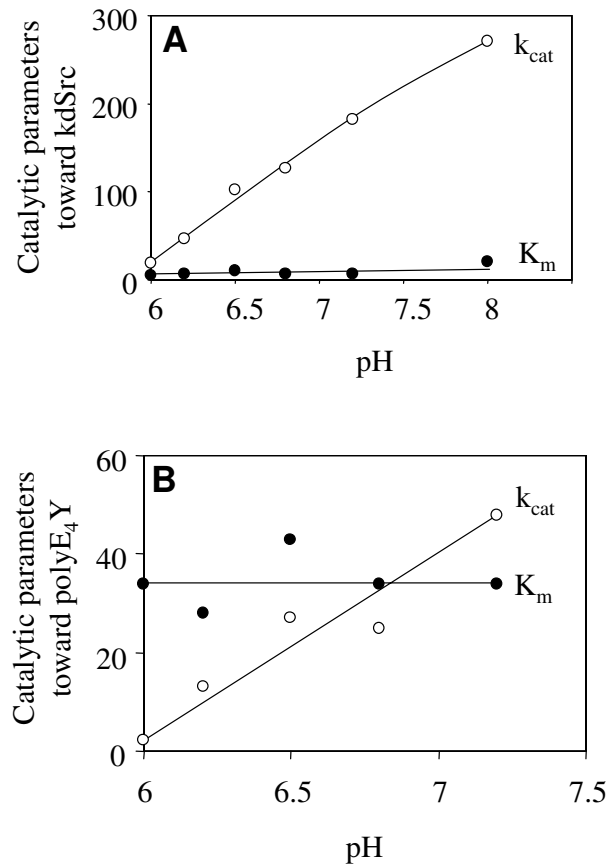
Because many PTKs are molecular targets for drug discovery, it is of high interest to understand the mechanisms of PTK catalysis. Like most protein tyrosine kinases, the catalytic mechanism of Csk is not fully understood. Csk has a bell-shaped pH response curve, with a pH optimum of around 8 (Figure 1). In the acidic arm of the pH curve, the activity is highly sensitive to pH. At pH 6, the enzyme shows very little activity, but at pH 7, the enzyme is nearly fully active. We hypothesize that this sensitivity likely reflects certain catalytically essential step(s) that is carried out by a functional group with an apparent  $pK_a$  in this range. Elucidation of such catalytically essential step(s) may shed light on the catalytic mechanism.

To test the above hypothesis, we characterized how steady state catalytic parameters responded to pH changes in this range (Figure 2). We first determined the catalytic parameters using ATP as the variable substrate. For this purpose, polyE<sub>4</sub>Y, a random polymer of Glu and Tyr (4:1), was used as the phosphate-accepting substrate at a fixed concentration of 1 mg ml<sup>-1</sup>. The apparent  $K_m$  for ATP decreased from 200  $\mu M$  at pH 6 to 91  $\mu M$  at pH 7.2 (relative  $K_m$  from 1 to 0.45 in Figure 2), but the  $k_{cat}$  increased



**Figure 2**  
**Effect of pH on Csk kinetics with ATP as the variable substrate.** (A) Double reciprocal plot of Csk activity using ATP as the variable substrate at pH 6, 6.1 and 6.2. (B) Plots of relative  $k_{cat}$  and  $K_m$  for ATP as a function of pH. The phosphate-accepting substrate was polyE<sub>4</sub>Y at 1 mg ml<sup>-1</sup>. The  $k_{cat}$  (9.1 min<sup>-1</sup>) and  $K_m$  (200  $\mu\text{M}$ ) at pH 6 are taken as 1 and all other values are relative to those.

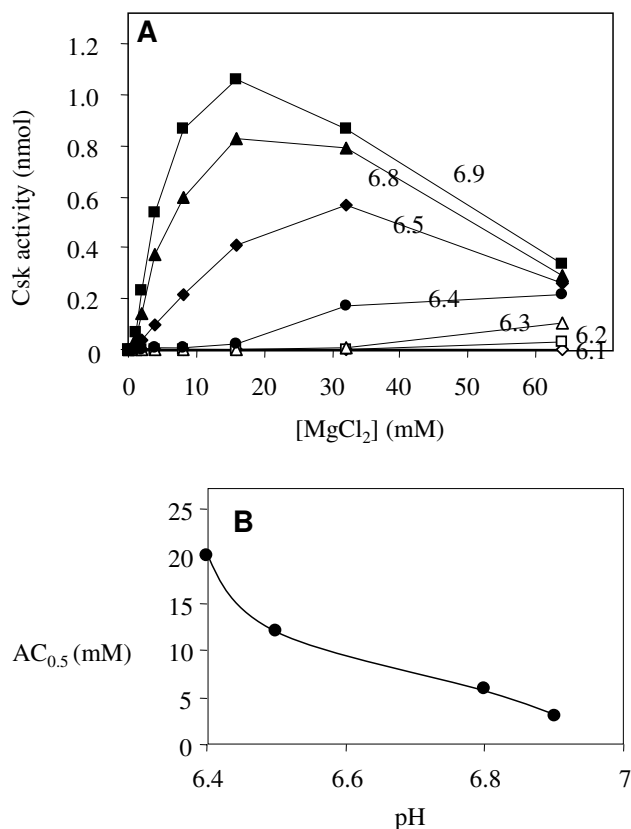
approximately 16 fold. The  $pK_a$  for this  $k_{cat}$  change is estimated to be 6.2. This indicates that phosphoryl transfer but not ATP binding to Csk is sensitive to pH. This pattern is different from those for PKA, whose apparent  $K_m$  for ATP gradually decreases by a factor of 7 when pH increases from 5.5 to 7 [22]. This contrast in kinetic patterns likely reflects differences in participation by pH-sensitive functional groups in catalysis by these two enzymes. In PKA, the sensitivity to pH is due to increased binding of ATP-M1 at a higher pH. Asp184 is a coordination ligand for M1 [21]. For Csk, the apparent  $K_m$  for ATP is also dependent on the presence of M1, with M1 resulting in



**Figure 3**  
**Effect of pH on Csk catalytic parameters using phosphate-accepting substrate as the variable substrate.** (A) KdSrc is used as the variable substrate. (B) PolyE<sub>4</sub>Y is used as the variable substrate.

lower  $K_m$  for ATP [7]. However, because the apparent  $K_m$  of Csk for ATP was largely independent of pH, the function of M1 is likely not affected by pH in this range. The contrast between Csk and PKA suggests that M1 coordination in Csk and PKA is different.

Steady state kinetics was then performed using the phosphate-accepting substrate as the variable substrate. The physiological substrates for Csk are the Src family kinases. Because SFKs are themselves PTKs that autophosphorylate, we used a kinase-defective mutant of Src (kdSrc) as the substrate. KdSrc contains a point mutation in the active site (Lys295Met), which abolishes Src kinase activity but does not affect its ability to serve as a Csk substrate [23,24]. The  $K_m$  of Csk for kdSrc did not change significantly in response to pH in the range of 6 to 7. However, the  $k_{cat}$  increased over 100-fold (Figure 3A). A similar kinetic pattern was observed for polyE<sub>4</sub>Y, a commonly



**Figure 4**  
**Mg<sup>2+</sup> activation of Csk as a function of pH.** (A) Mg<sup>2+</sup> activation of Csk is determined at different pH. The pH values are labeled in the graph. (B) Plot of AC<sub>50</sub> as a function of pH.

used artificial substrate for PTK activity assays (Figure 3B) [7,8]. This result indicates that the recognition of the phosphate-accepting substrate is not affected by pH changes in this range. Because the  $k_{cat}$  may be a function of both the phosphoryl transfer step and the ADP release [25], one of these steps is likely affected by pH in this range.

#### M2 binding to Csk is sensitive to pH

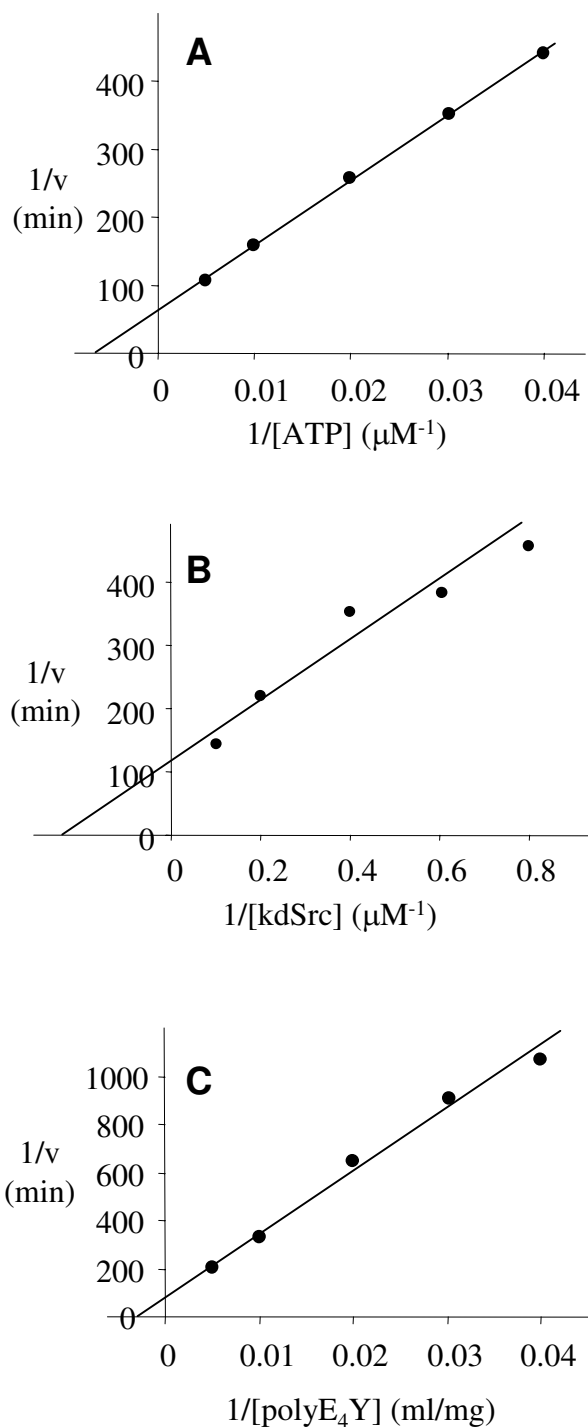
Because phosphoryl transfer, not substrate binding is sensitive to pH, it is likely that certain functional groups that catalyze phosphoryl transfer are subject to ionization in this pH range. Two types of functional groups could fall within this category. A functional group may directly participate in catalysis by interacting with the transition state, or it may interact with an essential activator. If only one ionization state of such a group is functional, it would render Csk phosphoryl transfer sensitive to pH. Because M2 acts as an essential activator, we determined if M2 binding to Csk was sensitive to pH in this range. The affin-

ity of Csk for M2 can be roughly measured by the AC<sub>50</sub>, the concentration of Mg<sup>2+</sup> that activates Csk to 50% of its full activity [7]. For example, at the optimal pH (8.0), Mg<sup>2+</sup> activates Csk with an AC<sub>50</sub> of 2.3 mM. We determined if the AC<sub>50</sub> of Csk for Mg<sup>2+</sup> activation was sensitive to pH. As shown in Figure 4A, the ability of Mg<sup>2+</sup> to activate Csk was indeed sensitive to pH. At pH 6.9, the AC<sub>50</sub> was 3 mM, close to the optimal AC<sub>50</sub>. As pH decreased, progressively higher concentration of Mg<sup>2+</sup> was required for Csk activation. At pH 6.4, 64 mM MgCl<sub>2</sub> nearly saturated Csk, and the AC<sub>50</sub> was estimated to be 20 mM. At pH 6.3 or below, highest activity was detected at 64 mM, making it difficult to estimate an AC<sub>50</sub>, but it is clear AC<sub>50</sub> continued to increase as pH decreased. It is known that Csk activity is highly sensitive to ionic inhibition [26], making it difficult to separate the effect of Mg<sup>2+</sup> as an activator from ionic inhibition at high MgCl<sub>2</sub> concentrations. Despite the lack of an accurate determination of the relationship between AC<sub>50</sub> and pH, it is clear that the AC<sub>50</sub> of Csk for Mg<sup>2+</sup> is dependent on pH. The pH dependence of metal binding is further illustrated by the pH dependence profile in Figure 4B. The pK<sub>a</sub> of this function was estimated in the range of 6.2 to 6.5, which correlated to the pH range where Csk activity was most sensitive to pH. This result strongly suggests that the sensitivity of phosphoryl transfer to pH is at least partly due to sensitivity of M2 binding to pH. Because the activity is gained with increased pH, the deprotonated form of the functional group is responsible for M2 binding.

#### Characterization of potential metal-coordinating residues in the active site of Csk

Only one protein tyrosine kinase, the insulin receptor kinase, has been co-crystallized with substrate analogs and divalent metal activators [17]. We compared the structures of Csk and IRK to identify Csk residues potentially involved in M2 coordination. In IRK, three residues are involved in metal cation coordination, Asp1150, Glu1047, and Asn1137. All three residues are conserved among PTKs, corresponding to Asp332, Glu236 and Asn319 in Csk [27]. Even though Csk and IRK displayed some differences in the kinetic patterns of Mg<sup>2+</sup> activation, it is likely that the conserved residues are playing similar roles in Mg<sup>2+</sup> coordination. We performed site-specific mutagenesis on these residues to determine if they are involved in metal activator coordination in Csk.

Asn319 is located in the catalytic loop and a universally conserved residue in all protein kinases, including Ser/Thr kinases. In IRK, the equivalent residue is involved in coordinating M1. Asn319 was mutated to Asp, His, Gln and Ser. Only one mutant, Asn319Ser, could be expressed as an active enzyme. Representative double reciprocal plots determining catalytic parameters of Asn319Ser mutant are presented in Figure 5 and summarized in Table 1. Overall,



**Figure 5**  
**Kinetic analysis of Asn319Ser.** Double reciprocal plots of Asn319Ser, using ATP (A), kdSrc (B) or polyE<sub>4</sub>Y (C) as the variable substrate. For the determination in (A), polyE<sub>4</sub>Y is used as the phosphate-accepting substrate.

the catalytic efficiency measured by  $k_{\text{cat}}$  decreased by a factor of approximately 10,000, while the apparent  $K_m$  values for ATP, polyE<sub>4</sub>Y and kdSrc did not change significantly (within a factor of 2). The Mg<sup>2+</sup> activation profile of this mutant was nearly identical to that of wt Csk (Figure 6) indicating that even though this residue is crucial for Csk catalysis, it is unlikely to be responsible for the M2 coordination. Because of the large decrease in  $k_{\text{cat}}$  due to the Asn319 mutation, there is a possibility that the mutation may have changed the rate-limiting step in Csk catalysis, making a direct conclusion about the role of Asn319 in M2 binding more complicated. The identical Mg<sup>2+</sup> responses by Asn319Ser and wild type Csk argue against this possibility.

Mutation of Glu236 and Asp332 to a number of residues, Ala, Asp, Gln for Glu236, Ala, Asn, Glu for Asp332, produced inactive mutants, thus kinetic analysis of their role in M2 binding is precluded. These two residues remain likely candidates for coordinating M2, but confirmation awaits further study by other methods.

#### **Divalent Metal cations of certain size bind to and activate Csk**

To characterize the physical properties required of the metal activator, it has been previously determined that Mn<sup>2+</sup>, Co<sup>2+</sup> and Ni<sup>2+</sup> could also serve as activators, while 14 other commercially available divalent metal cations could not [8,10]. To determine if the ability to activate Csk correlates to any specific physical attributes, we plotted the coordination number versus ionic radius [28] and determined where the activating metal cations were located on this map (Figure 7). The four divalent metal cations, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Co<sup>2+</sup> and Ni<sup>2+</sup>, that support Csk activity clustered together with a coordination number of 6 and an ionic radius of 0.65 to 0.8 Angstroms. Two other metal cations also met these criteria but apparently did not support the kinase activity: Cr<sup>2+</sup> and Fe<sup>2+</sup>. Both of these metal ions have multiple valencies, and interfere with the kinase assay. Fe<sup>2+</sup> forms brown precipitates likely with the enzyme and protein substrate, while Cr<sup>2+</sup> forms precipitates with ATP in the kinase assay. Such interferences prevent a definitive analysis if they could support Csk activity. Zn<sup>2+</sup> binds to Csk tightly but does not support Csk activity, thus it inhibits Csk activity as an inhibitor competitive against M2. Zn<sup>2+</sup> also falls within the range of the ionic radius, but has a coordination number of 4 or 5. This suggests that a coordination number of 6 may be required of the metal cation activator. Although this analysis suggests that the size and coordination number are likely important factors in determining if a metal cation could activate Csk catalysis, other factors may also be important.

**Table 1: Catalytic parameters of Csk and Asn319Ser mutant**

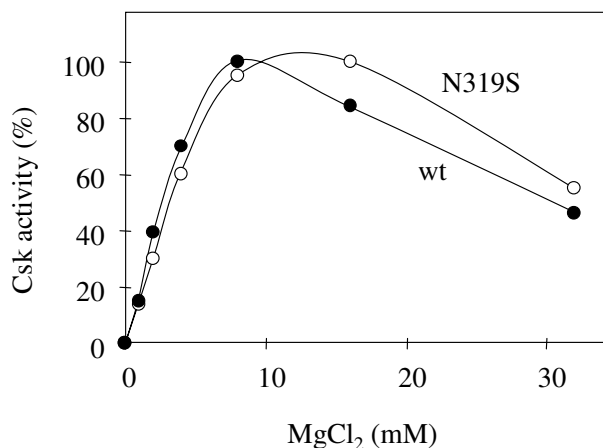
Parameter <sup>a</sup>	Wt Csk	Asn319Ser
$k_{cat}$ -ATP ( $\text{min}^{-1}$ )	160 ± 10	0.01 ± 0.007
$K_m$ -ATP ( $\mu\text{M}$ )	140 ± 12	150 ± 25
$k_{cat}$ -polyE <sub>4</sub> Y ( $\text{min}^{-1}$ )	82 ± 12	0.01 ± 0.001
$K_m$ -polyE <sub>4</sub> Y ( $\mu\text{g ml}^{-1}$ )	156 ± 30	220 ± 48
$k_{cat}$ -kdSrc ( $\text{min}^{-1}$ )	109 ± 3	0.01 ± 0.001
$K_m$ -kdSrc ( $\mu\text{M}$ )	6.4 ± 0.1	3.4 ± 0.1

<sup>a</sup>All assays were performed at least three times. The standard errors were calculated from three most consistent assays.

## Discussion

In this communication, we investigated the molecular basis of a commonly observed catalytic property of Csk. First, Csk activity is sensitive to pH change in the range of 6 to 7. Steady state kinetics demonstrates that the sensitivity is not due to the binding of Csk to either ATP-Mg or the protein substrate. The sensitivity is due to the sensitivity of M2 binding to pH in this range. Second, several residues that have the potential for M2 binding were studied by mutagenesis. These studies eliminated Asn319 in the active site as a potential ligand for M2 binding, but were inconclusive about the role of Asp332 and Glu236, because mutants at these two positions were inactive. Third, commercially available divalent metal cations were surveyed for their ability to support Csk activity. A strong correlation between the ability of divalent metal cation to support Csk activity and its physical parameters (ionic radius and the coordination number) was identified. Divalent metal cations with a coordination number of 6 and an ionic radius of 0.65–0.8 Å were able to support the activity while ions outside of this range were not. Overall this investigation provided insights into the kinase-divalent metal interaction in the active site.

The sensitivity of kinase activity to pH has been previously investigated in the cAMP-dependent protein kinase [22]. Interestingly, the binding of ATP was sensitive to pH for PKA while the binding of the second Mg<sup>2+</sup> to PKA is not sensitive to pH. This pattern is the opposite of that observed for Csk, likely reflecting different coordination patterns for M1 and M2 in Csk and PKA. This is also consistent with the structural information on IRK and PKA. Although in both PKA and IRK, three conserved residues (Glu1047, Asn1137, and Asp1150 in IRK, Glu91, Asn171 and Asp184 in PKA) are involved in coordinating M1 and M2, the positions of M1 and M2 are switched in the two kinases. In IRK, M1 is coordinated with Asn1127, while M2 is coordinated with Asp1150 directly and Glu1047 through two water molecules. In PKA, M2 is coordinated with Asn171, while M1 is coordinated with Asp184. Because M1 binding to PKA is sensitive to pH, it is likely due to deprotonation of Asp184. In this case, M2 binding to IRK would likely be sensitive to pH. This pattern is



**Figure 6**  
**Activation of wt and Asn319Ser mutant of Csk by Mg<sup>2+</sup>.** The maximum activities for both wt and Asn319Ser are set as 100%.

observed in Csk. This suggests that Csk and IRK likely uses a similar M2 binding site. In this case, Glu236 and Asp332 would be expected to be key ligands for M2 coordination.

Our effort to pinpoint the residues for coordinating M2 in Csk by mutagenic and kinetic studies is not fully successful. We were able to eliminate Asn319 as responsible for binding to M2, but our results are inconclusive regarding Asp332 and Glu236 due to the inability to generate active mutants at these two positions. This highlights the limitation of mutagenic approach to study catalytically essential residues. Further studies by other tools are required to solve these issues.

## Methods

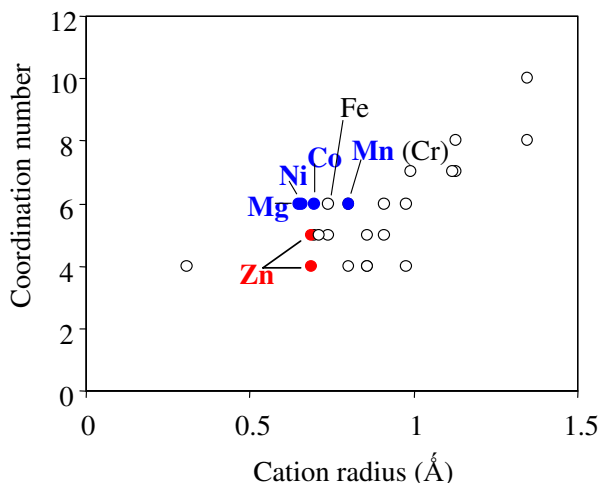
### Generation of Csk mutants

Glutathione S transferase (GST)-Csk fusion proteins were generated and purified as previously described [29]. Csk point mutants were generated using QuikChange (Stratagene) in the parental plasmid and were confirmed by DNA sequencing. Kinase-defective Src (kdSrc) was produced as described previously [23,24].

### Enzyme purification

Bacteria harbouring appropriate plasmids were cultured in LB medium at 37°C with shaking at 250 rpm overnight. The overnight culture was then mixed with an equal volume of fresh LB medium, cooled down to about 20°C. IPTG (0.2 mM) was added to the culture to induce recombinant protein expression at 20°C for 12 hours. The GST fusion proteins were purified by glutathione affinity chromatography as previously described [29]. The purified enzymes were desalted on a Sephadex G25 column equil-





**Figure 7**  
**Plot of coordination number versus cation radius for commercially available divalent metal cations.** Those cations that support Csk activity are shown in blue.  $Zn^{2+}$ , which also binds to Csk but does not support Csk activity, is shown in red.  $Fe^{2+}$  and  $Cr^{2+}$  are also labelled. Their ability to support Csk activity is discussed in the text. The coordination numbers and the cation radii are taken from reference [28] and the inhibition data is taken from reference [8].

ibrated with the storage buffer (100 mM Tris-Cl, pH 8.0, and 0.1%  $\beta$ -mercaptoethanol). Glycerol was added to the purified fractions to 30% and the enzymes were stored at  $-20^{\circ}C$ . Protein concentration was determined by the Bradford assay and the purity of purified proteins was assessed by SDS-PAGE with coomassie blue staining.

#### Kinase activity assay

For assaying PTK activity, phosphorylation of polyE<sub>4</sub>Y and kdSrc was measured using the acid precipitation assay as previously described [7]. Standard kinase assay buffer contains 100 mM EPPS, pH 8, 10% glycerol, 0.1% triton X-100 and 0.1%  $\beta$ -mercaptoethanol. Reaction time for the assays was 10 min. Standard assays used polyE<sub>4</sub>Y at 1 mg ml<sup>-1</sup>, or kdSrc at 10  $\mu$ M as the phosphate-accepting substrate and ATP at 0.2 mM as the phosphate-donating substrate. To determine the kinase activity at different pH, the kinase buffer contained all the standard buffer components except EPPS was replaced by 100 mM MES or Tris at designated pH. When  $K_m$  and  $k_{cat}$  were determined with regard to one substrate, the kinase activity was determined at various concentrations of that substrate in the range of 20 to 200  $\mu$ g ml<sup>-1</sup> for polyE<sub>4</sub>Y, 1 to 10  $\mu$ M for kdSrc or 20 to 200  $\mu$ M for ATP. When the phosphate-accepting substrate (either polyE<sub>4</sub>Y or kdSrc) was the variable substrate, ATP concentration was 0.2 mM. PolyE<sub>4</sub>Y at 1 mg ml<sup>-1</sup> was

used when ATP was the variable substrate. The  $k_{cat}$  and  $K_m$  values were determined by Lineweaver-Burk plots with linear regression using Microsoft Excel. All steady state kinetic assays were performed in duplicate, and repeated at least once. Standard errors were calculated if an assay was performed at least three times.

#### Abbreviation used

AC<sub>50</sub>, the concentration of a divalent metal cation that activates Csk to 50% of its full activity; Csk, C-terminal Src kinase; IRK, insulin receptor kinase; PKA, the camp-dependent protein kinase; PTK, protein tyrosine kinase(s); SFK, Src family kinase(s).

#### Authors' contributions

XL designed and performed the experiments and analyzed the results. MKA designed and performed experiments. GS designed the experiments, analyzed the results and wrote the paper.

#### Acknowledgements

This work was supported by grants from NIH (1R01CA111687 and 1P20RR16457), and the American Cancer Society (RSG-04-247-01-CDD). DNA sequencing was performed at URI Genomics and Sequencing Center.

#G. Sun is an American Cancer Society Research Scholar.

#### References

- Hubbard SR, Till JH: Protein tyrosine kinase structure and function. *Annu Rev Biochem* 2000, **69**:373-398.
- Hunter T: **The Croonian Lecture 1997. The phosphorylation of proteins on tyrosine: its role in cell growth and disease.** *Philos Trans R Soc Lond B Biol Sci* 1998, **353**:583-605.
- Krause DS, Van Etten RA: **Tyrosine kinases as targets for cancer therapy.** *N Engl J Med* 2005, **353**:172-187.
- Okada M, Nada S, Yamanashi Y, Yamamoto T, Nakagawa H: **CSK: a protein-tyrosine kinase involved in regulation of src family kinases.** *J Biol Chem* 1991, **266**:24249-24252.
- Sun G, Sharma AK, Budde RJ: **Autophosphorylation of Src and Yes blocks their inactivation by Csk phosphorylation.** *Oncogene* 1998, **17**:1587-1595.
- Cole PA, Burn P, Takacs B, Walsh CT: **Evaluation of the catalytic mechanism of recombinant human Csk (C-terminal Src kinase) using nucleotide analogs and viscosity effects.** *J Biol Chem* 1994, **269**:30880-30887.
- Sun G, Budde RJ: **Requirement for an additional divalent metal cation to activate protein tyrosine kinases.** *Biochemistry* 1997, **36**:2139-2146.
- Sun G, Budde RJ: **Substitution studies of the second divalent metal cation requirement of protein tyrosine kinase CSK.** *Biochemistry* 1999, **38**:5659-5665.
- Maguire ME: **Magnesium: a regulated and regulatory cation.** *Metal Ions Biol Syst* 1990, **26**:135-153.
- Grace MR, Walsh CT, Cole PA: **Divalent ion effects and insights into the catalytic mechanism of protein tyrosine kinase Csk.** *Biochemistry* 1997, **36**:1874-1881.
- Saylor P, Wang C, Hirai TJ, Adams JA: **A second magnesium ion is critical for ATP binding in the kinase domain of the oncoprotein v-Fps.** *Biochemistry* 1998, **37**:12624-12630.
- Sun G, Budde JA: **Expression, purification, and initial characterization of human Yes protein tyrosine kinase from a bacterial expression system.** *Arch Biochem Biophys* 1997, **345**:135-142.
- Budde JA, Ramdas L, Ke S: **Recombinant pp60c-src from baculovirus-infected insect cells: purification and characterization.** *Prep Biochem* 1993, **23**:493-515.

14. Wang QM, Srinivas PR, Harrison ML, Geahlen RL: **Partial purification and characterization of the Ick protein-tyrosine kinase from bovine thymus.** *Biochem J* 1991, **279**(Pt 2):567-574.
15. White MF, Haring HU, Kasuga M, Kahn CR: **Kinetic properties and sites of autophosphorylation of the partially purified insulin receptor from hepatoma cells.** *J Biol Chem* 1984, **259**:255-264.
16. Koland JG, Cerione RA: **Activation of the EGF receptor tyrosine kinase by divalent metal ions: comparison of holoreceptor and isolated kinase domain properties.** *Biochim Biophys Acta* 1990, **1052**:489-498.
17. Hubbard SR: **Crystal structure of the activated insulin receptor tyrosine kinase in complex with peptide substrate and ATP analog.** *EMBO J* 1997, **16**:5572-5581.
18. Vicario PP, Saperstein R, Bennun A: **Role of divalent metals in the kinetic mechanism of insulin receptor tyrosine kinase.** *Arch Biochem Biophys* 1988, **261**:336-345.
19. Mildvan AS, Rosevear PR, Fry DC, Bramson HN, Kaiser ET: **NMR studies of the mechanism of action and regulation of protein kinase.** *Curr Top Cell Regul* 1985, **27**:133-144.
20. Armstrong RN, Kondo H, Granot J, Kaiser ET, Mildvan AS: **Magnetic resonance and kinetic studies of the manganese(II) ion and substrate complexes of the catalytic subunit of adenosine 3',5'-monophosphate dependent protein kinase from bovine heart.** *Biochemistry* 1979, **18**:1230-1238.
21. Zheng J, Knighton DR, ten Eyck LF, Karlsson R, Xuong N, Taylor SS, Sowadski JM: **Crystal structure of the catalytic subunit of cAMP-dependent protein kinase complexed with MgATP and peptide inhibitor.** *Biochemistry* 1993, **32**:2154-2161.
22. Yoon MY, Cook PF: **Chemical mechanism of the adenosine cyclic 3',5'-monophosphate dependent protein kinase from pH studies.** *Biochemistry* 1987, **26**:4118-4125.
23. Wang D, Huang XY, Cole PA: **Molecular determinants for Csk-catalyzed tyrosine phosphorylation of the Src tail.** *Biochemistry* 2001, **40**:2004-2010.
24. Lee S, Lin X, Nam NH, Parang K, Sun G: **Determination of the substrate-docking site of protein tyrosine kinase C-terminal Src kinase.** *Proc Natl Acad Sci USA* 2003, **100**:14707-14712.
25. Shaffer J, Sun G, Adams JA: **Nucleotide release and associated conformational changes regulate function in the COOH-terminal Src kinase, Csk.** *Biochemistry* 2001, **40**:11149-11155.
26. Sondhi D, Xu W, Songyang Z, Eck MJ, Cole PA: **Peptide and protein phosphorylation by protein tyrosine kinase Csk: insights into specificity and mechanism.** *Biochemistry* 1998, **37**:165-172.
27. Ogawa A, Takayama Y, Sakai H, Chong KT, Takeuchi S, Nakagawa A, Nada S, Okada M, Tsukihara T: **Structure of the carboxyl-terminal Src kinase, Csk.** *J Biol Chem* 2002, **277**:14351-14354.
28. Glusker JP: **Structural aspects of metal liganding to functional groups in proteins.** *Adv Protein Chem* 1991, **42**:1-75.
29. Sun G, Budde RJ: **A modified pGEX expression system that eliminates degradation products and thrombin from the recombinant protein.** *Anal Biochem* 1995, **231**:458-460.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

